

AD _____

Award Number DAMD17-94-J-4205

TITLE: Cyclic AMP Modulation of Estrogen-Induced Effects: A Novel Mechanism for Hormonal Resistance in Breast Cancer

PRINCIPAL INVESTIGATOR: Benita S. Katzenellenbogen Ph.D.

CONTRACTING ORGANIZATION: University of Illinois
Champaign, Illinois 61820-6242

REPORT DATE: June 1999

TYPE OF REPORT: Final

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

20000306 096

DTIC QUALITY INSPECTED 3

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE June 1999	3. REPORT TYPE AND DATES COVERED Final (1 Oct 94 - 31 May 99)	
4. TITLE AND SUBTITLE Cyclic AMP Modulation of Estrogen-Induced Effects: A Novel Mechanism for Hormonal Resistance in Breast Cancer			5. FUNDING NUMBERS DAMD17-94-J-4205	
6. AUTHOR(S) Benita S. Katzenellenbogen Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) University of Illinois Champaign, Illinois 61820-6242			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited			12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200 words) <p>This research has been aimed at elucidating why breast cancer cells become resistant to antiestrogen treatment. Antiestrogens are used widely in the treatment of breast cancer, and although almost 50% of breast cancer patients benefit substantially from treatment with tamoxifen, many of these women eventually suffer relapse because some of the breast cancer cells become resistant to tamoxifen. We find that cyclic AMP stimulates the agonist activity of tamoxifen-like antiestrogens and reduces the ability of these antiestrogens to suppress estrogen-stimulated activity. In addition, estrogens and antiestrogens increase intracellular cAMP levels, which make tamoxifen more estrogenic and compromise its suppressive ability. We have developed several model breast cancer cell systems that differ in their sensitivity/resistance to antiestrogens and have characterized alterations in their proliferation, their production of and responsiveness to the transforming growth factors α and β, their intracellular cAMP levels, and their production of other estrogen-responsive proteins. We have used these breast cancer cells to study the involvement of intracellular cAMP in augmentation of antiestrogen agonist activity and in compromising the effectiveness of antiestrogens as estrogen antagonists, which can form a basis for tamoxifen resistance. These investigations highlight the important role of cAMP in modulation of estrogen and antiestrogen action in hormonal resistance, and suggest new directions for more effective strategies for the successful long-term treatment of hormone-responsive breast cancer.</p>				
14. SUBJECT TERMS Estrogen, Antiestrogen (Tamoxifen), Estrogen Receptor, Cyclic AMP, Hormone Resistance, Breast Cancer			15. NUMBER OF PAGES 206	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

FOREWORD

Opinions, interpretations, conclusions and recommendations are those of the author and are not necessarily endorsed by the U.S. Army.

____ Where copyrighted material is quoted, permission has been obtained to use such material.

____ Where material from documents designated for limited distribution is quoted, permission has been obtained to use the material.

____ Citations of commercial organizations and trade names in this report do not constitute an official Department of Army endorsement or approval of the products or services of these organizations.

NA In conducting research using animals, the investigator(s) adhered to the "Guide for the Care and Use of Laboratory Animals," prepared by the Committee on Care and use of Laboratory Animals of the Institute of Laboratory Resources, national Research Council (NIH Publication No. 86-23, Revised 1985).

NA For the protection of human subjects, the investigator(s) adhered to policies of applicable Federal Law 45 CFR 46.

NA In conducting research utilizing recombinant DNA technology, the investigator(s) adhered to current guidelines promulgated by the National Institutes of Health.

NA In the conduct of research utilizing recombinant DNA, the investigator(s) adhered to the NIH Guidelines for Research Involving Recombinant DNA Molecules.

____ In the conduct of research involving hazardous organisms, the investigator(s) adhered to the CDC-NIH Guide for Biosafety in Microbiological and Biomedical Laboratories.

Anta S. Katsellou
PI - Signature

June 20, 1999
Date

TABLE OF CONTENTS

	Page
FRONT COVER	1
SF 298 REPORT DOCUMENTATION PAGE	2
FOREWORD.....	3
TABLE OF CONTENTS.....	4
INTRODUCTION.....	5 - 6
BODY.....	6 - 15
KEY RESEARCH ACCOMPLISHMENTS.....	15 - 16
REPORTABLE OUTCOMES AND ALL PUBLICATIONS/ ABSTRACTS.....	16 - 21
CONCLUSIONS	21 - 22
PERSONNEL RECEIVING PAY FROM THE RESEARCH EFFORT	22
REFERENCES	22 - 25
APPENDIX Includes copies of publications	

(5) INTRODUCTION: Nature of the Problem, Background, Purpose and Scope of the Research

These studies are aimed at elucidating why breast cancer cells become resistant to antiestrogen treatment. Antiestrogens are used widely in the treatment of breast cancer, but development of resistance and patient relapse is a significant problem. The antiestrogen tamoxifen is the most widely prescribed drug for breast cancer treatment and it is usually considered the treatment of choice for the endocrine therapy of breast cancer because of its effectiveness, ease of use, and minimal side effects. It also appears to be of benefit in preventing the development of breast cancer in women at high risk for the disease, a very exciting recent finding in the major NCI-funded tamoxifen breast cancer prevention clinical trial (refs. 1-8; **see Reference list on pages 22-25**). Although almost one-half of breast cancer patients benefit substantially from treatment with tamoxifen, many of these women eventually suffer relapse because some of the breast cancer cells have become resistant to tamoxifen. This resistance to tamoxifen presents a major impediment to the long-term effectiveness of such treatments (refs. 9-17).

Our research is aimed at understanding and elucidating why breast cancer cells become resistant to antiestrogen treatment. In these studies we have used several model human breast cancer cell systems that differ in their sensitivity and resistance to tamoxifen, and we have investigated a novel mechanism and hypothesis that may explain antiestrogen resistance, namely the stimulation of adenylate cyclase by antiestrogens with increases in intracellular cAMP, augmentation of antiestrogen agonist character, and reduced effectiveness of antiestrogens as estrogen antagonists. Clinical experience has shown that hormonal resistance is often reversible, suggesting a cellular adaptation mechanism, rather than a genetic alteration in many breast cancers. This also seems to be the case in the tamoxifen-resistant human breast cancer cells (denoted MCF/TOT) we have developed (M. Herman and B. Katzenellenbogen, Publication #11; **see Publication list on pages 16 to 20**), and which are described in the section below entitled "Body". For example, patients that become resistant to tamoxifen often respond immediately to treatments with high dose estrogen or return to a state of tamoxifen responsiveness after a period of alternative therapy (refs. 18-20). Therefore, any mechanism that would explain tamoxifen resistance in these patients would have to involve mechanisms that would be reversible or adaptational, in contrast to other mechanisms for tamoxifen resistance that might involve mutations in the estrogen receptor or other critical transcription factor or growth factor genes. Therefore, we have been further investigating our observations regarding a two-way link between estrogen receptors and cAMP which would be consistent with a reversible and adaptational mechanism of antiestrogen resistance. Our observations that estrogens as well as antiestrogens are able to increase cAMP in breast cancer cells, and that cAMP increases the stimulatory effects of tamoxifen-like antiestrogens, could result in a feed-forward cascade that could result in the total compromising of the tumor growth suppressing activities of antiestrogens.

It is noteworthy that cAMP levels are significantly higher in breast tumors than in normal breast tissue (refs. 21, 22) and that elevated concentrations of cAMP binding proteins are associated with early disease recurrence and poor survival rates (refs. 23-25). Interestingly, as well, cAMP is both a mitogenic and a morphogenic factor in mammary cells (refs. 26-30) and it has been shown to enhance the mitogenic activity of several growth factors (as discussed in Fujimoto and Katzenellenbogen, Publication #1, and Herman and Katzenellenbogen, Publication #11 and references therein). Therefore, our overall goal in these studies has been to develop an understanding of the basis for the development of tamoxifen resistance in breast cancer. Understanding the basis for the development of tamoxifen resistance would be an important first step in developing more effective strategies for the successful long-term treatment of hormone-responsive breast cancer. In addition, this research should allow us to develop more effective therapies for antiestrogen-sensitive and antiestrogen-resistant breast cancers and should enable the use of antiestrogens to be approached most sensibly and effectively in the clinic.

(6) BODY: Experimental Methods Used, Results Obtained and the Relationship of Our Results to the Goals of the Research

Studies on cAMP and Estrogen Receptor Actions in Tamoxifen Responsive and Resistant Breast Cancer Cells (Statement of Work Task 1)

We completed and published our studies showing that cAMP alters the agonist/antagonist balance of tamoxifen-like antiestrogens and documented that this occurs in a promoter specific fashion (Publication #1, Fujimoto and Katzenellenbogen). In these studies, we have examined the effects of cAMP on the transcriptional activity of antiestrogens and estrogen (estradiol, E₂) in MCF-7 hormone responsive breast cancer cells, using several estrogen-responsive gene constructs. We have shown that increasing intracellular cAMP concentrations by treatment of MCF-7 human breast cancer cells with isobutyl methylxanthine (IBMX) plus cholera toxin or by enhancing intracellular protein kinase A activity by transfection of PKA catalytic subunits, stimulates the agonistic activity of the antiestrogens, tamoxifen, LY117018 and nafoxidine (Publication #1, see Appendix Reprints) on several estrogen responsive genes. In contrast, increasing intracellular cAMP levels or protein kinase A activity does not increase transcriptional stimulation by the more pure steroidal antiestrogen ICI 164,384. In addition, increasing intracellular cAMP in the breast cancer cells markedly reduced the ability of the tamoxifen-like antiestrogens to suppress estrogen stimulated activity (Figs. 1, 2 and 4 of Publication #1). Therefore, cAMP increased the agonist activity of tamoxifen-like but not ICI 164,384-like antiestrogens and also reduced the estrogen suppressing activity of the tamoxifen-like antiestrogens. Our observations (Publication #1 and also Publication #12) indicate that stimulation of the protein kinase A signal transduction pathway can change the interpretation of the character of the antiestrogen ligand, changing it from an estrogen antagonist to a partial, but significant, agonist. This stimulation of the protein kinase A signaling

phosphodiesterase (PDE) activity (Publication #2). The effect of estrogen and antiestrogens on adenylate cyclase and PDE activity was assessed in cell membranes isolated from MCF-7 cells after hormone treatment. The membrane fractions were combined with ^{32}P - α -ATP to monitor cAMP production; and harvested cells were homogenized and extracts monitored for PDE activity. Treatment of cells with CT (1 $\mu\text{g}/\text{ml}$) evoked a ca. three-fold increase in adenylate cyclase activity within 20 min, while treatment with E₂ or the antiestrogens TOT and ICI resulted in increases that were half as great. The effects of E₂, CT and ICI were not additive. By contrast, E₂, TOT or ICI did not alter PDE activity. These increases in cAMP result from enhanced membrane adenylate cyclase activity, by a mechanism that does not involve genomic actions of the hormones (i.e., is not blocked by inhibitors of RNA and protein synthesis). In addition, we find that estradiol and antiestrogens are able to stimulate adenylate cyclase in isolated membrane preparations from MCF-7 cells, indicating that these agents can act directly on the adenylate cyclase system in the membrane. These findings clearly imply a site of estrogen and antiestrogen action in the membrane.

Since we have shown that estrogens and antiestrogens increase cAMP within breast cancer cells, and cAMP alters the agonist/antagonist balance of antiestrogens, the increase in cAMP may result in a reduction in the tumor growth-suppressing activities of tamoxifen-like antiestrogens, a change that may underlie the development of tamoxifen resistance in some breast cancer patients. To examine this hypothesis in detail, we isolated and characterized estrogen growth-autonomous breast cancer cell sublines (Publication #3, Herman and Katzenellenbogen) and antiestrogen-resistant MCF-7 human breast cancer sublines that we have selected and cloned, and we have determined their responses to antiestrogens and cAMP in terms of cell proliferation and growth factor production, and the responses of other genes normally estrogen-regulated, such as pS2 and progesterone receptor (Publication #3, Herman and Katzenellenbogen; Publications #4 and #7, Nicholson et al; Publication #6, Ince et al.; and Publication #5, Katzenellenbogen et al.). These studies directly address the Statement of Work Task 1, points A, B, C and D.

In Publications #3, #4 and #7, we observed alterations in transforming growth factor α and β production and cell responsiveness during the progression of MCF-7 human breast cancer cells to estrogen-autonomous growth. We experimentally induced this progression from a steroid-regulated to a steroid-autonomous state, by long-term culture in the absence of steroids. These long-term steroid deprived cells, were initially slowed in their growth in response to steroid deprivation, but after several months, showed rapid, steroid-independent growth rates. Our findings documented a marked (ca. 80%) transient decrease in TGF α mRNA and protein production and a marked and transient increase (3-10-fold) increase in TGF β 1, β 2, and β 3 mRNA's and bioactive TGF- β proteins at two to 10 weeks of steroid deprivation which then returned by 24 weeks to the lower levels of the parental MCF-7 cells. Of note, these cells showed a loss of regulation of proliferation by TGF- α and a 10-fold decrease in sensitivity to the growth-suppressive effects of TGF β 1, despite little change

in receptor levels for these factors. The marked transient alterations in the levels of these growth factors indicate that they may play a role in the events which accompany the progression from estrogen-responsive to estrogen-autonomous growth in breast cancer cells.

To characterize the properties of tamoxifen-resistant breast cancer cells, we cultured MCF-7 breast cancer cells long-term (longer than 1 year) in the presence of the antiestrogen *trans*-hydroxy-tamoxifen (TOT) to generate a subline refractory to the growth-suppressive effects of TOT. This subline (designated MCF/TOT) showed growth stimulation, rather than inhibition, with TOT and diminished growth stimulation with estradiol (E_2), yet remained as sensitive as the parental cells to growth suppression by another antiestrogen, ICI 164,384 (Publication #11, Herman and Katzenellenbogen). Estrogen receptor (ER) levels were maintained at 40% that in parent MCF-7 cells, but MCF/TOT cells failed to show an increase in progesterone receptor content in response to E_2 or TOT treatment. In contrast, the MCF/TOT subline behaved like parental cells in terms of E_2 and TOT regulation of ER and pS2 expression and transactivation of a transiently transfected estrogen-responsive gene construct. DNA sequencing of the hormone binding domain of the ER from both MCF-7 and MCF/TOT cells confirmed the presence of wild-type ER and exon 5 and exon 7 deletion splice variants, but showed no point mutations. Compared to the parental cells, the MCF/TOT subline showed reduced sensitivity to the growth-suppressive effects of retinoic acid and complete resistance to exogenous TGF- β 1.

The altered growth responsiveness of MCF/TOT cells to TOT and TGF- β 1 was partially to fully reversible following TOT withdrawal for 16 weeks. Our findings underscore the fact that antiestrogen resistance is response-specific; that loss of growth suppression by TOT appears to be due to the acquisition of weak growth stimulation; and that resistance to TOT does not mean global resistance to other more pure antiestrogens such as ICI 164,384, implying that these antiestrogens must act by somewhat different mechanisms. The association of reduced retinoic acid responsiveness and insensitivity to exogenous TGF- β with antiestrogen growth-resistance in these cells supports the increasing evidence for interrelationships among cell regulatory pathways utilized by these three growth-suppressive agents in breast cancer cells. Since these MCF/TOT cells, resistant to the growth suppressive effects of antiestrogens or TGF- β continue to express TGF- β type I and II receptors of the correct size and in amounts equal to those observed in the parental cells, their lack of inhibition by the high levels of TGF- β 1 either being made by the cells or added by us to their culture media suggest a lesion after receptor binding, i.e. at some point in the TGF- β intracellular signaling pathway. In addition, our findings indicate that one mechanism of antiestrogen resistance, as seen in MCF/TOT cells, may involve alterations in growth factor and other hormonal pathways that affect the ER response pathway (Publications #8 and #11).

We have also used several MCF-7 cell clones with altered antiestrogen sensitivity to investigate the response to cAMP and antiestrogen as monitored by proliferation rates, colony formation ability and changes in regulation of several growth-related genes (TGF- β , TGF- α , pS2, and TGF- α /EGF receptor), (Publications #3, #4, #7, and #11). In addition, we have studied the regulation of the progesterone receptor in tamoxifen- and estrogen-sensitive and tamoxifen- and estrogen-resistant breast cancer cells, since the progesterone receptor is often used as an end-point or marker of hormone sensitivity and responsiveness. By monitoring progesterone receptor content in the cells, using several different progesterone receptor-specific antibodies, we have observed that the progesterone receptor B/A ratio is higher with trans-hydroxytamoxifen versus estrogen treatment of cells (a variety of different estrogens were tested) and progesterone receptors were further increased by treatment of cells with 8-Br-cAMP and trans-hydroxytamoxifen.

Factors Important in Regulation of cAMP Levels in Antiestrogen Responsive and Resistant Cells (Statement of Work Task 2)

We have monitored basal and stimulated levels of cAMP in parental MCF-7 cells and in our MCF/TOT (tamoxifen stimulated) MCF-7 cells and in estrogen receptor negative MDA-MB-231 breast cancer cells which are unresponsive to estrogen and antiestrogen. We have found that the antiestrogen-stimulated MCF-7 cells and the antiestrogen-unresponsive 231 cells showed 3-5 times higher intracellular cAMP levels than were observed in the parental MCF-7 cells. We observed no stimulation of cAMP levels by estrogen or antiestrogen treatment of 231 cells, while we observed only a 1.5-fold change in cAMP in the MCF/TOT cells and we observed a 3-4 fold increase in cAMP in the parental MCF-7 cells. Thus, hormone resistant and antiestrogen stimulated cells interestingly had elevated basal levels of cAMP, an observation we also made in breast cancer cells studied under Task 2, that were kindly provided by Dr. Fran Kern of the Lombardi Cancer Research Center at Georgetown University in Washington D. C.

Under Task 2, we have worked towards the identification of endogenous and exogenous agents and factors that result in elevation of cAMP levels in breast cancer cells. We have investigated the correlation between antiestrogen growth responsiveness/resistance and cellular cAMP levels and adenylate cyclase activities. Using 5 breast cancer cells lines (MCF-7 wild type versus MCF-7 tamoxifen stimulated, and 3 MCF-7 cell lines that are resistant to antiestrogen (MCF-7-v-Ha-ras, MCF-7-FGF1 and MCF-7-FGF4, which stably overexpress ras, FGF-1, or FGF-4, respectively, kindly provided to us by Dr. Fran Kern, we have observed that the overexpressing ras and FGF cells show basal cAMP levels 2.5-3.5 x higher than wild type MCF-7 cells. Values obtained were as follows (mean \pm S. D. : wild type MCF-7 cells, 35 \pm 10; MCF-7 ras, 121 \pm 9; MCF-7 FGF-1, 86 \pm 2; MCF-7 FGF-4, 103 \pm 9. Interestingly, these latter three cell types, which proliferate rapidly and do not have their rate of proliferation influenced by estrogen or antiestrogen, likewise did not have their intracellular cAMP levels

influenced by estrogen or antiestrogen treatment. Thus, elevated levels of cellular cAMP appear to correlate with altered growth responsiveness/resistance and with an estrogen and antiestrogen growth-autonomous state.

We also asked whether estradiol would affect intracellular cAMP in human endometrial cancer Ishikawa cells. These cells contain estrogen receptor and were of interest because tamoxifen is known to be quite agonistic (i.e. stimulatory) in endometrial cells, and in fact, a major concern in the Tamoxifen Prevention Trial in women has involved stimulation of the uterus by tamoxifen. We observed in these cells, basal and estrogen-stimulated and isobutyl methyl xanthine (IBMX)/cholera toxin-stimulated levels of cAMP similar in magnitude to those observed in the MCF-7 wild type breast cancer cells, namely an approximately 20-fold increase in response to IBMX and cholera toxin and an approximately 3-6 fold increase in response to estradiol. Thus, these uterine cells did not show a response to estrogen or to tamoxifen substantially different in magnitude from that observed with MCF-7 breast cancer cells.

Since antiestrogens such as tamoxifen can have partial estrogen-like activity in some cell types, and studies have implied that this stimulation is dependent on the amino-terminal activation function-1-containing region of the receptor, we studied this region of the receptor in detail (McInerney and Katzenellenbogen, Publication #13). In our investigations on the A/B domain of the estrogen receptor and its role in the transcriptional activity of the estrogen receptor elicited by estrogens and some antiestrogens, we have found that different regions within this domain are required for transcriptional stimulation by estrogen versus antiestrogen. We demonstrated that a specific 24-amino acid region of activation function-1 of the human estrogen receptor is necessary for agonism by trans-hydroxytamoxifen and other partial agonist/antagonist antiestrogens, but is not required for estradiol-dependent transactivation. As a consequence, the activity of estradiol and the estrogen agonist/antagonist character of trans-hydroxytamoxifen depended markedly, but not always concordantly, on the sequences present within the A/B domain in the receptor. Our studies show that hormone-dependent transcription utilizes a broad range of sequences within the amino terminal A/B domain and suggest that differences in the agonist/antagonist character of antiestrogens observed in cells could be due to altered levels of specific factors that interact with these regions of the receptor protein. In related work, we have demonstrated that a specific *cis* element from the promoter of an estrogen-responsive gene can alter the transcriptional activity of hormone and antihormone-occupied receptor bound at its response element near the promoter. Such ligand response modulatory elements, and changes in the levels and activity of factors that bind to such elements, may underlie the different sensitivities of steroid hormone-regulated genes to both hormones and antihormones (Publication #16).

During our work, a publication appeared in which a group of Italian researchers reported that sex steroid binding globulin (SSBG) was necessary in the stimulation of cAMP by estrogen in breast cancer cells (ref. 31). Because we felt it was essential for us

to determine if this was important in our work related to Task 1c and 1d and Task 3, we purchased SSBG from two different sources, namely Calbiochem and Scripps Laboratories, both SSBG preparations in highly purified form. We followed the Fissore protocol as closely as possible and also did several variations. Thus, we utilized 1nM and 3nM SSBG concentrations with cells in serum-free medium, and in 0.5% and 5% serum, and with cells in serum-free medium containing insulin, transferin and selenium. We also tested several different concentrations of estradiol, namely 10^{-8} , 10^{-9} and 10^{-10} M. In no case, did we observe a stimulatory (nor a suppressive) effect of SSBG on the cAMP response to hormone. Thus, despite several months of experiments, we were not able to confirm that sex steroid binding globulin was necessary for the stimulation of cAMP by hormone in our breast cancer cells. We therefore have ruled this out as a likely important factor in our studies in Tasks 1 and 3.

Efforts to Identify a Membrane Receptor for Estrogens (Statement of Work Task 3)

Our initial idea under Task 3 was to use estrogen radioligands, including the non-steroidal affinity labeling agent tamoxifen aziridine, to identify estrogen receptors in the membrane fraction of MCF-7 cells. These studies proved to be difficult, as we found it is technically difficult to perform quantitative binding studies on the cell membrane fraction; furthermore, tamoxifen aziridine failed to label any membrane protein covalently in a specific fashion (i.e., labeling that was significantly blocked by pretreatment with unlabeled estrogens).

The identification of interaction partners for proteins has been revolutionized by the yeast 2-hybrid screen. This interaction cloning method permits novel targets that interact with a bait protein (prepared as a fusion protein with a GAL4 DNA-binding domain; GAL4-DBD) to be identified in libraries of prey proteins (prepared as fusion proteins with a GAL4 activating domain; GAL4-AD). Interaction is scored by the activation of specific gene transcription that results when the two fusion proteins bind and generate a complex capable of activating transcription. This transcription can be tied to a colorimetric assay or a survival screen. As powerful as the yeast 2-hybrid screen is in identifying the interaction between two proteins, it is not useful, per se, in identifying the interaction between proteins and small molecules.

Jun Liu has reported a novel extension of the yeast 2-hybrid screen that can be used to identify the interaction between proteins and small molecules; he has named this extension the "yeast 3-hybrid screen" (ref. 32). In this extended version of the interaction cloning screen, the third hybrid or fusion component is a heterobivalent ligand that acts as a chemical adaptor. One end of this "chemical fusion" or hybrid species contains the small molecule bait; the other end contains a second small molecule that has distinctly different binding properties; this latter ligand acts as a tether by binding to a ligand binding domain that is fused to the GAL4-DBD. In Liu's version, the tethering ligand was a glucocorticoid ligand, which was bound by the ligand binding domain of the glucocorticoid receptor (GR-LBD) fused to the GAL4-DBD. In

his case, he used an immunosuppressant as bait and was able to identify immunophilin clones that were present in a library of prey proteins.

This 3-hybrid screen affords a new approach to search for novel estrogen receptors, some of which may be in the membrane, through their physical interaction with an estrogen ligand. This is a cloning method, but not one that depends on sequence homology or function, simply small molecule-receptor interaction. In order to undertake this 3-hybrid screen for novel estrogen receptors, we decided to synthesize a heterobivalent ligand in which an estrogen would be chemically tethered to a glucocorticoid. This chemical fusion would then be added to a screen in which a library of cellular cDNA, expressed as fusion proteins with a GAL4-AD, is presented to a GR-LBD/GAL4-DBD hybrid. We would isolate clones that showed activity only in the presence of the heterobivalent ligand. For this work, we completed the preparation of a cDNA library from MCF-7 human breast cancer cells fused to the GAL4-AD.

In work towards the development of the yeast 3-hybrid screen to identify novel estrogen binding proteins, that we proposed for work under Task 3, we synthesized various derivatives of estrogens and glucocorticoids. These were components to be joined together as the heterobivalent ligand component needed in the 3-hybrid screen. In the glucocorticoid series, we examined derivatives substituted at the most accessible C(21) and C(17 α) hydroxyl groups of cortisol and dexamethasone, and in the estrogens, we examined derivatives substituted at the 7 α , 16 α , and 17 α positions of estradiol. These are sites at which these ligands are generally considered to be most tolerant of long-chain substituents.

We were surprised by the degree to which binding affinity of each ligand was reduced by the attachment of the type of long chain substituent that would be needed to generate the heterobivalent ligand. Affinities of the parent ligand were reduced by a factor of 10-100. Even long-chain derivatives of other non-steroidal estrogens showed greatly reduced binding affinities. Because of this, we decided that it would not be worthwhile to prepare the heterobivalent ligands themselves and to undertake the yeast 3-hybrid screen. So this portion of the project was not pursued further.

Determination of the Mechanism by Which cAMP Alters the Agonist/Antagonist Activity of Antiestrogens: CREB-Estrogen Receptor Transcriptional Synergy on Estrogen Regulated Genes (Statement of Work Task 4)

We have performed these studies under Task 4. The estrogen receptor and cAMP signaling pathways appear to interact and one aspect of this interaction is that estradiol and antiestrogens like tamoxifen and protein kinase A activators can synergistically enhance transcription of both endogenous genes and reporter genes containing only estrogen response elements. Site-directed mutagenesis of potential protein kinase A phosphorylation sites on the estrogen receptor (S236 and S302) indicated that phosphorylation of these sites was not necessary for the transcriptional synergy. Transient transfection assays in two different cell backgrounds using three

different reporters containing either cAMP response elements, estrogen response elements or both types of elements in the presence and absence of cAMP response element binding protein (CREB) expression vector showed that CREB was involved in this synergistic interaction. The functional interaction of estrogen receptor and CREB on a reporter containing only an estrogen response element was also seen in a mammalian two-hybrid system. We therefore propose that in the transcriptional synergy between estrogen receptor and CREB, activated CREB can be recruited to DNA by a complex containing estradiol bound estrogen receptor, steroid receptor coactivator-1 and CREB binding protein. This larger complex containing both the estrogen receptor and CREB can be shown to synergistically enhance transcription of estrogen regulated genes and to alter the agonist/antagonist balance and activity of antiestrogens (Thomas, Lazennec and Katzenellenbogen, Publication #A13 and manuscript in preparation).

The Role of Estrogen Receptor Phosphorylation in Antiestrogen and cAMP Agonistic Activity (Statement of Work Task 4)

Under Task 4, we have also examined the role of specific phosphorylation sites in antiestrogen and cAMP agonistic activity. The estrogen receptor contains two potential cAMP-dependent protein kinase sites at serine 236 in the DNA binding domain and serine 302 at the very start of the hormone binding domain. We therefore have changed these serines to alanines by site-directed oligonucleotide mutagenesis of the estrogen receptor cDNA. The change from serine to alanine would thus eliminate the possibility of phosphorylation at these sites. We tested the response of these mutants to cAMP and estrogen and antiestrogen in order to identify sites of phosphorylation that may be associated with the alteration in tamoxifen agonist character in the presence of cAMP (Publication #A13 and manuscript in preparation). To our surprise, mutation of either of these sites, or both of these sites together, did not prevent nor reduce the synergism between cAMP and estrogen. Thus, these sites do not appear to be involved in mediating the enhanced transcriptional response when both cyclic AMP and antiestrogen are administered. Likewise, we examined the possible role of serine 118 since this is a MAP kinase site and there is now evidence for crosstalk and interrelationships between cAMP and MAP kinase pathways. The S118A estrogen receptor mutant was as effective as the wild type estrogen receptor in supporting tamoxifen agonism with cAMP present.

In addition to these three phosphorylation sites on the estrogen receptor, there are three additional calmodulin-dependent protein kinase II sites (S154, S167, S518), the first two sites being present in the activation function-1 region of the estrogen receptor, the region that we and others have shown to be responsible for tamoxifen agonism, and the latter amino acid in the hormone binding domain of the receptor. We therefore also examined if these phosphorylation sites could account for the synergy observed between ligand and PKA activators. Using site-directed mutagenesis, we changed these sites to alanine, thereby eliminating the possibility of their phosphorylation. We

observed that all of these serine mutants were activated by ligand in a very similar manner to that of the wild type estrogen receptor, and moreover, these mutants showed a synergistic activation of transcription when cells were treated with ligand and PKA activator, suggesting therefore that mutation of any one of these phosphorylation sites does not result in a loss of estrogen and protein kinase A transcriptional synergy. Thus, phosphorylation at these sites is not required for the transcriptional synergism elicited in the presence of cAMP.

Based on these findings, we also investigated the possible role of tyrosine 537 as a potentially important phosphorylation site (Publications #15 and #18). Intriguingly, changing this tyrosine to alanine, an amino acid not capable of being phosphorylated resulted in partial constitutive activity of the estrogen receptor, and changing tyrosine to serine resulted in full constitutive activity. Changing the tyrosine to several other amino acids had no effect on estrogen receptor activity. Our findings, that changing the tyrosine to another amino acid resulted in receptors fully responsive to estrogen and that several mutants show constitutive activity, indicate that response to estrogen does not require phosphorylation at this site but that the position of this tyrosine, near the start of helix 12 and the activation function-2 region of the receptor, can result in a receptor conformation in which the receptor is active even in the absence of hormone.

Based on our findings with phosphorylation site mutants, we also investigated the role of the steroid receptor coactivator SRC-1 in the activity of the estrogen receptor (Publications #14 and #19). Since we found that SRC-1 markedly enhances the activity of the estrogen receptor and the functional interaction between the N- and C- terminal regions of the receptor, it is possible that the transcriptional synergism between cAMP and the ligand occupied receptor may result both from changes in phosphorylation of the estrogen receptor itself as well as coregulators such as SRC-1 which are highly phosphorylated proteins. Such coregulators are now well-documented to be involved in enhancing the level of transcriptional activity of the estrogen receptor (discussed in Publications #9 and #10).

Some of these mechanisms involved in antiestrogen resistance were described in a review article (Publication #17).

(7) KEY RESEARCH ACCOMPLISHMENTS

We have shown that:

- Stimulation of the protein kinase A signaling pathway activates the stimulatory activity of tamoxifen-like antiestrogens and may contribute to the development of tamoxifen resistance.
- The N-terminal activation function-1 region of the estrogen receptor is responsible for tamoxifen's stimulatory activity.

- Elevated cAMP levels can compromise the growth suppressive activities of antiestrogens, rendering cells insensitive to these normally growth suppressive compounds.
- There is synergistic activation of transcription when breast cancer cells are treated with hormone and protein kinase A activator; this synergism does not require the phosphorylation of the two potential cAMP-dependent protein kinase phosphorylation sites on the estrogen receptor but may involve phosphorylation of important estrogen receptor coregulators such as SRC-1.
- Breast cancer cells resistant to the growth inhibitory effects of antiestrogen (our MCF/TOT cells or cells overexpressing ras or FGF-1 or FGF-4) contain substantially elevated levels of intracellular cAMP.
- Estrogens as well as antiestrogens and some growth factors and oncogenes increase intracellular cAMP.
- Antiestrogen resistance is reversible upon the removal of tamoxifen implying that resistance is adaptational and not explained by genetic/mutational alterations in the cells.
- These investigations highlight the important role of cAMP modulation of estrogen and antiestrogen action in hormonal resistance, and suggest new directions for more effective implementation of antiestrogen treatments in breast cancer patients that may prove to be more long-term and effective.

(8) REPORTABLE OUTCOMES

Manuscript Publications Resulting from this Research: (*indicates copy present in Appendix)

- *1. Fujimoto, N., and Katzenellenbogen, B. S. Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen-selectivity and promoter-dependence. Molec. Endocrinol. 8:296-304, 1994.
- *2. Aronica, S. M., Kraus, W. L., and Katzenellenbogen, B. S. Estrogen action via the cAMP signaling pathway: Stimulation of adenylate cyclase and cAMP-regulated gene transcription. Proc. Natl. Acad. Sci. USA 91:8517-8521, 1994.
- *3. Herman, M. E. and Katzenellenbogen, B. S. Alterations in transforming growth factor alpha and beta production and cell responsiveness during the progression of MCF-7 human breast cancer cells to estrogen-autonomous growth. Cancer Research 54:5867-5874, 1994.

- *4. Nicholson, R. I., Gee, J. M. W., Francis, A. B., Manning, D. L., Wakeling, A. E., and Katzenellenbogen, B. S. Observations arising from the use of pure antioestrogens on oestrogen-responsive (MCF-7) and oestrogen growth-independent (K3) human breast cancer cells. Endocrine Related Cancer 2:115-121, 1995.
- *5. Katzenellenbogen, B. S., Montano, M., Le Goff, P., Schodin, D. J., Kraus, W. L., Bhardwaj, B., and Fujimoto, N. Antiestrogens: Mechanisms and actions in target cells. J. Steroid Biochem. Molec. Biol. 53:387-393, 1995.
- *6. Ince, B. A., Schodin, D. J., Shapiro, D. J., and Katzenellenbogen, B. S. Repression of endogenous estrogen receptor activity in MCF-7 human breast cancer cells by dominant negative estrogen receptors. Endocrinology, 136: 3194-3199, 1995.
- *7. Nicholson, R. I., Gee, J. M., W., Manning, D. L., Wakeling, A. E., Montano, M. M., and Katzenellenbogen, B. S. Responses to pure antioestrogens (ICI 164384, ICI 182780) in oestrogen sensitive and resistant experimental and clinical breast cancer. Annals N. Y. Acad. Sci. 761:148-163, 1995.
- *8. Katzenellenbogen, B. S. Estrogen receptors: Bioactivities and interactions with cell signaling pathways. Biol. Reprod. 54:287-293, 1996.
- *9. Katzenellenbogen, J. A., O'Malley, B. W., and Katzenellenbogen, B. S. Tripartite steroid hormone receptor pharmacology: Interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. Molec. Endocrinol. 10:119-131, 1996.
- *10. Katzenellenbogen, J. A. and Katzenellenbogen, B. S. Nuclear hormone receptors: Ligand-activated regulators of transcription and diverse cell responses. Chem. and Biol. 3: 529-536, 1996.
- *11. Herman, M. E. and Katzenellenbogen, B. S. Response-specific antiestrogen resistance in a newly characterized MCF-7 human breast cancer cell line resulting from long-term exposure to trans-hydroxytamoxifen. J. Steroid Biochem. Molec. Biol. 59: 121-134, 1996.
- *12. Katzenellenbogen, B.S., Montano, M. M., Kraus, W. L., Aronica, S. M., Fujimoto, N., and LeGoff, P. Estrogen-receptor and antiestrogen-receptor complexes: cell and promoter specific effects and interactions with second messenger signaling pathways. In: Baird et al. (Eds.), Ernst Schering Foundation Workshop 16, Organ-Selective Actions of Steroid Hormones, Springer-Verlag Publishers, Heidelberg, Germany, pp. 29-50, 1995.

- *13. McInerney, E. M. and Katzenellenbogen, B. S. Different regions in activation function-1 of the human estrogen receptor required for antiestrogen-dependent and estradiol-dependent transcription activation. J. Biol. Chem. **271**: 24172-24178, 1996.
- *14. McInerney, E. M., Tsai, M. J., O'Malley, B. W., and Katzenellenbogen, B. S. Analysis of estrogen receptor transcriptional enhancement by a nuclear hormone receptor coactivator. Proc. Natl. Acad. Sci. USA **93**: 10069-10073, 1996.
- *15. Weis, K. E., Ekena, K., Thomas, J. A., Lazennec, G., and Katzenellenbogen, B. S. Constitutively active human estrogen receptors containing amino acid substitutions for tyrosine 537 in the receptor protein. Mol. Endocrinol. **10**: 1388-1398, 1996.
- *16. Montano, M. M., Kraus, W. L., and Katzenellenbogen, B. S. Identification of a novel transferable cis element in the promoter of an estrogen responsive gene that modulates sensitivity to hormone and antihormone. Mol. Endocrinol. **11**:330-341, 1997.
- *17. Katzenellenbogen, B. S., Montano, M. M., Ekena, K., Herman, M. E., and McInerney, E. M. Antiestrogens: Mechanisms of action and resistance in breast cancer. Breast Cancer Res. Treatment **44**:23-38, 1997.
- *18. Lazennec, G., Ediger, T. R., Petz, L. N., Nardulli, A. M., and Katzenellenbogen, B. S. Mechanistic aspects of estrogen receptor activation probed with constitutively active estrogen receptors: correlations with DNA and coregulator interactions and receptor conformational changes. Mol. Endocrinol. **11**:1375-1386, 1997.
- *19. Webb, P., Nguyen, P., Shinsako, J., Anderson, C., Feng, W., Nguyen, M.P., Chen, D., Huang, S-M., Subramanian, S., M., McInerney, E. M., Katzenellenbogen, B. S., Stallcup, M. R., and Kushner, P. J. Estrogen receptor activation function 1 works by binding p160 coactivator protein. Mol. Endocrinol. **12**:1605-1618, 1998

Published Abstracts on this Research:

- A1. Herman, M. E. and Katzenellenbogen, B. S. Alterations in estrogen and TGF- β pathways in antiestrogen-resistant MCF7 human breast cancer cells. Proceedings, Amer. Assoc. for Cancer Research **35**: Abstract 1527, p. 256, 1994.
- A2. Katzenellenbogen, B. S., Montano, M. M., Schodin, D. J., LeGoff, P., Aronica, S. M., and Cho, H. Estrogen receptor interactions with cell signaling pathways. J. Cell. Biochem. (Suppl. 18B): Abstract K018, p. 337, 1994.

- A3. Katzenellenbogen, B. S. Structure-activity relationships of estrogens and their antagonists: Estrogen receptor interactions and transcriptional regulation. Plenary Talk, First International Conference on Steroids and Bone, Florence, Italy, Calcified Tissue International 54:331, 1994.
- A4. Katzenellenbogen, B. S. Estrogen and antiestrogen actions in breast cancer: Regulation of proliferation and oncogene expression. Plenary Talk, Ninth Workshop on Vitamin D, Orlando, Florida, Proceedings, p. 171, 1994.
- A5. Katzenellenbogen, B. S., Montano, M., Herman, M., Schodin, D., Ince, B. A., Kraus, W. L., and Bhardwaj, B. Antiestrogens: Mechanisms and actions in target cells. Proceedings, IX International Congress on Hormonal Steroids, Dallas, Texas, 1994.
- A6. Katzenellenbogen, B. S. Estrogen receptor structure-function relationships and bioactivity regulation. Amer. Assoc. for Cancer Res. Special Conference on the Mechanism of Action of Retinoids, Vitamin D and Steroid Hormones. Whistler, British Columbia, January, 1995.
- A7. Katzenellenbogen, B. S. Estrogen receptor bioactivities and interactions with signal transduction pathways in breast cancer cells. Proceedings, Amer. Assoc. for Cancer Research 37:630, 1996
- A8. Katzenellenbogen, B. S. Estrogen receptor mechanisms and actions in target cells. Endocrinology 138 (Suppl.): Plenary lecture abstract L1, p.13, 1996.
- A9. Katzenellenbogen, B. S., Ekena, K., Lazennec, G., McInerney, E., Montano, M. M., and Katzenellenbogen J. A. Estrogen receptor mechanisms and activity regulation. EMBO Workshop on Nuclear Receptors, Lecture abstract, p.44, Sicily, Italy, May 1997.
- A10. Katzenellenbogen, B. S., Ekena, K., Lazennec, G., McInerney, E., Montano, M. M., and Katzenellenbogen J. A. Tripartite estrogen receptor pharmacology: Regulation of gene expression by estrogens and antiestrogens. Frontiers in Estrogen Action, Lecture abstract, p. 13-14, Key West, Florida, May, 1997.
- A11. Katzenellenbogen, B. S. Estrogen receptors: Mechanisms and actions in breast cancer. International Union Against Cancer (UICC) Study Group Meeting on Cancer Research, Woods Hole, Massachusetts, Lecture abstract, p.12, June 1997.
- A12. Katzenellenbogen, B. S., Montano, M. M., Ekena, K., Lazennec, G., Ediger, T., Choi, I., and Katzenellenbogen, J. A. New dimensions in estrogen receptor pharmacology. Keystone Symposium on Nuclear Receptor Gene Superfamily, Lake Tahoe, CA, March 1998.

- A13. Thomas, J. A., Lazennec, G., and Katzenellenbogen, B. S. CREB-estrogen receptor transcriptional synergy on estrogen regulated genes. Keystone Symposium on Nuclear Receptor Gene Superfamily, Lake Tahoe, CA, March 1998.
- A14. Katzenellenbogen, B. S., Montano, M. M., Ekena, K., Lazennec, G., Ediger, T., McInernery, E., Choi, I., Sun, J., Weis, K., and Katzenellenbogen, J. A. Estrogen receptor pharmacology. Symposium Abstract S39-1, p.42, 80th Annual Meeting of the Endocrine Society, New Orleans, LA, June 1998.
- A15. Katzenellenbogen, B. S., Ediger, T., Ekena, K., Sun, J., Choi, I., Weis, K., Martini, P., Montano, M., Delage-Mourroux. Molecular mechanisms of estrogen and antiestrogen actions via estrogen receptors alpha and beta. Keystone Symposium on Molecular Pathogenesis of Bone Disease, Lake Tahoe, CA., March 1999.
- A16. Katzenellenbogen, B. S., Ediger, T. R., Choi, I., Sun, J., Montano, M. M., Martini, P., Delage-Mourroux, R., Rajendran, R. Transcriptional regulation by estrogen receptors and insights into the actions of antiestrogens. Proceedings, Amer. Assoc. for Cancer Research 40:752-753. Symposium 6 Invited Talk: Transcriptional Regulation and Cancer, April 1999.
- A17. Katzenellenbogen, B. S., Choi, I., Delage-Mourroux, R., Ediger, T., Martini, P., Montano, M. M., Sun, J., Weis, K., and Katzenellenbogen, J. A. Molecular mechanisms of estrogen action: selective ligands and receptor pharmacology. Nobel Symposium on Estrogens and Women's Health - Benefit or Threat?, Karlskoga, Sweden, July 1999.

Employment or Research Opportunities Received Based on Experiences Supported by this Award:

Dr. William Lee Kraus, a Postdoctoral Researcher on this work, substantially benefited from his experiences and training such that he is currently an Assistant Professor in the Department of Biochemistry and Molecular Biology at Cornell University in Ithaca, NY.

Dr. Gwendal Lazennec, a Postdoctoral Researcher on this work, is now an Assistant Professor at the INSERM Unit 148, "Hormones and Cancer," Montpellier, France.

Dr. Kirk Ekena, a Postdoctoral Researcher on this work, has been able to move on to an independent research position at Mirus Corporation, Madison, WI.

Dr. Eileen McNerney had valuable research opportunities associated with this research, and is currently a very productive postdoctoral scientist at the Howard Hughes Medical Institute at the University of California at San Diego in the unit headed by Dr. Michael G. Rosenfeld.

(9) CONCLUSIONS: Implications and Importance of Our Research Findings

The results of our studies indicate that agents or factors that elevate cAMP in breast cancer cells reduce the effectiveness of tamoxifen-like antiestrogens used in hormonal therapy of breast cancer and may lead to antiestrogen resistance. In addition, we find that antiestrogens themselves can increase cAMP levels, rendering the antiestrogens less potent antagonists of estrogen action and more potent stimulators of estrogen-induced effects, resulting in compromising of the tumor growth suppressing activities of antiestrogens. Our observations in studies under this grant, which indicate that cells resistant to the growth suppressive affects of antiestrogen (including our MCF/TOT cells or cells overexpressing ras or FGF-1 or FGF-4) contain substantially elevated levels of intracellular cAMP, are consistent with the hypothesis that elevated cAMP levels may compromise the growth suppressive activities of antiestrogens, rendering the cells insensitive to these normally growth suppressive compounds.

In contrast to mechanisms for tamoxifen resistance that involve mutations in the estrogen receptor or other critical growth regulatory genes, which would not be reversible, this mechanism involving a compromising of tamoxifen effectiveness as an antiestrogen in the presence of elevated levels of intracellular cAMP, would be a progressive, adaptational response, which would be reversible upon cessation of tamoxifen therapy. Indeed, our findings in Publication #8, and also clinical experience support a mechanism of this type in that patients who become resistant to tamoxifen often return to a state of tamoxifen responsiveness after a period of alternate therapy (during which time cAMP levels in tumor cells may drop such that newly administered tamoxifen would again be effective as a growth suppressive agent). In addition, our data could account for the observation that hormonal resistance in model mammary tumor systems develops much more slowly to ICI 164,384 than to tamoxifen in that the agonistic character of ICI 164,384 is not augmented by cAMP. Therefore, ICI 164,384-like antiestrogens may prove to be more long-term effective antiestrogens compared with tamoxifen.

We have mutated the seven potential phosphorylation sites on the estrogen receptor, namely two cAMP-dependent protein kinase sites (S236 and S305), one site for MAP kinase (S118) and four additional phosphorylation sites for calmodulin-dependent protein kinase II (S154, S167, S338, and S518) since protein kinase phosphorylation cascades might result in the involvement of these sites in the transcriptional enhancement and magnified agonistic activity of antiestrogens. To our surprise, we found that elimination of phosphorylation at any one of these phosphorylation sites in the estrogen receptor did not eliminate the transcriptional

synergism elicited by cAMP. In addition, we have observed the involvement of the cAMP response element binding protein (CREB) in the synergistic activation of the estrogen receptor by hormone and protein kinase activators, and documented the importance of coregulator SRC-1 involvement and modulation of its potential phosphorylation state.

These investigations have provided insight into the role of cAMP modulation of estrogen and antiestrogen action in hormonal resistance. They allow an understanding of tamoxifen resistance at the molecular level, and thus point toward new directions for more effective implementation of antiestrogen treatments in breast cancer patients that may prove to be more long-term and effective.

(10) LIST OF PERSONNEL RECEIVING PAY FROM THE RESEARCH EFFORT

Bhardwaj, Bhavna (M.D.)
Ediger, Tracy
Ekena, Kirk (Ph.D.)
En, Julie
Hoeksema, Jason
Keller, Anne
Kraus, William Lee (Ph.D.)
Krueger, Kristopher
Lazennec, Gwendal (Ph.D.)
McInerney, Eileen (Ph.D.)
Sun, Jun
Thomas, James
Turpin, Katherine
Wilson, Carole
Xi, Xiaojan

(11) REFERENCES

1. Wakeling, AE (1993) The future of new pure antiestrogens in clinical breast cancer. Breast Canc. Res. Treat. 25:1-9.
2. Fisher, B, Costantino, JP, Wickerham, DL, Redmond, CK, Kavanah, M, Cronin, WM, Vogel, V, Robidoux, A, Dimitrov, N, Atkins, J, Daly, M, Wieand, S, Tan-Chiu, E, Ford, L and Wolmark, N (1998) Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. J. Natl. Cancer Inst. 90:1371-1388.
3. Pakdel, F and Katzenellenbogen, BS (1992) Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. J. Biol. Chem. 267:3429-3437.

4. Dickson, RB and Lippman, ME (1987) Estrogenic regulation of growth and polypeptide growth factor secretion in human breast carcinoma. Endocr. Rev. 8:29-43.
5. Read, LD and Katzenellenbogen, BS (1992). Characterization and regulation of estrogen and progesterone receptors in breast cancer. Genes, Oncogenes and Hormones: Advances in cellular and molecular biology of breast cancer. R. B. Dickson and M. E. Lippman. Nowell, Massachusetts, Kluwer-Nijhoff: 277-299.
6. Chalbos, D, Philips, A, Galtier, F and Rochefort, H (1993) Synthetic antiestrogens modulate induction of pS2 and cathepsin-D messenger ribonucleic acid by growth factors and adenosine 3',5'-monophosphate in MCF-7 cells. Endocrinology 133:571-576.
7. Reese, JR and Katzenellenbogen, BS (1991) Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: Studies using human estrogen receptor overexpressed in mammalian cells. Nucleic Acids Res. 19:6595-6602.
8. Fawell, SE, White, R, Hoare, S, Sydenham, M, Page, M and Parker, MG (1990) Inhibition of estrogen receptor-DNA binding by the "pure" antiestrogen ICI164,384 appears to be mediated by impaired receptor dimerization. Proc. Natl. Acad. Sci. USA 87:6883-6887.
9. Katzenellenbogen, BS (1991) Antiestrogen resistance: Mechanisms by which breast cancer cells undermine the effectiveness of endocrine therapy. J. Natl. Cancer Inst. 83:1434-1435.
10. Katzenellenbogen, BS, Fang, H, Ince, BA, Pakdel, F, Reese, JC, Wooge, CH and Wrenn, CK (1993) Estrogen receptors: Ligand discrimination and antiestrogen action. Breast Cancer Res. Treat. 27:17-26.
11. Fuqua, SAW, Fitzgerald, SD and Chamness, GC (1991) Variant human breast tumor estrogen receptor with constitutive transcriptional activity. Cancer Res. 51:105-109.
12. Murphy, LC and Dotzlaw, H (1989) Variant estrogen receptor mRNA species detected in human breast cancer biopsy samples. Mol. Endocrinol. 3:687-693.
13. Graham, ML, II, Krett, NL, Miller, LA and Horwitz, KB (1990) T47DCO cells, genetically unstable and containing estrogen receptor mutants, are a model for the progression of breast cancers to hormone resistance. Cancer Res. 50:6208-6217.

14. Katzenellenbogen, BS, Kendra, KL, Norman, MJ and Berthois, Y (1987) Proliferation, hormonal responsiveness, and estrogen receptor content of MCF-7 human breast cancer cells grown in the short-term and long-term absence of estrogens. Cancer Res. 47:4355-4360.
15. Clarke, R, Br  nner, N, Katzenellenbogen, BS, Thompson, EW, Norman, MJ, Koppi, C, Soonmyoung, P, Lippman, ME and Dickson, RB (1989) Progression of human breast cancer cells from hormone-dependent to hormone-independent growth both *in vitro* and *in vivo*. Proc. Natl. Acad. Sci. USA 86:3649-3653.
16. Jiang, S, Wolf, DM, Yingling, JM, Chang, C and Jordan, VC (1992) An estrogen receptor positive MCF-7 clone that is resistant to antiestrogens and estradiol. Mol. Cell. Endocrinol. 90:77-86.
17. Katzenellenbogen, BS, Montano, MM, Ekena, K, Herman, ME and McInerney, EM (1997) Antiestrogens: Mechanisms of action and resistance in breast cancer. Breast Cancer Res. Treat. 44:23-38.
18. Vassilomanolakis, ME, Tsoussis, S, Kandyliis, K, Hajichristou, E and Efredmidis, AP (1991) Rechallenge by tamoxifen in metastatic breast cancer: Prospective study of different dose levels. Breast Dis. 4:129-134.
19. Stoll, BA (1983) Intermittent antiestrogen therapy in advanced breast cancer. Cancer Treat. Rep. 67:98.
20. Stoll, BA (1983) Rechallenging breast cancer with tamoxifen therapy. Clin. Oncol. 9:347-351.
21. Minton, JP, Wisenbaugh, T and Matthews, RH (1974) Elevated cyclic AMP levels in human breast-cancer tissue. J. Natl. Cancer Inst. 53:283.
22. Cohen, LA and Chan, P (1974) Intracellular cAMP levels in normal rat mammary gland and adenocarcinoma: *in vivo* vs. *in vitro*. Life Sci. 16:107-115.
23. Silberstein, GB, Strickland, PS, Trumpbour, V, Coleman, S and Daniel, CW (1984) cAMP stimulates growth and morphogenesis of mouse mammary ducts. Proc. Natl. Acad. Sci. USA 81:4950-4954.
24. Miller, WR, Elton, RA, Dixon, JM, Chetty, U and Watson, DMA (1990) Cyclic AMP binding proteins and prognosis in breast cancer. Br. J. Cancer 61:263-266.

25. Miller, WR, Watson, DMA, Jack, W, Chetty, U and Elton, RA (1993) Tumour cyclic AMP binding proteins: An independent prognostic factor for disease recurrence and survival in breast cancer. Breast Cancer Res Treat 26:89-94.
26. Kvinnsland, S, Ekanger, R, Døskeland, SO and Thorsen, T (1983) Relationship of cyclic AMP binding capacity and estrogen receptor to hormone sensitivity in human breast cancer. Breast Cancer Res. Treat. 3:67-72.
27. Watson, DMA, Hawkins, RA, Bundred, NJ, Stewart, HJ and Miller, WR (1987) Tumour cyclic AMP binding proteins and endocrine responsiveness in patients with inoperable breast cancer. Br. J. Cancer 56:141-142.
28. Stampfer, MR (1982) Cholera toxin stimulation of human mammary epithelial cells in culture. In Vitro 18:531-537.
29. Taylor-Papadimitriou, J, Purfis, P and Fentiman, IS (1980) Cholera toxin and analogues of cyclic AMP stimulate the growth of cultured human mammary epithelial cells. J. Cell Physiol. 102:317-321.
30. Sheffield, LG and Welsch, CW (1985) Cholera-toxin-enhanced growth of human breast cancer cell lines in vitro and in vivo: Interaction with estrogen. Int. J. Cancer 36:479-483.
31. Fissore, F, Fortunati, N, Comba, A, Fazzari, A, Gaidano, G, Berta, L and Frairia, R (1994) The receptor-mediated action of sex steroid binding protein (SBP, SHBG): Accumulation of cAMP in MCF-7 cells under SBP and estradiol treatment. Steroids 59:661-667.
32. Licitra, EJ and Liu, JO (1996) A three-hybrid system for detecting small ligand-protein receptor interactions. Proc. Natl. Acad. Sci. USA 93:12817-12821.

Alteration in the Agonist/Antagonist Balance of Antiestrogens by Activation of Protein Kinase A Signaling Pathways in Breast Cancer Cells: Antiestrogen Selectivity and Promoter Dependence

Nariaki Fujimoto and Benita S. Katzenellenbogen

Department of Physiology and Biophysics
University of Illinois
Urbana, Illinois 61801

We find that stimulation of the protein kinase A (PKA) signaling pathway in MCF-7 human breast cancer cells changes the agonist/antagonist activity of tamoxifen and related antiestrogens; it activates or enhances their estrogen agonist activity and reduces their ability to antagonize the effects of estradiol (E_2). In MCF-7 human breast cancer cells which contain high levels of endogenous estrogen receptor (ER), the antiestrogen *trans*-hydroxy-tamoxifen (TOT) fails to stimulate transcription of the estrogen-responsive promoter-reporter constructs estrogen response element (ERE)-TATA-chloramphenicol acetyl transferase (CAT), (ERE)₂-TATA-CAT, and pS2-CAT. However, when cells are treated with isobutyl methylxanthine plus cholera toxin (which increases intracellular cAMP approximately 10-fold), or with 8-bromo-cAMP, or are transfected with expression vectors for the PKA catalytic subunits, the transcriptional activity of the antiestrogen-ER complex is now increased, to levels 20–75% that of E_2 , and TOT also becomes much less effective in antagonizing the stimulation of transcription by E_2 . Although this alteration in the agonist and antagonist activity of TOT is observed with three promoter-reporter constructs, containing a simple TATA promoter or a more complex, pS2 promoter, elevation of cAMP did not enhance the transcription by either TOT or E_2 of the reporter plasmid ERE-thymidine kinase-CAT. Thus, this phenomenon is promoter specific. The maximal stimulatory effects of isobutylmethylxanthine plus cholera toxin and PKA catalytic subunits on TOT and E_2 transcriptional enhancement were not additive, consistent with the hypothesis that they are both acting via stimulation of the same signal transduction pathway. By contrast, cAMP and PKA catalytic subunit transfection failed to evoke any transcription by the more pure

antiestrogen ICI164,384 with any of the four promoter-reporter constructs tested. Our findings, documenting that stimulation of the PKA signaling pathway activates the agonist activity of tamoxifen-like antiestrogens, may in part explain the development of tamoxifen resistance by some ER-containing breast cancers. They also suggest that the use of antiestrogens like ICI164,384, that fail to activate ER transcription in the presence of cAMP, may prove more effective for long-term antiestrogen therapy in breast cancer. (Molecular Endocrinology 8: 296–304, 1994)

INTRODUCTION

The estrogen receptor (ER), a member of a large nuclear hormone receptor superfamily, binds steroidal or nonsteroidal ligands and functions as a hormone-activated transcription regulator. Upon binding estrogen, the receptor is believed to bind most usually to estrogen response element (ERE) DNA, often located in the 5'-flanking region of estrogen-responsive genes (1–5). The estrogen-occupied receptor is then thought to interact with transcription factors and other components of the transcriptional complex to modulate gene transcription (6, 7). The actions of estrogens are antagonized by antiestrogens which bind to the ER in a manner that is competitive with estrogen, but antiestrogens fail to effectively activate gene transcription (8–13). Data suggest that the hormone and antihormone receptor complexes display different conformations which are dependent on the nature of the ligand (5, 12, 14, 15). Presumably, the transcription apparatus reads an antiestrogen-ER complex differently from an estrogen-ER complex, possibly through interaction with factors exclusive for one complex or the other.

Estrogen increases proliferative and metastatic activity of breast cancer cells, in part via the induction of

growth factors, growth factor receptors, basement membrane receptors, and proteases (16–18), effects believed to be largely antagonized by antiestrogens. Using the present knowledge of the mechanisms of estrogen action, breast cancer patients are chosen for hormonal therapy on the basis of their hormone receptor status. Approximately two-thirds of breast cancer patients that have tumors containing good levels of the ER respond well to antiestrogen therapy, most commonly treatment with the antiestrogen tamoxifen, due to its effectiveness and minimal side effects (10, 11, 19). There remain about one-third of ER-positive breast cancer patients who, for unknown reasons, do not respond to antiestrogen therapy. Likewise, tumor resistance to antiestrogens frequently develops over time, even in patients who respond well initially (20). This resistance presents a major impediment to the long-term effectiveness of such antiestrogen treatments. There is intense interest in understanding why most breast cancer cells inevitably progress from a hormone-responsive to a hormone-resistant phenotype and in elucidating the alterations in the cells that accompany this change to hormonal resistance (21).

We have shown recently that treatment of breast cancer cells and other cells with agents that increase intracellular cAMP can result in synergistic activation of ER-mediated transcription by estradiol (E_2) (22). In this study, we have therefore examined the effects of cAMP on the transcriptional activity of antiestrogen-liganded ER complexes, since such alterations in transcriptional activity might underlie the stimulatory effects of antiestrogens, such as tamoxifen, that are observed during the progression of cells toward antiestrogen resistance.

In these studies, we have compared the effects of cAMP on the transcriptional activity of ER complexes occupied by the widely used type 1 tamoxifen-like antiestrogens, known to sometimes exhibit partial agonist/antagonist gene activity, and type 2 (more pure antagonist) antiestrogens (8, 10, 12, 20); and we have used several different estrogen-responsive promoter-reporter gene constructs containing both simple and more complex promoters. Differences in the character of the type 1 and 2 antiestrogen ligands suggest that they may exert their actions through different mechanisms and that the nature of the interaction of antiestrogen-occupied receptor complexes with chromatin may be influenced by which of the two classes of antiestrogens occupy the receptor (12, 13, 23, 24).

We find that increasing the intracellular concentration of cAMP, or of protein kinase A (PKA) catalytic subunits by transfection, activates and/or enhances the transcriptional activity of type 1 but not type 2 antiestrogen-occupied ER complexes and reduces the estrogen antagonist activity of the type 1 antiestrogens. These findings imply the important involvement of PKA signaling pathways in the activity of antiestrogen-ER complexes and indicate that cAMP can alter the estrogen agonist/antagonist balance of antiestrogens in breast cancer cells. Stimulation of the PKA signal transduction pathway can, therefore, change the interpretation of

the character of the antiestrogen ligand, changing it from an estrogen antagonist to a partial, but significant, estrogen agonist.

RESULTS

Increases in Intracellular cAMP Can Activate the Transcriptional Activity of Some Antiestrogen-ER Complexes and Alter the Agonist/Antagonist Balance of Antiestrogens

As seen in Fig. 1, we examined the effects of cAMP on the transcriptional activity of ERs exposed to E_2 and several antiestrogens. For these studies, we used MCF-7 human cancer cells, which contain high levels of endogenous ERs. Using the estrogen-responsive reporter construct pATC2, which contains two estrogen response elements (EREs) upstream of a TATA promoter and chloramphenicol acetyl transferase (CAT) gene, transfected into MCF-7 cells, we observed an

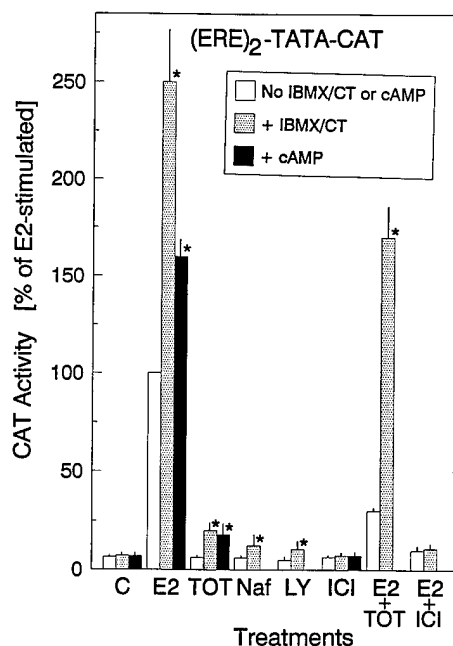


Fig. 1. Effect of IBMX/CT or cAMP on the Ability of E_2 and Antiestrogens to Stimulate *Trans*-Activation of $(ERE)_2$ -TATA-CAT, and on the Ability of Antiestrogens to Suppress E_2 -Stimulated *Trans*-Activation

MCF-7 cells were transfected with the reporter plasmid $(ERE)_2$ -TATA-CAT plus an internal control plasmid that expresses β -galactosidase. The cells were treated with the indicated agents for 24 h and harvested for CAT assay. The CAT activity of the E_2 -treated cells was set at 100%. Each bar represents the mean \pm SEM ($n = 3$ –6 experiments). *, Significant difference ($P < 0.05$ by Student's t test) from cells receiving the indicated treatment but no IBMX/CT or cAMP (open bar). C, Control ethanol vehicle; E_2 , 10^{-9} M; TOT, 10^{-6} M; Naf, 10^{-6} M; LY, 10^{-6} M; ICI, 10^{-6} M; IBMX/CT, 10^{-4} M IBMX and 1 μ g/ml CT; cAMP, 10^{-4} M 8-Br-cAMP.

approximately 12-fold increase in CAT activity by E_2 but no stimulation by any of the four antiestrogens tested. However, when cells were exposed to isobutylmethylxanthine (IBMX) and cholera toxin (CT), which result in marked increases in intracellular cAMP in these cells (15-fold by 1 h and 5-fold at 24 h; Ref. 25), three of the antiestrogens [*trans*-hydroxytamoxifen (TOT), Nafoxidine (Naf), and LY117018 (LY)] now showed stimulatory transcriptional activity, approximately 15–25% of that of E_2 alone. The antiestrogen ICI164,384 (ICI) showed no stimulation of transcription in cells in the presence or absence of IBMX/CT (Fig. 1, *stippled bar*) or in cells exposed to 8-bromo-cAMP (Fig. 1, *black bar*), while these treatments enhanced transcription by E_2 - and TOT-occupied ERs. As reported previously (22), IBMX/CT markedly enhanced the transcriptional activity of the E_2 -liganded ER complex, increasing its activity to approximately 250% of that of the E_2 -ER complex alone in the absence of cAMP stimulators. Of interest, although the antiestrogen ICI fully blocked the transcriptional response to E_2 in the presence and absence of IBMX/CT, TOT was much less effective in blocking E_2 transcriptional activity in the presence of IBMX/CT than in its absence. Hence, IBMX/CT changed both the agonist activity and the estrogen antagonist activity of some, but not all, antiestrogens.

Stimulation of the Agonist Activity of Antiestrogens by cAMP Is Promotor Dependent

In order to determine the generality of this phenomenon, we examined the effects of cAMP on the transcriptional activity of antiestrogens and E_2 in the same (MCF-7) cells, but using several different estrogen-responsive promoter-reporter constructs. In Fig. 2, A and B, we have determined the effect of cAMP on the activity of TOT, ICI, and E_2 on a simple TATA promoter with one consensus ERE upstream of the CAT gene (pATC1) and on the more complex pS2 gene promoter and 5'-flanking region (–3000 to +10) containing an imperfect ERE (26–28). The endogenous pS2 gene is regulated by E_2 in MCF-7 breast cancer cells (29, 30). E_2 increased the transcription of both of these gene constructs, and treatment with IBMX/CT and E_2 evoked a synergistic increase in transcription, with activity being approximately 2.5-fold that of E_2 alone. Both antiestrogens (TOT and ICI) failed to stimulate *trans*-activation of these reporter gene constructs, but in the presence of IBMX/CT, TOT gave significant stimulation of transcription (85% or 60% that of E_2 alone; Fig. 2, A or B). ICI failed to stimulate *trans*-activation even in the presence of IBMX/CT, and ICI fully blocked E_2 stimulation in the presence or absence of cAMP. By contrast, treatment with IBMX/CT reduced the ability of TOT to inhibit E_2 *trans*-activation. While TOT returned E_2 inhibition down to that of the control in the absence of IBMX/CT (Fig. 2, A and B, compare *open bars*, E_2 vs. E_2 + TOT), TOT only partially suppressed the E_2 stimulation in the presence of IBMX/CT (Fig. 2, A and B, compare *stippled bars*, E_2 vs. E_2 + TOT).

With ERE-thymidine kinase (tk)-CAT, an estrogen-

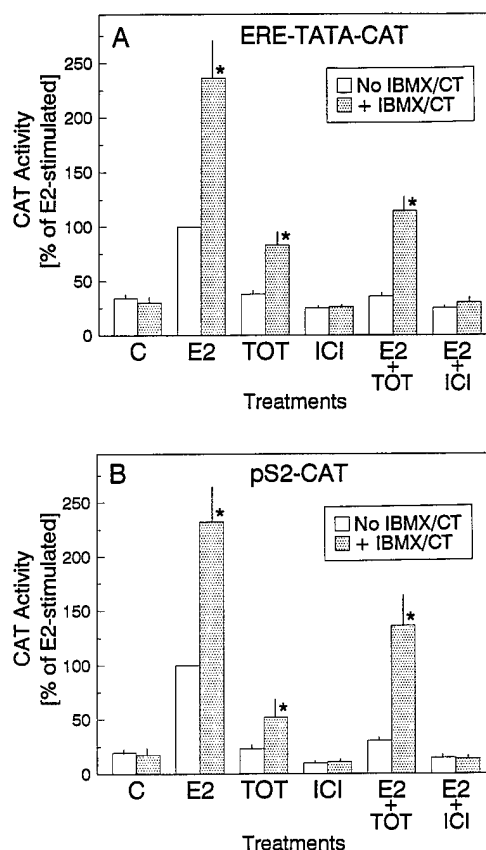


Fig. 2. Effect of IBMX/CT on the Ability of E_2 and Antiestrogens to Stimulate *Trans*-Activation of ERE-TATA-CAT (A) and pS2-CAT (B) and on the Ability of Antiestrogens to Suppress E_2 -Stimulated *Trans*-Activation

MCF-7 cells were transfected with the indicated reporter plasmid and an internal control plasmid that expresses β -galactosidase and were treated with the agents indicated for 24 h. Each bar represents the mean \pm SEM ($n = 3$ experiments). *, Significant difference from the cells with no IBMX/CT ($P < 0.05$ by Student's t test). C, Control ethanol vehicle; E_2 , 10^{-9} M; TOT, 10^{-6} M; ICI, 10^{-6} M; IBMX, 10^{-4} M; and CT, 1 μ g/ml.

responsive promoter-gene construct containing the tk promoter, however, we found no effect of IBMX/CT on transcriptional activity of either E_2 or antiestrogen-receptor complexes (Fig. 3A). The antiestrogens TOT, Naf, and LY alone showed severalfold increases in CAT activity (5- to 10-fold that of the control), but neither their activity, nor that of E_2 , was altered by IBMX/CT treatment. TOT and ICI also both effectively suppressed E_2 activity in the presence or absence of IBMX/CT. As seen in Fig. 3B, we observed no enhancement of E_2 *trans*-activation by IBMX/CT over a broad concentration range of ligand, and we observed the same absence of transcriptional enhancement with a broad range of TOT concentrations (10^{-9} – 10^{-6} M, data not presented).

Increasing the Levels of cAMP-Dependent PK α - or β -Catalytic Subunits in MCF-7 Cells Stimulates the Transcriptional Activity of E_2 -Occupied and TOT-Occupied ERs but Not ICI-Occupied ERs

The studies in Fig. 4 show that exposure of MCF-7 cells to E_2 or to the antiestrogen TOT in the presence

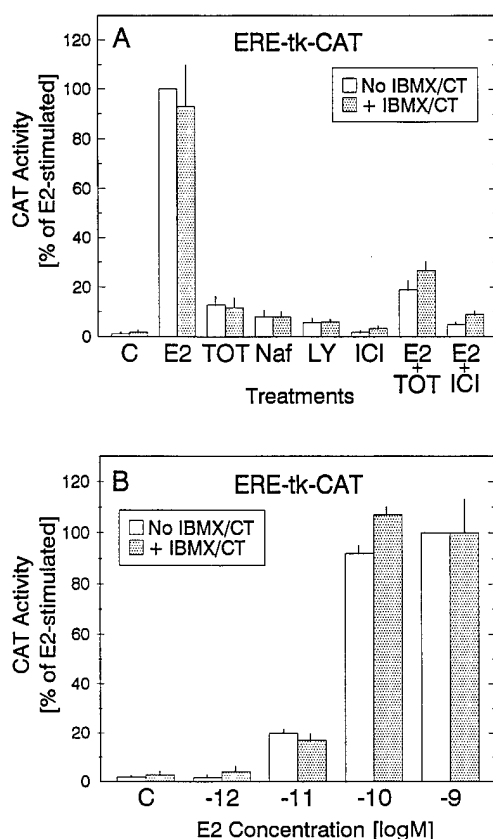


Fig. 3. Effect of IBMX/CT on the Ability of E₂ and Antiestrogens to Stimulate *Trans*-Activation of ERE-tk-CAT and on the Ability of Antiestrogens to Suppress E₂-Stimulated *Trans*-Activation.

MCF-7 cells transfected with ERE-tk-CAT were treated with E₂ or antiestrogens (A) or various concentrations of E₂ (B) in the presence or absence of IBMX/CT for 24 h. Each bar presents the mean \pm SEM ($n = 3$ experiments). *, Significant difference from the treatment alone in the absence of IBMX/CT ($P < 0.05$ by Student's t test). C, Control ethanol vehicle; E₂, 10^{-9} M; TOT, 10^{-6} M; Naf, 10^{-6} M; LY, 10^{-6} M; ICI, 10^{-6} M; IBMX, 10^{-4} M; and CT, 1 μ g/ml.

of IBMX/CT (which raises intracellular cAMP) results in increased transcriptional activity of the receptor complexes. Since cAMP is a PKA activator, we asked whether directly increasing the intracellular level of cAMP-dependent protein kinase catalytic subunit isoforms α or β by transfection might also bring about transcriptional enhancement. Expressing either the PK α - or β -catalytic subunits in MCF-7 cells resulted in a marked enhancement of transcription by the E₂-ER or TOT-ER (but not ICI-ER) complexes, equivalent in magnitude to that evoked by IBMX/CT alone (Fig. 4). Treatment of cells expressing PK α - or β -catalytic subunits with IBMX/CT plus E₂ or TOT failed to enhance transcription above that achieved by ligand in the presence of either IBMX/CT or cAMP-dependent PK subunit transfection alone. As might have been expected, ERs remained transcriptionally inactive with the antiestrogen ICI in the presence of PK catalytic subunit or IBMX/CT, and these agents (IBMX/CT; PK α - or β -subunit) also failed to enhance the low transcriptional activity ob-

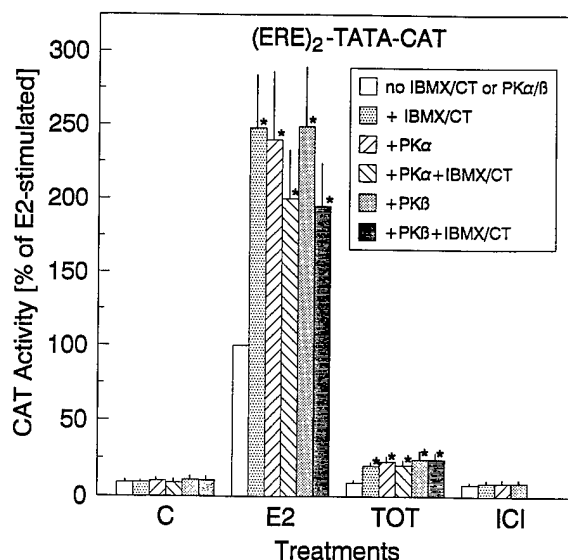


Fig. 4. Effect of Transfection of PK α - or β -Catalytic Subunit Expression Plasmids on the Ability of E₂ or Antiestrogens to Stimulate *Trans*-Activation of (ERE)₂-TATA-CAT in MCF-7 Cells

Cells were transfected with (ERE)₂-TATA-CAT reporter plasmid and were treated with control vehicle, E₂, TOT, or ICI in the presence or absence of IBMX/CT for 24 h, or were transfected with PK α - or β -expression plasmids and (ERE)₂-TATA-CAT. All cell transfections also included the internal control plasmid pCH110 to normalize for transfection efficiency. Eighty-micromolar ZnSO₄ was added to induce the mouse metallothionein promoter in the PK α/β constructs. Some cells were also exposed to IBMX/CT, as indicated. Each bar presents the mean \pm SEM ($n = 3$ or 4). *, Significant difference ($P < 0.05$ by Student's t test) from the no-PK α/β transfection and/or no-IBMX/CT treatment (open bar). C, Control ethanol vehicle; E₂, 10^{-9} M; TOT, 10^{-6} M; ICI, 10^{-6} M; IBMX, 10^{-4} M; CT, 1 μ g/ml; PK α , PK α -catalytic subunit plasmid transfected; PK β , PK β -catalytic subunit plasmid transfected.

served with the (ERE)₂-TATA-CAT plasmid in control MCF-7 cells.

As shown in Fig. 5, treatment of MCF-7 cells with IBMX/CT or transfection with the PK α - or β -catalytic subunit dramatically increased the transcriptional activation of a somatostatin (SRIF)-CAT reporter gene construct which contains a cAMP response element (CRE), documenting the increased cAMP-mediated, PKA-pathway activity in these cells.

PKA Activation Does Not Affect Antiestrogen or E₂ Modulation of ER Levels in MCF-7 Cells

We determined the level of ER in MCF-7 cells after treatment with E₂ and TOT, in the presence and absence of IBMX/CT, to determine if changes in ER levels might account for the enhanced transcriptional activity of ERs observed upon IBMX/CT and E₂ or TOT exposure. As shown in Fig. 6, IBMX/CT alone had no effect on the high ER levels in control cells. Likewise, the well documented decrease in ER level over 24 h brought about by E₂ exposure (13, for example), was unaffected by IBMX/CT, as was the absence of change from

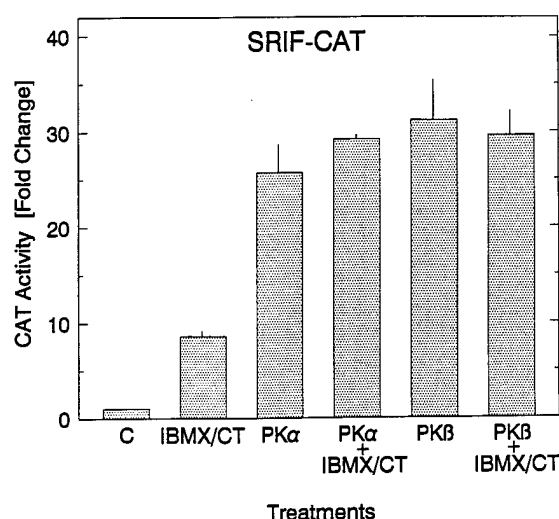


Fig. 5. Activation of SRIF-CAT Transcription by IBMX/CT and/or Transfection of PK α or PK β Expression Plasmids

MCF-7 cells were transfected with the SRIF-CAT plasmid which contains a CRE, PK α or β expression plasmids, and an internal control β -galactosidase pCH110 plasmid. Eighty-micromolar ZnSO₄ was used to induce the mouse metallothionein promoter in PK α/β constructs. Cells were harvested for assay at 24 h. Each bar presents the mean \pm SEM ($n = 3-5$ experiments). *, Significant difference from the control ($P < 0.05$ by Student's t test). C, Control ethanol vehicle; E₂, 10⁻⁹ M; TOT, 10⁻⁶ M; ICI, 10⁻⁶ M; IBMX, 10⁻⁴ M; and CT, 1 μ g/ml.

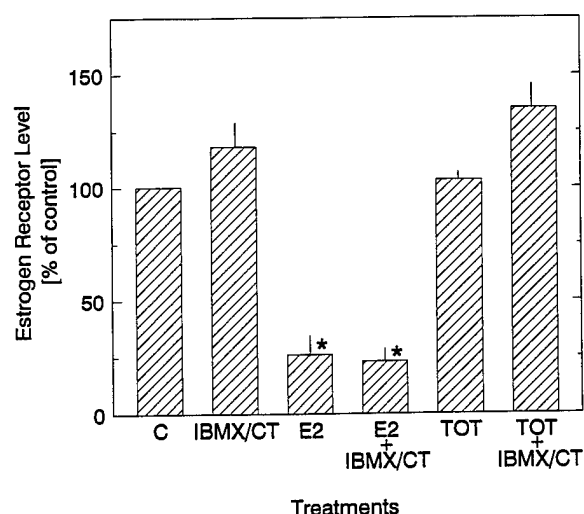


Fig. 6. Effects of E₂, TOT, and IBMX/CT on the Level of ER in MCF-7 Cells

Cells were treated with E₂ and TOT with and without IBMX/CT for 24 h. The cell extracts were analyzed by Western immunoblot with the ER-specific antibody H222. Autoradiograms were quantified with a densitometer. Each bar presents the mean \pm SEM ($n = 3-5$). *, Significant difference from the control ($P < 0.05$ by Student's t test). C, Control ethanol vehicle; E₂, 10⁻⁹ M; TOT, 10⁻⁶ M; IBMX, 10⁻⁴ M; and CT, 1 μ g/ml.

control ER level in cells treated with TOT alone or with PKA activator (IBMX/CT). Also, gel mobility shift assays showed that treatment of cells with IBMX/CT did not alter qualitatively or quantitatively the binding of TOT-liganded receptor complexes to ERE-DNA (data not presented), as observed previously for E₂-occupied receptors (22).

DISCUSSION

cAMP Changes the Interpretation of the Character of the Antiestrogen Ligand

These studies reveal that cAMP changes the interpretation of the character of the antiestrogen ligand, changing it from an ineffective transcriptional activator and an effective antagonist of estrogen-stimulated transcription to an agonist which now stimulates estrogen-dependent gene transcription. cAMP also blunts the antagonist activity of the tamoxifen antiestrogen, reducing its ability to antagonize E₂-stimulated transcription, an observation that may have important implications for understanding antiestrogen resistance in breast cancer, which often occurs despite the presence of functionally intact ERs (21).

Three aspects revealed by our studies are of note. First, cAMP only affected the agonist/antagonist activity of so called type 1 antiestrogens like TOT. The type 2, pure antiestrogen ICI remained a pure antiestrogen despite changes in intracellular levels of cAMP or cAMP-dependent protein kinase. Second, the cAMP enhancement/activation of gene transcription by TOT was promoter dependent and occurred with several, but not all, estrogen-responsive promoter-reporter constructs examined. Interestingly, constructs showing synergistic activation of ER-mediated transcription by E₂ and PKA activators also showed synergistic activation of ER-mediated transcription by tamoxifen, while the ERE-tk promoter-CAT gene construct failed to exhibit synergistic activation by E₂ or TOT with PKA activators. Third, the fact that we could activate the transcriptional ability of TOT-occupied and E₂-occupied ERs by increasing cAMP or by increasing intracellular PKA implies the important involvement of cAMP-dependent PK signaling pathways in this transcriptional activation/enhancement.

Activation of the Transcriptional Activity of Antiestrogen-Occupied ERs Occurs Only with Type 1, Tamoxifen-Like Antiestrogens and Is Promoter Specific

Differences in the abilities of the four antiestrogens we examined to have their transcriptional activity activated or enhanced by cAMP may be a reflection of differences in their ability to allow the ER to form stable dimers and interact effectively with DNA. Several studies with ICI164,384, the antiestrogen that fully blocked E₂-stimulated transcriptional activation and failed to itself gen-

erate ER complexes that were activated by cAMP or PKA catalytic subunits, have shown that ICI reduces DNA binding of the ER and also impairs the stability of ER dimers (13, 23, 24). Likewise, in the case of the human progesterone receptor, antiprogesterone-occupied PR complexes that do not interact effectively with DNA also fail to have their transcriptional activity influenced by cAMP (31, 32). In contrast to ICI, the type 1 antiestrogen TOT, like E_2 , binds to ERs and receptors occupied by this antiestrogen or by E_2 bind to ERE-DNA *in vitro* or *in vivo* (12–14). These antiestrogen-receptor complexes, like the RU486 antiprogesterone and antigluco-corticoid-occupied progesterone receptor and glucocorticoid receptor complexes, act as hormone agonists in the presence of cAMP (31–34).

Studies in this and other laboratories over the past 15 yr have documented that estrogen antagonism is complex and that the agonist/antagonist character of antiestrogens depends upon the particular antiestrogen compound, the target cell and even the particular response or gene being evaluated (8, 10, 35). Our present studies indicate that cAMP can activate the agonist activity of tamoxifen-like antiestrogens on some but not all promoter-gene constructs. Although the response to antiestrogen in the presence of elevated cAMP was significant, the magnitude of the response was dependent on the promoter-gene construct and was always less than that observed with E_2 . We observed transcriptional activation/enhancement with constructs containing a minimal promoter (ERE-TATA-CAT) and with a more complex promoter construct, pS2-CAT, containing the pS2 promoter and 5'-flanking region. By contrast, we observed no transcriptional activation/enhancement using the ERE-tk-CAT construct containing the tk promoter which contains binding sites for several transcription factors. Of note also, when we saw transcriptional enhancement or synergism with E_2 and PKA activator, we also saw synergism with tamoxifen-like antiestrogens; and with the tk promoter we failed to see transcriptional synergism with cAMP and E_2 or TOT. Although we do not know the basis for this promoter-specific transcriptional activation/enhancement, these promoters contain binding sites for different transcription factors and interact with different protein factors. It is possible that cAMP may influence the phosphorylation state of various coactivator/adaptor proteins or transcription factors that mediate promoter-specific interactions with the ER (see below).

Mechanisms and cAMP-Dependent Signaling Pathway Involvement in Activation of the Transcriptional Activity of ERs Occupied by Tamoxifen-Like but Not ICI-Like Antiestrogens

Our data provide strong evidence for the involvement of cAMP-dependent signaling pathways in the agonist actions of tamoxifen-like estrogen antagonists. It seems reasonable to assume then that the transcriptional response in the presence of cAMP may result from phosphorylation or other conformational changes in the ER

itself or in other transcription factors or adaptor proteins with which the ER interacts to enhance ER-specific transcription. While we do not know what these components are, we do know that cAMP exposure increases by approximately 4-fold the overall phosphorylation of the human ER (36–38) and rat ER (39) and that there is synergistic activation of E_2 -liganded receptors by PKA-mediated pathways (22). This occurs with no change in ERE gel mobility shift patterns with E_2 - or antiestrogen-occupied ERs in the presence of cAMP, suggesting that cAMP does not change the DNA interaction of the receptor. However, since we know that E_2 , as well as the antiestrogens TOT and ICI, enhance phosphorylation of the ER (36–39), and show quite similar phosphotryptic peptide patterns (37), the role of cAMP-induced and hormone-induced ER phosphorylation in transcriptional enhancement is not straightforward and may implicate contributions of both receptor phosphorylation as well as phosphorylation of other factors involved in ER-specific transcription.

The promoter specificity of the transcriptional enhancement phenomenon suggests that factors in addition to ER are probably being modulated by PKA pathway stimulation. Clearly, there is still much we need to learn about how PKA activation affects the phosphorylation/bioactivity of ER and/or other transcription factors or adapter/coactivator proteins with which the ligand-occupied ER interacts so as to preferentially evoke promoter-dependent transcriptional enhancement by tamoxifen and E_2 , but not the antiestrogen ICI164,384. In contrast to the observed cAMP stimulation of ER phosphorylation, enhancement of progesterone receptor transcription by okadaic acid (a protein phosphatase inhibitor), and cAMP stimulation of glucocorticoid receptor and progesterone receptor transcriptional enhancement did not appear to be associated with major changes in phosphorylation of these receptors (31, 32, 40), suggesting, in these cases, that transcriptional enhancement resulted from alterations in components other than the receptor itself.

Implications for Antiestrogen Resistance in Breast Cancer

Our studies imply that changes in the cAMP content of cells, which can result in activation of the agonist activity of tamoxifen-like antiestrogens, might account, at least in part, for the resistance to antiestrogen therapy that is observed in some breast cancer patients. Of interest, MCF-7 cells transplanted into nude mice fail to grow with tamoxifen treatment initially, but some hormone-resistant cells grow out into tumors after several months of tamoxifen exposure (10, 41, 42). Studies have shown that this resistance to tamoxifen is, more correctly, a reflection of tamoxifen stimulation of proliferation, representing a change in the interpretation of the tamoxifen-ER complex and its agonist/antagonist balance. It is of interest that we found the pS2 gene, which is under estrogen and antiestrogen regulation in breast cancer (26–30), to be activated by tamoxifen in

the presence of elevated cAMP. By contrast, however, antiestrogens such as ICI, shown in many systems to be more complete estrogen antagonists, are not changed in their agonist/antagonist balance by increasing intracellular concentrations of cAMP. Therefore, ICI-like compounds may prove to be more efficacious and less likely to result in antiestrogen-stimulated growth (20).

High consumption of caffeine and methyl xanthines has been thought to be associated with breast cysts and benign breast lumps, conditions associated with estrogenic stimulation (43). This might in part reflect elevated cAMP brought about by suppression of cAMP phosphodiesterase by methylxanthines like caffeine and consequent transcriptional enhancement by estrogens. CT and cAMP analogs have also been shown to stimulate growth of cultured human mammary epithelial cells (44) and human breast cancer cells (45), and to enhance the growth and morphogenesis of mouse mammary ducts (46). In this regard, it is noteworthy that cAMP levels are significantly higher in breast tumors than in normal breast tissue (47, 48) and that elevated concentrations of cAMP binding proteins are associated with early disease recurrence and poor survival rates (49, 50).

There is evidence now that MCF-7 breast cancer cells that become resistant to tamoxifen can have their growth suppressed by the more pure ICI antiestrogen (20, 51). Therefore, resistance to tamoxifen-like antiestrogens does not necessarily result in resistance to more pure ICI-like antiestrogens. Our data would be consistent with these biological observations and would suggest that fluctuations in cAMP cellular concentrations, which might accompany progression or changes in the proliferative or metastatic state of breast cancers, may influence the outcome and response to tamoxifen but not to the more pure antiestrogens. These more pure antiestrogens may therefore prove to be less problematic in long-term treatment protocols.

The transcriptional enhancement we have observed between PKA activators and ER occupied by tamoxifen-like antiestrogens and E_2 provides further evidence for cross-talk between the ER and signal transduction pathways regulated by cAMP that are important in ER-dependent transcription.

MATERIALS AND METHODS

Chemicals

Estradiol-17 β , IBMX, CT and 8-bromo-cAMP were purchased from Sigma Chemical Co. (St. Louis, MO). The antiestrogens *trans*-4-hydroxytamoxifen and ICI164,384 were kindly provided by ICI Pharmaceuticals (Macclesfield, UK). The antiestrogens LY 117018 and Nafoxidine (U11,100A) were generously provided by the Eli Lilly Co. (Indianapolis, IN) and the Upjohn Co. (Kalamazoo, MI), respectively.

Plasmids

The plasmids ERE-TATA-CAT, (ERE) $_2$ -TATA-CAT, ERE-tk-CAT, and pS2-CAT were used as estrogen-responsive tran-

scriptional reporters. In ERE-TATA-CAT and (ERE) $_2$ -TATA-CAT, described in Ref. 52, one or two consensus EREs are directly linked to the TATA box of the *Xenopus* vitellogenin B1 promoter (-42/+14). In ERE-tk-CAT, a consensus ERE was inserted into the polycloning region at the *Bam*HI site of pTZ-tk-CAT. pTZ-tk-CAT was constructed by cloning the 1.82-kilobase *Bam*HI/*Bam*HI fragment of pS2-tk-CAT (30), which includes the -150 to +55 region of the herpes simplex virus tk promoter, the bacterial CAT gene, and SV40 splice and polyadenylation signals, into the *Hind*III site of pTZ19R (53) with the tk promoter and CAT gene oriented 3' to 5' relative to the T7 promoter. The pS2-CAT construct has 3000 base pairs of the 5'-flanking region (-3000/+10) of the human estrogen-responsive (pS2) gene that includes the promoter and estrogen-responsive region (30).

For expression of the catalytic subunits of cAMP-dependent PK, pC α EV and pC β EV, which contain a mouse metallothionein promoter followed by catalytic subunits α/β and a human GH polyadenylation signal, were used (54). These encode two isoforms of the catalytic subunit of PKA; although derived from separate genes, the protein isoforms are very similar, with more than 90% of the amino acids being the same (55). The SRIF-CAT reporter contains the -71 to +53 region of the rat somatostatin gene in which there is a CRE (56). It is referred to as pSRIF Δ (-71) (56).

ERE-TATA-CAT and (ERE) $_2$ -TATA-CAT were kindly provided by Dr. David Shapiro (University of Illinois, Urbana, IL). pC α EV and pC β EV were gifts from Dr. Stanley McKnight (University of Washington, Seattle, WA). SRIF-CAT was kindly provided by Dr. Marc Montminy (Salk Institute, La Jolla, CA).

A β -galactosidase expression vector, pCH110 (Pharmacia, Piscataway, NJ) was used as an internal control for transfection efficiency in all transfection experiments. pTZ19 was used as carrier DNA.

MCF-7 Cell Culture

MCF-7 human breast cancer cells were grown in Eagle's minimum essential medium (MEM; GIBCO, Grand Island, NY) containing 5% calf serum (Hyclone, Logan, UT), 25 μ g/ml gentamycin, 100 U/ml penicillin, and 100 μ g/ml streptomycin. Before the experiments, the cells were cultured in MEM containing 5% dextran coated charcoal treated calf serum (CDCS) and the above antibiotics for 1 week and finally in phenol red-free MEM containing 5% CDCS and the same antibiotics for 1 more week.

Transfection

Transient transfection and CAT assays were carried out as described previously (22). Briefly, MCF-7 cells were plated in phenol red-free MEM containing 5% CDCS. They were transfected by the calcium phosphate-DNA coprecipitation method followed by glycerol shock. In all of the experiments, 15 μ g total plasmid DNA were used [2 μ g (ERE) $_2$ -TATA-CAT or ERE-tk-CAT, 11 μ g ERE-TATA-CAT or pS2-CAT, 4 μ g SRIF-CAT, 4 μ g pCH110, 50 ng pC α EV or pC β EV]. At 6 h after transfection, the cells were shocked with 25% glycerol in MEM containing 5% CDCS, and they were then treated with E_2 or antiestrogens and/or 1 μ g/ml CT plus 10^{-4} M IBMX. For zinc induction of the metallothionein promoter of pC α EV and pC β EV, 80 μ M ZnSO $_4$ was added. Cells were harvested at 24 h and prepared for CAT assay as described (22). All cell transfections included the pCH110 β -galactosidase plasmid as an internal control to correct for transfection efficiency between plates (22).

Immunoblots

Whole cell extracts were used in Western immunoblot analyses as described previously (22). The anti-ER monoclonal antibody H222, kindly provided by Abbott Laboratories (North

Chicago, IL), was used as the primary antibody, and rabbit antirat immunoglobulin G (Zymed Lab, Inc., San Francisco, CA) was used as the second antibody. The immune complexes were detected with [¹²⁵I]protein-A (New England Nuclear, Boston, MA), and autoradiograms were quantified by densitometry (22).

Acknowledgments

We thank Stanley McKnight for providing protein kinase A expression plasmids, Marc Montminy for providing the SRIF-CAT plasmid, David Shapiro for providing the plasmids (ERE)₂-TATA-CAT and ERE-TATA-CAT, and Pierre Chambon for providing the pS2 promoter and 5'-flanking region used in our preparation of the pS2-CAT reporter plasmid. We also thank W. Lee Kraus and Joseph Reese of our laboratory for preparation of the plasmid ERE-tk-CAT. We are grateful to Abbott Laboratories for providing ER antibodies, ICI Pharmaceuticals for providing the antiestrogens *trans*-hydroxy-tamoxifen and ICI164,384, the Eli Lilly Company for providing the antiestrogen LY117018, and the Upjohn Company for providing the antiestrogen Nafoxidine.

Received September 9, 1993. Revision received December 1, 1993. Accepted December 13, 1993.

Address requests for reprints to: Prof. Benita S. Katzenellenbogen, Department of Physiology and Biophysics, University of Illinois, 524 Burrill Hall, 407 S. Goodwin Avenue, Urbana, Illinois 61801.

This work was supported by NIH Grants CA-18119 and CA-51482 (to B.S.K.).

REFERENCES

1. Beato M 1989 Gene regulation by steroid hormones. *Cell* 56:335-344
2. Evans RM 1988 The steroid and thyroid hormone receptor superfamily. *Science* 240:889-895
3. Carson-Jurica MA, Schrader W, O'Malley B 1990 Steroid receptor family: structure and functions. *Endocr Rev* 11:201-220
4. Ryffel GU, Klein-Hitpass L, Druege P, Doebbeling U, Heitlinger E, Cato AC 1988 The estrogen-responsive DNA-element: Structure and interaction with the estrogen receptor. *J Steroid Biochem* 35:219-222
5. Kumar V, Chambon P 1988 The estrogen receptor binds tightly to its responsive element as a ligand-induced homodimer. *Cell* 55:145-156
6. Green S, Chambon P 1991 The oestrogen receptor: from perception to mechanism. In: Parker M (ed) *Nuclear Hormone Receptors*. Academic Press, New York, pp 15-38
7. Meyer ME, Gronemeyer H, Turcotte B, Bocquel M, Tasset D, Chambon P 1989 Steroid hormone receptors compete for factors that mediate their enhancer function. *Cell* 57:433-442
8. Katzenellenbogen BS, Miller MA, Mullick A, Sheen YY 1985 Antiestrogen action in breast cancer cells: modulation of proliferation and protein synthesis, and interaction with estrogen receptors and additional antiestrogen binding sites. *Breast Cancer Res Treat* 5:231-243
9. Thompson EW, Katz D, Shima TB, Wakeling AE, Lippman ME, Dickson RB 1989 ICI164,384, a pure antagonist of estrogen-stimulated MCF-7 cell proliferation and invasiveness. *Cancer Res* 49:6929-6934
10. Jordan VC, Murphy CS 1990 Endocrine pharmacology of antiestrogens as antitumor agents. *Endocr Rev* 11:578-610
11. Santen R, Manni A, Harvey H, Redmond C 1990 Endocrine treatment of breast cancer in women. *Endocr Rev* 11:221-265
12. Reese JR, Katzenellenbogen BS 1991 Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor overexpressed in mammalian cells. *Nucleic Acids Res* 19:6595-6602
13. Reese JC, Katzenellenbogen BS 1992 Examination of the DNA binding abilities of estrogen receptor in whole cells: implications for hormone-independent transactivation and the action of the pure antiestrogen ICI164,384. *Mol Cell Biol* 12:4531-4538
14. Pakdel F, Katzenellenbogen BS 1992 Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J Biol Chem* 267:3429-3437
15. Brown M, Sharp P 1990 Human estrogen receptor forms multiple DNA complexes. *J Biol Chem* 265:11238-11243
16. Dickson RB, Lippman ME 1987 Estrogenic regulation of growth and polypeptide growth factor secretion in human breast carcinoma. *Endocr Rev* 8:29-43
17. Lippman ME, Dickson RB, Gelmann EP, Knabbe C, Jasid K, Bates S, Swain S 1987 Mechanisms of estrogenic and antiestrogenic regulation of growth of human breast carcinoma. In: Klijn JGM (ed) *Hormonal Manipulation of Cancer: Peptides, Growth Factors, and New (anti)Steroidal Agents*. Raven Press, New York, pp 381-403
18. Read LD, Katzenellenbogen BS 1991 Characterization and regulation of estrogen and progesterone receptors in breast cancer. In: Dickson RB, Lippman ME (eds) *Genes, Oncogenes and Hormones: Advances in Cellular and Molecular Biology of Breast Cancer*. Kluwer-Nijhoff Publishers, Nowell, Massachusetts, pp 277-299
19. Patterson JS 1987 Ten years of tamoxifen in breast cancer: a review. In: Klijn JGM (ed) *Hormonal Manipulation of Cancer: Peptides, Growth Factors, and New (Anti)Steroidal Agents*. Raven Press, New York, pp 1-6
20. Wakeling AE 1993 The future of new pure antiestrogens in clinical breast cancer. *Breast Cancer Res Treat* 25:1-9
21. Katzenellenbogen BS 1991 Antiestrogen resistance: mechanisms by which breast cancer cells undermine the effectiveness of endocrine therapy. *J Natl Cancer Inst* 83:1434-1435
22. Cho H, Katzenellenbogen BS 1993 Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Mol Endocrinol* 7:441-452
23. Fawell SE, White R, Hoare S, Sydenham M, Page M, Parker MG 1990 Inhibition of estrogen receptor-DNA binding by the "pure" antiestrogen ICI 164,384 appears to be mediated by impaired receptor dimerization. *Proc Natl Acad Sci USA* 87:6883-6887
24. Arbuckle ND, Dauvois S, Parker MG 1992 Effects of antiestrogens on the DNA binding activity of oestrogen receptor *in vitro*. *Nucleic Acids Res* 20:3839-3844
25. Cho H, Aronica SM, Katzenellenbogen BS, Regulation of progesterone receptor gene expression in MCF-7 breast cancer cells: a comparison of the effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology*, in press
26. Masiakowski P, Breathnach R, Bioch J, Gannon F, Krust A, Chambon P 1982 Cloning of cDNA sequences of hormone-regulated genes from the MCF-7 human breast cancer cell line. *Nucleic Acids Res* 10:7895-7903
27. Jakowlew SB, Breathnach R, Jeltsch JM, Masiakowski P, Chambon P 1984 Sequence of the pS2 mRNA induced by estrogen in the human breast cancer cell line MCF-7. *Nucleic Acids Res* 12:2861-2878
28. Jeltsch JM, Roberts M, Schatz C, Garnier JM, Brown AMC, Chambon P 1987 Structure of the human oestrogen-responsive gene pS2. *Nucleic Acids Res* 15:1401-1414
29. Brown AMC, Jeltsch JM, Roberts M, Chambon P 1984 Activation of pS2 gene transcription is a primary response

- to estrogen in the human breast cancer cell line MCF-7. *Proc Natl Acad Sci USA* 81:6344-6348
30. Weaver CA, Springer PA, Katzenellenbogen BS 1988 Regulation of pS2 gene expression by affinity labeling and reversibly binding estrogens and antiestrogens: comparison of effects on the native gene and on pS2-chloramphenicol acetyltransferase fusion genes transfected into MCF-7 human breast cancer cells. *Mol Endocrinol* 2:936-945
31. Sartorius CA, Tung L, Takimoto GS, Horwitz KB 1993 Antagonist-occupied human progesterone receptors bound to DNA are functionally switched to transcriptional agonists by cAMP. *J Biol Chem* 268:9262-9266
32. Beck CA, Weigel NL, Moyer ML, Nordeen SK, Edwards DP 1993 The progesterone antagonist RU486 acquires agonist activity upon stimulation of cAMP signaling pathways. *Proc Natl Acad Sci USA* 90:4441-4445
33. Gruol DJ, Altschmied J 1993 Synergistic induction of apoptosis with glucocorticoids and 3',5'-cyclic adenosine monophosphate reveals agonist activity by RU 486. *Mol Endocrinol* 7:104-113
34. Nordeen SK, Bona BJ, Moyer ML 1993 Latent agonist activity of the steroid antagonist, RU486, is unmasked in cells treated with activators of protein kinase A. *Mol Endocrinol* 7:731-742
35. Berry M, Metzger D, Chambon P 1990 Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-2818
36. LeGoff P, Aronica SM, Katzenellenbogen BS, Phosphorylation of wild type and mutant estrogen receptor transiently expressed in COS-1 cells: enhancement by estrogen, antiestrogen and protein kinase activators. *Proceedings of the 74th Annual Meeting of the Endocrine Society, San Antonio, Texas, 1992, p 230 (Abstract 716)*
37. LeGoff P, Montano MM, Schodin DJ, Katzenellenbogen BS, Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem*, in press
38. Ali S, Metzger D, Bornert J, Chambon P 1993 Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J* 12:1153-1160
39. Aronica SM, Katzenellenbogen BS 1993 Stimulation of estrogen receptor-mediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic adenosine monophosphate, and insulin-like growth factor-1. *Mol Endocrinol* 7:743-752
40. Somers JP, DeFranco DB 1992 Effects of okadaic acid, a protein phosphatase inhibitor, on glucocorticoid receptor-mediated enhancement. *Mol Endocrinol* 6:26-34
41. Osborne CK, Coronado E, Allred DC, Wiebe V, DeGregorio M 1991 Acquired tamoxifen resistance: correlation with reduced breast tumor levels of tamoxifen and isomerization of trans-4-hydroxytamoxifen. *J Natl Cancer Inst* 83:1477-1482
42. Gottardis MM, Jordan VC 1988 Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res* 48:5183-5187
43. Minton JP 1979 Benign breast disease tied to coffee, tea, cocoa, cola. *Med World News* 19:1-2
44. Taylor-Papadimitriou J, Purfis P, Fentiman IS 1980 Cholera toxin and analogues of cyclic AMP stimulate the growth of cultured human mammary epithelial cells. *J Cell Physiol* 102:317-321
45. Sheffield LG, Welsch CW 1985 Cholera-toxin-enhanced growth of human breast cancer cell lines *in vitro* and *in vivo*: interaction with estrogen. *Int J Cancer* 36:479-483
46. Silberstein GB, Strickland P, Trumpbour V, Coleman S, Daniel CW 1984 *In vivo* cAMP stimulates growth and morphogenesis of mouse mammary ducts. *Proc Natl Acad Sci USA* 81:4950-4954
47. Minton JP, Wisenbaugh T, Matthews RH 1974 Elevated cyclic AMP levels in human breast-cancer tissue. *J Natl Cancer Inst* 53:283
48. Cohen LA, Chan P 1974 Intracellular cAMP levels in normal rat mammary gland and adenocarcinoma: *In vivo* vs. *in vitro*. *Life Sci* 16:107-115
49. Miller WR, Elton RA, Dixon JM, Chetty U, Watson DMA 1990 Cyclic AMP binding proteins and prognosis in breast cancer. *Br J Cancer* 61:263-266
50. Miller WR, Watson DMA, Jack W, Chetty U, Elton RA 1993 Tumour cyclic AMP binding proteins: an independent prognostic factor for disease recurrence and survival in breast cancer. *Breast Cancer Res Treat* 26:89-94
51. Gottardis MM, Jiang SY, Jeng MH, Jordan VC 1989 Inhibition of tamoxifen-stimulated growth of an MCF-7 tumor variant in athymic mice by novel steroidal antiestrogens. *Cancer Res* 49:4090-4093
52. Chang T, Nardulli A, Lew D, Shapiro 1992 The role of estrogen response elements in expression of the *Xenopus laevis* Vitellogenin B1 gene. *Mol Endocrinol* 6:346-354
53. Mead D, Szczesna-Skorupa E, Kemper B 1986 Single-stranded DNA "blue" T7 promoter plasmids: a versatile tandem promoter system for cloning and protein engineering. *Protein Eng* 1:67-74
54. Uhler MD, McKnight GS 1987 Expression of cDNAs for two isoforms of the catalytic subunit of cAMP-dependent protein kinase. *J Biol Chem* 15202-15207
55. Spaulding SW 1993 The ways in which hormones change cyclic adenosine 3', 5'-monophosphate-dependent protein kinase subunits, and how such changes affect cell behavior. *Endocr Rev* 14:632-650
56. Montminy MR, Sevarino KA, Wagner JA, Mandel G, Goodman RH 1986 Identification of a cyclic-AMP-responsive element within the rat somatostatin gene. *Proc Natl Acad Sci USA* 83:6682-6686

Estrogen action via the cAMP signaling pathway: Stimulation of adenylate cyclase and cAMP-regulated gene transcription

(estrogens/antiestrogens/breast cancer/uterus)

SUSAN M. ARONICA, W. LEE KRAUS, AND BENITA S. KATZENELLENBOGEN

Department of Physiology and Biophysics, University of Illinois, Urbana, IL 61801

Communicated by Jack Gorski, May 9, 1994 (received for review January 28, 1994)

ABSTRACT Estrogenic hormones, believed to exert most of their effects via the direct interaction of their receptors with chromatin, are found to increase cAMP in target breast cancer and uterine cells in culture and in the intact uterus *in vivo*. Increases in intracellular cAMP are evoked by very low concentrations of estradiol (half maximal at 10 pM) and by other physiologically active estrogens and antiestrogens, but not by an inactive estrogen stereoisomer. These increases in cAMP result from enhanced membrane adenylate cyclase activity by a mechanism that does not involve genomic actions of the hormones (are not blocked by inhibitors of RNA and protein synthesis). The estrogen-stimulated levels of cAMP are sufficient to activate transcription from cAMP response element-containing genes and reporter plasmid constructs. Our findings document a nongenomic action of estrogenic hormones that involves the activation of an important second-messenger signaling system and suggest that estrogen regulation of cAMP may provide an additional mechanism by which this steroid hormone can alter the expression of genes.

For many years, steroid hormones and peptide hormones have been considered to act via distinctly different mechanisms, the former via intracellular receptors acting through the genome (1, 2) and the latter via membrane-localized receptors that initially affect extranuclear activities, including the generation of second messengers such as cAMP. However, there has been increasing evidence for interactions between cAMP and estrogen in enhancing the growth of the mammary gland and breast cancer cells (3, 4) and for cAMP induction of estrogen-like uterine growth (5). As early as 1967, Szego and Davis (6) demonstrated a very rapid, acute elevation of uterine cAMP by estrogen treatment of rats *in vivo* that was confirmed in other reports (7, 8), but several subsequent studies either failed to confirm this observation or reported only minimal effects that were considered to represent indirect effects of estrogen on cAMP mediated by estrogen-induced release of uterine epinephrine (9–12). Recently, cAMP and other protein kinase activators have been documented to synergize with steroid hormone-occupied receptors, leading to enhanced steroid receptor-mediated transcription (13–18), possibly by a mechanism involving phosphorylation of the receptor or associated transcription factors (14, 19–21).

In this paper, we show that estrogen activates adenylate cyclase, markedly increasing the concentration of cAMP in estrogen-responsive breast cancer and uterine cells in culture and in the intact uterus of rats treated with estrogen *in vivo*, in a manner that does not require new RNA or protein synthesis. The intracellular concentrations of cAMP achieved by low, physiological levels of estrogen are substantial and sufficient to stimulate cAMP response element

(CRE)-mediated gene transcription. Therefore, this nongenomic action of the steroid hormone estrogen involves the production of an important second messenger and the resultant activation of second messenger-stimulated genes. These findings document a two-way directionality in the cross talk between steroid hormone- and cAMP-signaling pathways.

MATERIALS AND METHODS

Plasmid DNA. The SRIF-CAT plasmid (22), containing the CRE region of the somatostatin [somatotropin release-inhibiting factor (SRIF)] gene promoter linked to the gene encoding chloramphenicol acetyltransferase (CAT), was kindly provided by Marc Montminy (Salk Institute, La Jolla, CA). PR_d-CAT, containing the distal promoter of the rat progesterone receptor (PR) gene linked to the CAT gene, has been described (23). pS2-CAT was constructed by replacing the thymidine kinase (tk) promoter of pTZ-tk-CAT (23), released by digestion with *Bam*HI and *Bgl* II, with the promoter region of the gene encoding protein pS2 (–90 to +10; ref. 24), which was released from a genomic clone provided by Pierre Chambon (Institut National de la Santé et de la Recherche Médicale, Strasbourg, France) by *Bam*HI digestion. The PR_d and pS2 promoter regions do not contain any estrogen-response elements (EREs) or CREs and are unresponsive to 17 β -estradiol (E₂) and cAMP (refs. 23 and 24 and our unpublished observations). CRE-pS2-CAT and CRE-PR_d-CAT were constructed by annealing the oligonucleotide 5'-TGGCTGACGTCAGAGA-3' with its complement and cloning the resulting double-stranded oligomer, which contains the CRE of the rat somatostatin gene and its immediate flanking sequences (22) into *Sma* I-digested pS2-CAT and PR_d-CAT, respectively.

Cell Culture, Transfections, and CAT Assays. Primary cultures of uterine cells from 18-day-old Sprague–Dawley rats (Holtzman, Madison, WI) were prepared and maintained as described (14). MCF-7 human breast cancer cells were maintained as described (23) and were switched to culture medium lacking phenol red 2 days prior to plating for experiments. For cAMP studies, uterine and MCF-7 cells were plated at a density of $1.5\text{--}2 \times 10^6$ cells per 160-mm dish. For transfection experiments, both cell types were plated at $4\text{--}5 \times 10^6$ cells per 100-mm dish, and the medium was changed 48 hr later. Cells were transfected 24 hr later as described (14, 23) with 15 μ g of DNA containing 5 μ g of CAT reporter plasmid, 3 μ g of pCH110 β -galactosidase reporter plasmid, and 7 μ g of pTZ19 carrier DNA. Treatments were added in fresh medium following glycerol shock, and cells

Abbreviations: CAT, chloramphenicol acetyltransferase; CRE, cAMP response element; CT, cholera toxin; E₂, 17 β -estradiol; ER, estrogen receptor; ERE, estrogen response element; PDE, phosphodiesterase; RIA, radioimmunoassay; TOT, the antiestrogen *trans*-4-hydroxytamoxifen; IBMX, isobutylmethylxanthine; SRIF, somatotropin release-inhibiting factor (i.e., somatostatin); PR, progesterone receptor.

were harvested 24 hr later. CAT assays, normalized for the β -galactosidase activity of each extract, were performed as described (14).

cAMP Radioimmunoassay (RIA). Nearly confluent plates of cells were treated with various agents and were harvested in 500 μ l of TNE (40 mM Tris-HCl, pH 7.5/140 mM NaCl/1.5 mM EDTA). Cells were scraped loose using a rubber policeman, collected into microcentrifuge tubes kept on ice, and pelleted ($100 \times g$ for 5 min). Cell pellets were resuspended in 200 μ l of ice-cold cAMP extraction buffer (50 mM Tris-HCl, pH 7.5/4 mM EDTA). For whole uterus treated *in vivo*, the tissue was cut into 1-mm pieces prior to homogenization. Extracts remained on ice for 15 min, with vigorous mixing every 5 min. Extract aliquots were analyzed for protein content by the BCA (bicinchoninic acid) method (Pierce). Extracts were boiled for 10 min, and cell debris was pelleted. A [3 H]cAMP RIA kit and instructions (Amersham) were used to measure cAMP content of samples. Incubations were carried out at 4°C for 3 hr, and charcoal-dextran-treated samples were analyzed for radioactivity.

Adenylate Cyclase Assay. Membrane preparations isolated from cultured cells or uteri were tested for adenylate cyclase activity (25). In brief, homogenates in 75 mM Tris-HCl, pH 7.4/2 mM EDTA/8 mM $MgCl_2$ were subjected to two centrifugations ($1000 \times g$ for 5 min at 4°C and $40,000 \times g$ for 20 min at 4°C). Membrane protein (20 μ g) was used from each sample. Reaction conditions were as described (25) except that 0.4 μ Ci (1 μ Ci = 37 kBq) of [α - 32 P]ATP was used per tube and the incubation time was extended to 20 min. Reactions were terminated upon addition of 1 mM ATP and 0.1 μ Ci of [3 H]cAMP. Aliquots were assayed for ^{32}P and 3H as a measure of total radioactivity in each sample.

The amount of [^{32}P]ATP converted into cAMP was determined by immunoprecipitation. Precleared samples were incubated with polyclonal antiserum directed against cAMP (provided by V. Ramirez, University of Illinois) for 1 hr on ice. This antiserum elicited no detectable cross-reactivity with ATP, GTP, AMP, or cGMP in our assay system. A second bridging goat anti-rabbit monoclonal antibody was added for an additional 1 hr (Zymed), and antibody complexes were precipitated by using a 10% slurry of Zysorbin (Zymed) as described (14). Zysorbin cells were boiled for 10 min and pelleted. ^{32}P and 3H radioactivity were determined in the supernatant, and recovery of the known [3 H]cAMP cpm added (which ranged from 70% to 90%) was used to assess the efficacy of the precipitation and to adjust the ATP-to-cAMP conversion values accordingly. Adenylate cyclase activity is calculated as the percentage of [^{32}P]ATP converted to [^{32}P]cAMP during the 20-min incubation.

Phosphodiesterase (PDE) Assay. The method of Prosser (26), as adapted from Thompson *et al.* (27), was used to assess cAMP PDE activity in cells exposed to various agents. After homogenization in 50 mM Tris-HCl, pH 7.4/1.5 mM EDTA/10% (vol/vol) glycerol/0.6 M NaCl, samples were sonicated for 30 sec on ice (setting no. 3, Branson Sonifier 200) and centrifuged ($30,000 \times g$ at 4°C for 15 min). Supernatants (20 μ g of cell protein) were incubated with 10 μ g of snake venom (*Ophiophagus hannah*; Sigma) and 0.15 μ Ci of [3 H]cAMP in assay buffer (4 mM Tris/1 mM 2-mercaptoethanol/1 μ M AMP/1 mM $MgCl_2$ /0.6 μ M cAMP) at 30°C for 15 min. Reactions were stopped by addition of 1 ml of AG 1-X2 resin (Bio-Rad), diluted 1:1 with water, and centrifuged 10 min at $2500 \times g$. The supernatant was collected, added to a second 1-ml aliquot of the resin mixture, and centrifuged a second time. PDE activity of the final supernatant is expressed as pmol of AMP produced from the [3 H]cAMP per mg of extract protein per min.

RESULTS

We first examined the effects of estrogen on cAMP in two estrogen-responsive cell types—namely, MCF-7 human breast cancer cells, which contain high levels of endogenous estrogen receptor (ER) and primary cultures of rat uterine cells, which contain lower levels of ER (Fig. 1). In both cell systems in culture, we observed that the estrogen E_2 increased cAMP levels dramatically, as did the antiestrogens *trans*-hydroxytamoxifen (TOT) and ICI 164,384, although the response to antiestrogen was somewhat lower. Maximal, *ca.* 10-fold, increases in intracellular cAMP, monitored by RIA, were observed at 0.5 or 1 hr after exposure to E_2 in MCF-7 and uterine cells, respectively; these represented changes that were one-third to one-half of those achieved by exposure to cholera toxin (CT), a powerful stimulator of adenylate cyclase activity. Intracellular cAMP levels returned to the control level by 6 hr (MCF-7 cells) or 3 hr (uterine cells) despite continued exposure to hormonal ligands. By contrast, cAMP elevation was more prolonged in the presence of CT.

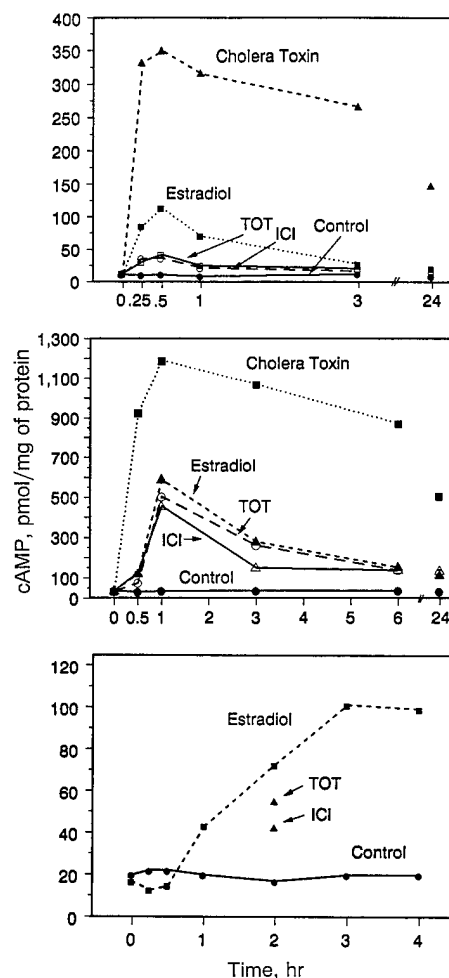


FIG. 1. Effect of estrogen and antiestrogens on the level of cAMP in uterine cells (Top), MCF-7 cells (Middle), and uterus *in vivo* (Bottom). Cultures of immature rat uterine cells (Top) or MCF-7 breast cancer cells (Middle) were treated with various agents and harvested at the times indicated. Cell extracts were monitored for cAMP content by RIA. Treatments were control vehicle (control), CT at 1 μ g/ml, 1 nM E_2 , 1 μ M TOT, or 1 μ M ICI 164,384 (ICI). Each point represents the mean from three experiments. (Bottom) Immature 18-day-old rats were given a single s.c. injection of E_2 (5 μ g), TOT or ICI (50 μ g), or control (0.2 ml of corn oil/ethanol vehicle). Uterine extracts were analyzed for cAMP content by RIA. Each point represents the mean from two experiments.

Exposure of rats to estrogen (E_2) or antiestrogen (TOT, ICI 164,384) *in vivo* also resulted in substantial increases in intracellular uterine cAMP, suggesting that the changes observed *in vitro* are neither artifactual nor associated only with *in vitro* cell culture. Tamoxifen *in vivo* has also been reported to alter cAMP accumulation in quail oviduct and mouse uterus (28).

Dose-response studies reveal that low, physiological concentrations of E_2 were able to stimulate cAMP accumulation (Fig. 2). Half-maximal stimulation was achieved with *ca.* 10 pM E_2 and full stimulation with 1 nM E_2 . Slightly higher concentrations of ICI 164,384 and TOT were needed for equal stimulation of intracellular cAMP, consistent with their lower availability in serum-containing medium and their restricted entry into cells. A nearly identical dose-response relationship was observed for cAMP stimulation by these ligands in rat uterine cells (data not shown).

The hormone specificity of the cAMP stimulation was examined in MCF-7 and uterine cells (Table 1). The potent estrogens E_2 and diethylstilbestrol (DES) evoked large fold increases in cAMP, although DES (at 1 nM) was less effective than E_2 , while the weaker estrogen 2-hydroxyestrone (2-OH-estrone) effected a substantial but smaller increase in cAMP; the E_2 stereoisomer 17 α -estradiol, known to be biologically ineffective as an estrogen, failed to alter intracellular cAMP. The glucocorticoid dexamethasone and the androgen testosterone failed to increase cAMP. As seen in Table 1, nearly identical results were observed in both cell types.

The ability of estrogen to increase cAMP was associated with increased adenylate cyclase activity, with no significant change in PDE activity (Fig. 3). Treatment of cells with CT evoked an increase of *ca.* 3-fold in adenylate cyclase activity within 20 min, while treatment with E_2 or the antiestrogens TOT and ICI 164,384 resulted in increases that were half as great. The effects of E_2 , CT, and ICI 164,384 were not additive. Similar increases in uterine adenylate cyclase activity were also observed after treatment of rats with these agents *in vivo* or after treatment of uterine cell cultures *in vitro* (data not shown). By contrast, E_2 , TOT, or ICI 164,384 did not alter PDE activity of MCF-7 cells (Fig. 3 *Right*) or uterine cells (data not shown); the potent inhibitor IBMX was effective in suppressing PDE activity, as expected.

To assess whether RNA or protein synthesis was required for the stimulatory effect of these agents on intracellular cAMP concentration, cells were pretreated with actinomycin D or cycloheximide for 30 min prior to the addition of hormone or CT, and cAMP was monitored 1 hr thereafter. Neither of these agents showed any inhibition of the estrogen- or antiestrogen- or CT-induced stimulation of cAMP,

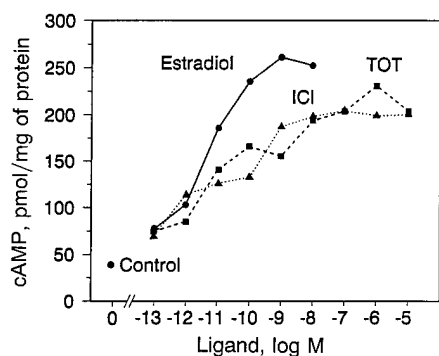


FIG. 2. Dose-response relationship for the increase in cAMP stimulated by estrogen and antiestrogens. MCF-7 cells were treated with various concentrations of E_2 , TOT, ICI 164,384 (ICI), or control ethanol vehicle for 1 hr. Cell extracts were monitored for cAMP content by RIA. Each point represents the mean from two experiments.

Table 1. Hormone specificity of the cAMP increase in target cells

Hormone	cAMP level, fold increase over control	
	MCF-7 cells	Uterine cells
E_2	9.2	8.2
DES	5.2	3.8
2-OH-estrone	3.6	3.6
17 α -Estradiol	1.1	1.4
Dexamethasone	1.1	1.2
Testosterone	1.0	0.8

Results shown are the mean from two experiments. MCF-7 or uterine cells were treated with various hormones (at 1 nM) for 1 hr or 30 min, respectively, and cell extracts were monitored for cAMP content by RIA. DES, diethylstilbestrol.

although they inhibited RNA or protein synthesis, respectively, by >95% (data not shown). This implies that stimulation of cAMP accumulation does not require RNA or protein synthesis. The very early time course of cAMP increase (Fig. 1) would also be consistent with the apparent absence of a requirement for RNA or protein synthesis.

Since estrogens and antiestrogens increased intracellular cAMP, we asked whether the changes in cAMP were significant enough to affect transcriptional activation of cAMP-regulated genes. To examine this, cells were transfected with a cAMP-responsive gene construct (SRIF-CAT) containing the CRE region of the somatostatin gene promoter (−71 to +55) linked to a CAT reporter gene (22). Cells were treated as shown in Fig. 4, and CAT activity was measured at 24 hr. E_2 exposure evoked a 5-fold (uterine cells) or 13-fold (MCF-7 cells) increase in CRE-mediated CAT activity, nearly equal to that evoked by exposure of cells to 0.1 mM 8-bromo-cAMP. As expected, CT evoked an even larger response based on its greater and more prolonged elevation of cAMP. TOT and ICI 164,384 also stimulated SRIF-CAT activity, although less dramatically than E_2 , consistent with their lesser stimulation of intracellular cAMP. The effects of E_2 and CT or cAMP were not additive, consistent with their action via the same pathway. The effects of E_2 plus antiestrogen were also not additive. Insulin-like growth factor 1, which failed to increase intracellular cAMP (data not shown), also failed to increase SRIF-CAT transcription.

The SRIF-CAT cAMP-responsive reporter gene construct contains the somatostatin promoter and flanking region (−71 to +55) and thus might potentially contain elements other than CREs that might contribute to the response to the hormonal agents. Therefore, we prepared two more simple consensus CRE-containing CAT reporter plasmids; the first contained one CRE upstream of the pS2 promoter (−90 to +10) and CAT gene (CRE-pS2-CAT), and the second contained one CRE upstream of the rat PR gene distal promoter and CAT gene (CRE-PR_d-CAT). Transactivation of these reporter genes was stimulated by estrogen and cAMP (Fig. 5), as seen for the SRIF-CAT plasmid, confirming that transactivation of the SRIF-CAT plasmid was indeed mediated via the CRE and, therefore, was due to intracellular cAMP generated by the hormone. The lack of response to E_2 or cAMP in control plasmids (pS2-CAT or PR_d-CAT) containing the pS2 or rat PR promoters and CAT reporter gene but lacking the CRE further confirms the requirement for the CRE in the transactivation.

The possibility of any binding of ER to consensus CRE monomers and dimers was analyzed by standard and competitive gel mobility-shift assays using human ER overexpressed in COS-1 cells (29). In the standard gel mobility-shift assay, direct binding of ER to a radiolabeled CRE dimer could not be detected, whereas binding of bacterially ex-

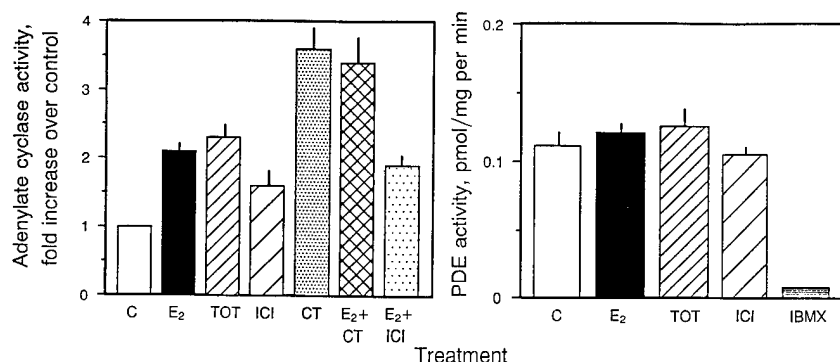


FIG. 3. Regulation of adenylate cyclase and PDE activities by estrogen and antiestrogens. MCF-7 cells were treated for 1 hr with ethanol vehicle (C, control), CT at 1 μ g/ml, 1 nM E₂, 1 μ M TOT, 1 μ M ICI 164,384 (ICI), 0.1 mM isobutylmethylxanthine (IBMX), or E₂ in combination with CT or ICI. Cell extracts were monitored for PDE activity (Right), or membrane fractions were monitored for adenylate cyclase activity (Left). Each bar represents the mean and range from two experiments. All adenylate cyclase values of treated groups were significantly different from the control ($P < 0.05$ by Student's *t* test), while only PDE activity of IBMX-treated cells was significantly different from the control.

pressed CREB protein (CRE-binding protein; provided by Richard Goodman, Vollum Institute, Portland, OR) was observed (not shown). In the competitive assay, where the binding of ER to radiolabeled consensus EREs was done in the presence of increasing amounts (up to 400-fold excess) of unlabeled CRE monomer or CRE dimer, no competition at all was observed, suggesting no interaction of ER with the CREs (not shown); however, with unlabeled ERE, a 5-fold molar excess gave nearly complete competition.

DISCUSSION

Three pieces of evidence indicate that the cAMP stimulation by estrogen and antiestrogens is mediated by a high-affinity hormone binder, possibly the ER: first, the steroid specificity and relative effectiveness of different estrogens in stimulation of cAMP (Table 1); second, the low concentrations of E₂ needed for the cAMP stimulation response (Fig. 2), with a half-maximum approximately that of the K_d for E₂ binding to ER; and third, the stimulation by estrogen of increased intracellular cAMP in ER-containing cells with little or no stimulation in cells lacking significant numbers of ERs (such as ER-negative 231 breast cancer cells and 3T3 mouse fibroblast cells, data not presented).

The implications of these observations may be significant. They suggest that estrogens and antiestrogens, by increasing cAMP within target cells, may modulate expression of cAMP-regulated genes and thereby possibly influence other

cAMP-regulated bioactivities. Although our studies document an increase in adenylate cyclase activity stimulated by estrogen, the mechanism by which estrogen enhances adenylate cyclase activity remains to be determined. It is of note that almost all responses to estrogen, even early ones such as the induction of creatine kinase B, require RNA synthesis, with some later responses to estrogen also often requiring prior protein synthesis (30, 31). Therefore, the cAMP increase evoked by estrogen is unusual in that prior RNA and protein synthesis are not required. It is also of interest that both estrogens and antiestrogens have stimulatory effects on cellular cAMP levels and enhance CRE-mediated gene transcription. Both categories of ligands also increase ER phosphorylation, and for both end-points, E₂ is somewhat more stimulatory than the antiestrogens (14, 19, 20). By contrast, antiestrogens fail to stimulate, or stimulate only weakly, the transcription of genes containing EREs, which are strongly activated by estrogen (refs. 13, 14, and 23 and references therein).

The clear change in adenylate cyclase activity in response to estrogen suggests a possible membrane action of this hormone, an aspect that will require further study. There is now evidence for ER-like binding sites for estrogens (32) and glucocorticoid receptor-like binding sites for glucocorticoids in the cell membrane (33). Earlier work (34) and more recent studies (35) have provided evidence for ER-like sites in the membrane and cytoplasmic portion of the cell that do not appear to translocate into the nuclear compartment. In the

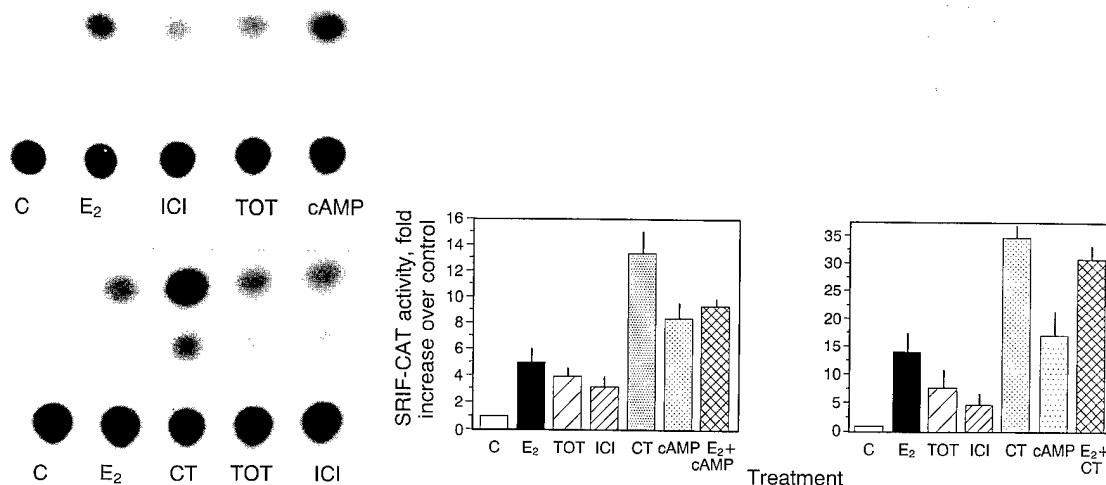


FIG. 4. Induction of CRE-mediated CAT activity by E₂, cAMP, TOT, and ICI 164,384 (ICI). (Left) Uterine cells (Left Upper) or MCF-7 cells (Left Lower) were transfected with the reporter plasmid SRIF-CAT plus an internal control plasmid that expresses β -galactosidase. Treatments were control vehicle (C), 1 nM E₂, 1 μ M ICI 164,384 (ICI), 1 μ M TOT, 0.1 mM 8-Br-cAMP (cAMP), or CT at 1 μ g/ml. Cells were harvested after 24 hr, and extracts were assayed for β -gal activity to normalize for transfection efficiency. Autoradiograms show CAT activity from transfected cells after the indicated treatments. (Center and Right) Summary of CAT data from uterine cells (Center) and MCF-7 cells (Right). Bars represent the mean and range from two experiments. All treatment values are significantly different from the control with $P < 0.05$ by Student's *t* test.

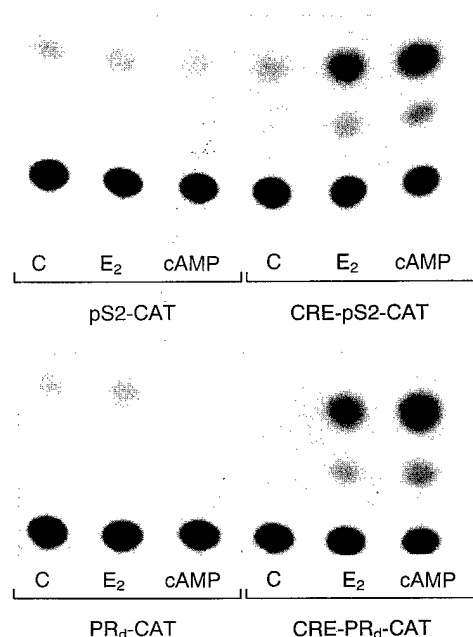


FIG. 5. Stimulation of CAT activity by estrogen and cAMP on target reporter genes containing minimal CREs. (Upper) Uterine cells were transfected with plasmid DNA containing either the promoter of the pS2 gene linked to the CAT gene (pS2-CAT) or the pS2-CAT construct containing a single consensus CRE placed upstream of the promoter (CRE-pS2-CAT) plus an internal control plasmid that expresses β -galactosidase. (Lower) Uterine cells were transfected with plasmid DNA containing either the distal promoter of the rat PR gene linked to the CAT gene (PR_d-CAT), or the PR_d-CAT construct containing a consensus CRE placed directly upstream from the PR promoter (CRE-PR_d-CAT) plus an internal control plasmid expressing β -galactosidase. Cells were treated for 24 hr with control vehicle (C), 1 nM E₂, or 0.1 mM 8-bromo-cAMP (cAMP). Cell extracts analyzed for CAT activity contained equal amounts of β -galactosidase activity. Autoradiograms show the CAT activity from transfected cells.

case of the steroid hormone progesterone, the evidence is for relatively high-affinity binding sites in the membrane with ligand binding selectivity and other properties clearly different from that of intracellular PRs (36). In addition, several recent reports present clear evidence for other nongenomic effects of E₂ and of vitamin D (37).

Considerable recent data indicate the modulation of steroid hormone receptor action by cAMP and by activation of protein kinase A as well as protein kinase C and tyrosine kinase pathways (13–18, 21, 38, 39). In addition, some growth factors, such as epidermal growth factor and insulin-like growth factor 1, which may act via alterations in signaling pathways involving protein kinases, have been shown to mimic some of the effects of estrogen (14, 40). cAMP has also been shown to be an important modulator of ER bioactivity, able to synergize with estrogen in enhancing ER transcriptional activity (13, 18). Our current observations, plus the reports that vitamin D action increases intracellular cGMP (41), imply a possibly broad involvement of steroid hormone action on cyclic nucleotide and second-messenger systems. Our data provide evidence for extensive two-way cross talk between estrogen and cAMP signaling pathways: in one way as shown previously (13, 14), cAMP can enhance the transcription of estrogen-regulated genes containing EREs; in the other way, as shown here, estrogens can act via the cAMP system to regulate cAMP-mediated gene expression. The

consequences of this in terms of understanding the biology and regulation of estrogen-responsive cells seem manifold.

We are very grateful to Drs. Marc Montminy and Pierre Chambon for providing plasmids, Dr. Richard Goodman for providing CRE-binding protein CREB, and Dr. Victor Ramirez for providing antiserum to cAMP. We thank Dr. Alan Wakeling and ICI Pharmaceuticals for providing the antiestrogens. This work was supported in part by National Institutes of Health Grants CA51482 and CA18119 and Predoctoral Traineeship 5T32GM7143.

- Gronemeyer, H. (1991) *Annu. Rev. Genet.* **25**, 89–123.
- Gorski, J., Furlow, J. D., Murdoch, F. E., Fritsch, M., Kaneko, K., Ying, C. & Malayer, J. R. (1993) *Biol. Reprod.* **48**, 8–14.
- Sheffield, L. G. & Welsch, C. W. (1985) *Int. J. Cancer* **36**, 479–483.
- Silberstein, G. B., Strickland, P. & Trumpbour, V. (1984) *Proc. Natl. Acad. Sci. USA* **81**, 4950–4954.
- Stewart, P. J. & Webster, R. A. (1983) *Biol. Reprod.* **29**, 671–679.
- Szego, C. M. & Davis, J. S. (1967) *Proc. Natl. Acad. Sci. USA* **58**, 1711–1718.
- Dupont-Maioresse, N., Van Sande, J., Rooryck, J., Fastrez-Boute, A. & Galand, P. (1974) *J. Steroid Biochem.* **5**, 173–178.
- Szego, C. M. & Davis, J. S. (1969) *Mol. Pharmacol.* **5**, 470–480.
- Chew, C. S. & Rinard, G. A. (1974) *Biochim. Biophys. Acta* **362**, 493–500.
- Rosenfeld, M. G. & O'Malley, B. W. (1970) *Science* **168**, 253–255.
- Sanborn, B. M., Bhalla, R. C. & Korenman, S. G. (1973) *Endocrinology* **92**, 494–499.
- Kuehl, F. A., Jr., Ham, E. A., Zanetti, M. E., Sanford, C. H., Nicol, S. E. & Goldberg, N. D. (1974) *Proc. Natl. Acad. Sci. USA* **71**, 1866–1870.
- Cho, H. & Katzenellenbogen, B. S. (1993) *Mol. Endocrinol.* **7**, 441–452.
- Aronica, S. M. & Katzenellenbogen, B. S. (1993) *Mol. Endocrinol.* **7**, 743–752.
- Sartorius, C. A., Tung, L., Takimoto, G. S. & Horwitz, K. B. (1993) *J. Biol. Chem.* **268**, 9262–9266.
- Beck, C. A., Weigel, N. L., Moyer, M. L., Nordeen, S. K. & Edwards, D. P. (1993) *Proc. Natl. Acad. Sci. USA* **90**, 4441–4445.
- Groul, D. J. & Altschmid, J. (1993) *Mol. Endocrinol.* **7**, 104–113.
- Fujimoto, N. & Katzenellenbogen, B. S. (1994) *Mol. Endocrinol.* **8**, 296–304.
- Le Goff, P., Montano, M. M., Schodin, D. J. & Katzenellenbogen, B. S. (1994) *J. Biol. Chem.* **269**, 4458–4466.
- Ali, S., Metzger, D. & Bornert, J. (1993) *EMBO J.* **12**, 1153–1160.
- Somers, J. P. & DeFranco, D. B. (1992) *Mol. Endocrinol.* **6**, 26–34.
- Montminy, M. R., Sevarino, K. A., Wagner, J. A., Mandel, G. & Goodman, R. H. (1986) *Proc. Natl. Acad. Sci. USA* **83**, 6682–6686.
- Kraus, W. L., Montano, M. M. & Katzenellenbogen, B. S. (1993) *Mol. Endocrinol.* **7**, 1603–1616.
- Berry, M., Nunez, A. & Chambon, P. (1989) *Proc. Natl. Acad. Sci. USA* **86**, 1218–1222.
- Lohse, M. J., Benovic, J. L., Caron, M. G. & Lefkowitz, R. J. (1990) *J. Biol. Chem.* **265**, 3202–3209.
- Prosser, R. A. (1989) Ph.D. thesis (Univ. Illinois, Urbana).
- Thompson, W. J., Terasaki, W. L., Epstein, P. M. & Strada, S. J. (1979) *Adv. Cyclic Nucleotide Res.* **10**, 69–92.
- Fanidi, A., Ahnadi, C., Fayard, J. M., Pageaux, J. F. & Laugier, C. (1992) *J. Steroid Biochem. Mol. Biol.* **41**, 571–577.
- Reese, J. C. & Katzenellenbogen, B. S. (1991) *Nucleic Acids Res.* **19**, 6595–6602.
- Katzenellenbogen, B. S. & Gorski, J. (1975) in *Biochemical Actions of Hormones*, ed. Litwack, G. (Academic, New York), Vol. 3, pp. 187–243.
- Katzenellenbogen, B. S. (1980) *Annu. Rev. Physiol.* **42**, 17–35.
- Pappas, T. C. & Watson, C. S. (1993) *Endocrinology* **132**, Suppl., 512 (abstr. 1846).
- Gametchu, B., Watson, C. S. & Wu, J. (1993) *FASEB J.* **7**, 1283–1292.
- Pietras, R. J. & Szego, C. M. (1979) *J. Steroid Biochem.* **11**, 1471–1483.
- Welshons, W. V., Grady, L. H. & Judy, B. M. (1993) *Mol. Cell. Endocrinol.* **94**, 183–194.
- Tischkau, S. A. & Ramirez, V. D. (1993) *Proc. Natl. Acad. Sci. USA* **90**, 1285–1289.
- Lieberherr, M., Grosse, B., Kachkache, M. & Balsan, S. (1993) *J. Bone Mineral Res.* **8**, 1365–1376.
- Rangarajan, P. N., Umesono, K. & Evans, R. M. (1992) *Mol. Endocrinol.* **6**, 1451–1457.
- Migliaccio, A., Pagano, M. & Auricchio, F. (1993) *Oncogene* **8**, 2183–2191.
- Ignar-Trowbridge, D. M., Teng, C. T., Ross, K. A., Parker, M. G., Korach, K. S. & McLachlan, J. A. (1993) *Mol. Endocrinol.* **7**, 992–998.
- Barsony, J. & Marx, S. J. (1988) *Proc. Natl. Acad. Sci. USA* **85**, 1223–1226.

Alterations in Transforming Growth Factor- α and - β Production and Cell Responsiveness during the Progression of MCF-7 Human Breast Cancer Cells to Estrogen-Autonomous Growth¹

Mary E. Herman and Benita S. Katzenellenbogen²

Department of Physiology and Biophysics, University of Illinois and University of Illinois College of Medicine, Urbana, Illinois 61801

ABSTRACT

Hormonal management of breast cancer is confounded by an almost inevitable progression of cell growth from a steroid-regulated to a steroid-autonomous state. We have experimentally induced this progression in the estrogen growth-responsive MCF-7 human breast cancer cell line by long-term culture in the absence of steroids. After an initial period (10–12 weeks) of slowed growth in response to steroid deprivation, rapid, steroid-independent growth rates were consistently established. In these cells, which contained 3-fold elevated, functional estrogen receptor levels (as determined by induction of PgR and transactivation of a transiently transfected estrogen-responsive gene construct), antiestrogens still effectively suppressed cell proliferation, although estrogens only minimally increased the proliferation rate. Depletion of steroids from the growth media also resulted in a marked (70–80%) transient decrease in transforming growth factor (TGF) α mRNA and TGF- α protein production at 2 weeks that was followed by a progressive, partial return to the initial parental TGF- α mRNA and protein levels. In contrast, the mRNAs for TGF- β 1, - β 2, and - β 3 and bioactive TGF- β proteins transiently increased (3–10-fold) at 2 to 10 weeks of steroid deprivation and then returned by 24 weeks to the lower levels of the parental MCF-7 cells. These results suggest that the cells acquired steroid-independent means to regulate the production of these peptides. The long-term steroid-deprived sublines showed a loss of regulation of proliferation by TGF- α or anti-TGF- α antibodies and a 10-fold decrease in sensitivity to the growth-suppressive effects of TGF- β 1, despite little change in receptor levels for these factors. The diminished contributions of TGF- α and TGF- β s to the regulation of cell proliferation in long-term steroid-deprived MCF-7 breast cancer cells suggest that the TGFs do not act as major growth regulators in these estrogen-autonomous sublines. However, the marked, transient alterations in the levels of these growth factors indicate that they may play a role in the events which accompany the progression from estrogen-responsive to estrogen-autonomous growth. In addition, continued exposure to estrogen may be needed for the long-term maintenance of cell responsiveness to these TGFs.

INTRODUCTION

The almost inevitable shift from the estrogen-dependent growth character of early stage breast cancer to an estrogen-independent growth phenotype is correlated with increased tumor aggressiveness and metastatic potential and decreased patient survival. Few models exist for the study of this transition. We have reported previously that long-term steroid-deprivation of the hormone-dependent MCF-7 breast cancer cell line resulted in the ability of these cells to form estrogen-independent tumors in nude mice and the acquisition of a more invasive phenotype in a subline selected *in vivo* (1). Thus,

long-term steroid-deprived MCF-7 sublines appear to be appropriate models for the study of breast cancer progression.

In the present study, we expand our investigation of this model to characterize the role of TGF- α and the TGF- β s during the time period in which MCF-7 cells progress to an estrogen-autonomous growth state. It has been hypothesized that growth factor autocrine loops may have supplanted estrogen-mediated mitogenic pathways in more progressed breast tumors (2). The known mitogenic and angiogenic regulating properties of the TGFs make them good candidates in this capacity.

Experiments using exogenous TGF- α or anti-TGF- α antibodies have demonstrated that TGF- α is a growth-promoting autocrine regulator in a number of cancer cell lines (2, 3). Furthermore, TGF- α mRNA levels have been found to be elevated in estrogen-independent, as compared to estrogen-dependent, human breast cancer cell lines (4), and overexpression of TGF- α has been shown to function as a transforming oncogene in cell-specific (5, 6) and pregnancy-specific (7, 8) contexts. Since the expression of this mRNA is modulated by ligand-mediated up-regulation and by phorbol esters (9, 10), as well as by the estrogen receptor (4, 11), it responds to steroid-independent as well as steroid-dependent signaling pathways. Thus, TGF- α may modulate cell proliferation in the absence of estrogen-regulated growth as well as in its presence.

The TGF- β s have been most frequently identified as negative autocrine regulators in breast neoplasms and cell lines, including the MCF-7 cell line (12, 13). The TGF- β s have been shown to be elevated in more aggressive cancers (14, 15), and Arteaga *et al.* (16) recently reported that introduction of TGF- β 1 cDNA into MCF-7 cells resulted in the formation of estrogen-independent tumors in nude mice. Like TGF- α , TGF- β expression has been found to be regulated by steroidal (17) as well as nonsteroidal (18) mechanisms. This finding, in combination with the known growth-regulating potential of these factors and their often aberrant production in breast cancers, suggest that these peptides might function as possible mediators of events that accompany the progression from estrogen-responsive to estrogen-autonomous growth in breast cancer. Our goal in this work was to evaluate alterations in TGF- α and TGF- β expression and in mitogenic sensitivity to these factors during the progression of MCF-7 cells to estrogen-autonomous growth.

MATERIALS AND METHODS

Materials. Radioinert estradiol and R5020 (promegestone; 17,21-dimethyl-19-nor-pregna-4,9-diene-3,20-dione), bovine insulin, hydrocortisone, nutritional supplements for growth in serum-free conditions, protease inhibitors, and sera were purchased from Sigma Chemical Co. (St. Louis, MO). *trans*-Hydroxytamoxifen and ICI 164,384 were generously provided by ICI Pharmaceuticals (Macclesfield, England). Tissue culture media and antibiotics

Received 6/30/94; accepted 9/29/94.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

¹ Supported in part by NIH Grants CA18119 and CA51482 (to B. S. K.), predoctoral fellowship GM07283, and Carle Foundation Cancer Research Funds (to M. E. H.), and a Susan G. Komen Foundation grant.

² To whom requests for reprints should be addressed, at Department of Physiology and Biophysics, University of Illinois, 524 Burrill Hall, 407 S. Goodwin Ave., Urbana, IL 61801.

³ The abbreviations used are: TGF, transforming growth factor; cDNA, complementary DNA; ER, estrogen receptor; MEM, phenol red-containing Eagle's minimal essential medium; IMEM, phenol red-free Richter's improved minimal essential medium; PgR, progesterone receptor; FCS, fetal calf serum; CDFCS, charcoal-dextran-treated FCS; BSA, bovine serum albumin; HBSS, Hanks' balanced salt solution; CAT, chloramphenicol acetyltransferase; E₂, estradiol; EGF, epidermal growth factor.

were purchased from GIBCO (Grand Island, NY). Tritiated estradiol (2,4,6,7- ^3H]N-estradiol), tritiated R5020 (17 α -methyl- ^3H]promegestone), and [^{125}I]TGF- β 1 were purchased from New England Nuclear Corp. (Boston, MA), and [methyl- ^3H]thymidine was purchased from ICN (Costa Mesa, CA). The ER-specific antibody H222 was generously provided by Geoffrey L. Greene, University of Chicago. Activated human recombinant TGF- β 1 was purchased from R&D Systems (Minneapolis, MN), and TGF- α was purchased from InterGen Co. (Purchase, NY) and United States Biochemical Corp. (Cleveland, OH). Anti-TGF- α antibody was purchased from Oncogene Science (Uniondale, NY); control antibody was purchased from Zymed Immunologicals (South San Francisco, CA). RNase protection assay supplies and protocols were purchased from Ambion Corp. (Austin, TX). TGF- α , - β 1, - β 2, and - β 3 cDNAs and gene constructs were generously provided by Rik Derynck (Genentech, Inc., South San Francisco, CA).

Cell Culture. MCF-7 human breast cancer cells were acquired from the Michigan Cancer Foundation; cells between passage numbers 150–300 were used in these studies. Parent MCF-7 cells were routinely cultured in MEM supplemented with 5% heat-inactivated FCS, 10^{-12} M estradiol, 10 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid buffer, 6 ng/ml insulin, 100 units/ml penicillin, 100 $\mu\text{g}/\text{ml}$ streptomycin, and 50 $\mu\text{g}/\text{ml}$ gentamicin. To generate long-term steroid-deprived MCF-7 sublines, cells were switched from standard estrogenic conditions to 5% CDFCS in MEM containing phenol red. After 2 weeks, the sublines were switched to estrogen-free growth media containing 5% CDFCS in IMEM. Steroid-deprived sublines were thereafter routinely maintained in this growth medium. Cells were subcultured weekly at near confluence using 1 mM EDTA prepared in HBSS, and media were replenished every other day.

Cell Proliferation Studies. Steroid-deprived cells prepared as described above were grown in 5% CDFCS IMEM without insulin for 5 days prior to the experiment. Cells were seeded at 150,000 cells/T25 flask; after 2 days, Day 0 flasks were counted, and the media in triplicate flasks were replaced with 7.5% CDFCS IMEM and treatments. Media were changed every 2 days, and cells in logarithmic phase were harvested on Day 6 and counted in a Coulter particle counter (Hialeah, FL).

Whole-Cell Hormone Binding Assays. Whole-cell ER and PgR binding assays were modified from that of MacIndoe *et al.* (19) as described previously (20). Cells were grown in 5% CDFCS IMEM without insulin and, for PgR studies, with and without estradiol as indicated. One million cells/tube were incubated with 10 nM [^3H]estradiol or [^3H]R5020 in the absence or presence of a 100-fold excess of unlabeled ligand and for PgR studies, with 3.75 ng/ml hydrocortisone. After incubating at 37°C for 30 min, cells were washed twice with 1% Tween-80 in phosphate-buffered saline, and bound radiolabeled ligand was counted in a scintillation counter.

Western Blot Analysis. Cells at subconfluence were harvested, pelleted, and resuspended in 50 mM Tris (pH 7.4), 7.5 mM EDTA, 0.6 M NaCl, 10% glycerol, and 20 mM sodium molybdate in the presence of proteinase inhibitors (0.5 mg/ml leupeptin, 50 mg/ml pepstatin A, and 1 mM phenylmethylsulfonylfluoride) and homogenized on ice. Samples were centrifuged for 25 min at 46,000 rpm, and the protein content in the supernatants was determined by BCA assay (Pierce Chemical Co., Rockford, IL). Samples (150 μg) were boiled for 5 min in loading buffer, separated on an sodium dodecyl sulfate polyacrylamide gel and transferred to nitrocellulose. Blots were incubated sequentially with ER-specific antibody H222, a bridging rabbit anti-rat IgG, and finally with [^{125}I]protein A and then exposed to film (21).

[^3H]Thymidine Incorporation Studies. Cells were seeded at 2000 cells/well in 24-well dishes. Two or three days later, the wells were washed in serum-free media for 2 h and then treated in serum-supplemented or in serum-free IMEM with 1 $\mu\text{g}/\text{ml}$ fibronectin, 2 $\mu\text{g}/\text{ml}$ transferrin, and 1:100 dilution of trace elements. After 3 or 4 days without media changes, the cells were incubated with 0.5 mCi methyl [^3H]thymidine at 37°C for 2 h. Plates were sequentially washed and fixed with ice-cold phosphate-buffered saline, 10% trichloroacetic acid (2X), and methanol, and incorporated label was then recovered by incubation of the wells in 0.5 N NaOH for 30 min at 37°C. Lysates were transferred to vials containing ScintiVerse cocktail (Fisher Scientific), and [^3H]thymidine radioactivity was determined in a scintillation counter.

TGF- α and TGF- β Protein Determinations. Subconfluent layers in T150 flasks were washed three times for 1 h in serum-free media and then incubated in serum-free media supplemented with 2 $\mu\text{g}/\text{ml}$ transferrin, 1 $\mu\text{g}/\text{ml}$

fibronectin, and 1:100 trace elements for 48 h. For the TGF- β assay, BSA was added to a final concentration of 0.5 mg/ml conditioned media, and the samples were snap frozen and later tested for the ability to inhibit [^3H]thymidine incorporation by Mv 1 Lu (mink lung) epithelial cells. Active and total TGF- β bioactivity was kindly determined by Anita Roberts and Nan Roche (National Cancer Institute, Bethesda, MD). For the TGF- α radioimmunoassay, conditioned media was collected, proteinase inhibitors (leupeptin, PMSF, aprotinin, and pepstatin) were added, and samples were dialyzed three times against distilled water over a period of 2 days at 4°C. Samples were then lyophilized, and resuspensions were assayed in a TGF- α radioimmunoassay kit (Biomedical Technologies, Inc., Stoughton, MA) as directed.

Isolation of mRNA. Isolation of total RNA from near confluent monolayers of cells was performed using guanidinium thiocyanate-phenol-chloroform extraction according to the method of Chomczynski and Sacchi (22) with some modifications (21). RNA concentration and purity was determined by absorbance at 260 and 280 nm.

Ribonuclease Protection Assay. Ten to 20 μg of RNA was coprecipitated with *in vitro*-transcribed, gel-purified complementary RNA labeled with phosphorus-32 and resuspended in 80% formamide/0.1 M sodium citrate (pH 6.4)/0.3 M NaOAc (pH 6.4)/1 mM EDTA. Samples were heated to 85°C for 5 min and hybridized overnight at 45°C. Unhybridized total RNA and probe was digested in a final concentration of 5 units/ml RNase A and 1000 units/ml RNase T1 for 30 min at 37°C. The sizes of protected fragments were confirmed by comparison to a lane loaded with a 0.16–1.77 kilobases RNA ladder (GIBCO). The probes used were a 168-base pair *Nco*I segment of TGF- α cDNA (23), a 240-base pair *Mbo*II segment of TGF- β 1 cDNA (24), a *Hpa*I segment of TGF- β 2/sp72 cDNA (25), and a 125-base pair *Nde*I segment of TGF- β 3 cDNA (26). A 99-base pair *Mbo*II segment or 68-base pair *Sfa*NI segment of human acidic phosphoprotein PO (36B4; Ref. 27) was used as an internal control as described previously (28). We did not find any changes in the level of expression of 36B4 mRNA in the sublines used in this paper.

[^{125}I]TGF- β 1 Binding Assay. Cells at $3.0\text{--}3.6 \times 10^5$ cells/well in 24-well plates (75–90% confluent) were washed three times over 1 h with serum-free media supplemented with 0.1% BSA and were then incubated with 10^{-10} M [^{125}I]TGF- β 1, with and without a 100-fold excess of radioinert TGF- β 1, for 45 min. Cells were then washed four times with ice-cold HBSS (with 0.1% BSA) and solubilized with 1% Triton X-100, 20 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (pH 7.4), 10% glycerol, and 0.01% BSA for 15 min at 37°C. Solubilized fractions were counted in a gamma counter.

Transient Transfections and CAT Assays. Two days after plating at 3×10^6 cells per 100-mm dish, cells were transfected by the calcium phosphate precipitation method (29). Three μg of a construct containing the consensus estrogen response element linked to a thymidine kinase promoter and the CAT gene (ERE-TK-CAT) were cotransfected with 0.5 μg of the internal control plasmid, CMV- β -gal, and 11.5 μg of the carrier DNA, pTZ19. After 6 h incubation at 37°C, cells were incubated for 3 min with 20% glycerol in 2% CDFCS IMEM, washed with HBSS, and then treated for 24 h in fresh media. Cells were harvested in cold 40 mM Tris (pH 7.5), 140 mM NaCl, and 1.5 mM EDTA and centrifuged; then cell extracts were prepared by three quick freeze-thaw cycles in 250 mM Tris (pH 7.5). The mixed-phase CAT assay was performed as described previously (30). Fold inductions within each assay were normalized against β -galactosidase activity (31, 32).

RESULTS

Generation and Steroid Responsiveness of Long-term, Steroid-deprived MCF-7 Sublines. In order to monitor the progression of the MCF-7 cells to an estrogen-autonomous growth state, we transferred MCF-7 parent cells from estrogen-supplemented medium containing phenol red and FCS to steroid-depleted, phenol red-free medium containing CDFCS. The growth rate slowed from doubling times of 1.4 to 4.0 days at 2 weeks of steroid deprivation. Cells showed even slower doubling rates between 3 and 8 weeks (data not shown) and subsequently regained a rapid growth rate of 1.6 days by 13 weeks (Fig. 1A). This rate was observed reproducibly and found to be maintained during at least 150 weeks of steroid deprivation.

The growth rate of short-term (2-week), steroid-deprived cells was dramatically increased by treatment with estrogen (to 1.1 *versus* 4.0

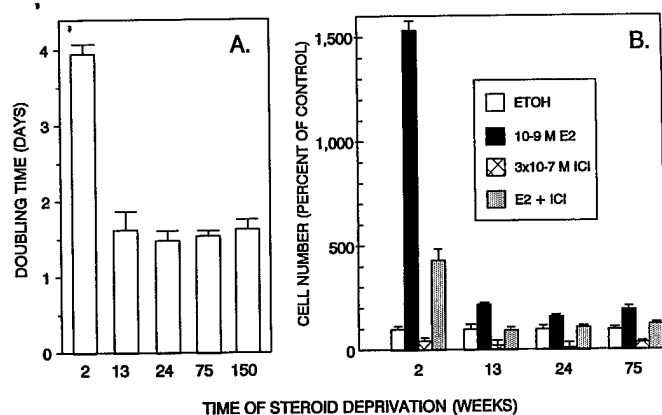


Fig. 1. A, basal growth rates of steroid-deprived MCF-7 cells. Values were determined from triplicate subconfluent flasks of cells and are expressed as doubling times in days. B, growth responsiveness of steroid-deprived MCF-7 cells to estrogen and antiestrogen. Cells were grown in steroid-deprived cell culture medium for the weeks indicated and were then treated with control vehicle (0.1% ethanol), 10^{-9} M estradiol (E₂), or 3×10^{-7} M ICI 164,384 (ICI), alone or in combination. Cell number was determined on Day 6 of treatment; values are expressed as a percentage of control cell number/flask. In (A) and (B), values are the mean of three determinations and are representative of at least three experiments; bars, SEM.

Table 1 ER content of MCF-7 cells maintained for different periods of time in steroid-deprived cell culture medium

Receptor content was determined by whole-cell hormone binding assay ($n = 3$; mean \pm SEM) or Western blot analysis of immunoreactive ER in fractionated cellular protein detected by binding of the ER-specific monoclonal antibody H222 (mean and range of two separate experiments).

Time of steroid deprivation (wks)	[³ H]E ₂ -bound/ 10^6 cells (fmol)	Immunoreactive ER (%)
2	59.7 \pm 4.6	100 \pm 11
8	105.7 \pm 2.2	165 \pm 7
24	183.6 \pm 11.1	ND ^a
75	163.4 \pm 4.3	315 \pm 43

^a ND, not determined.

day doubling time; Fig. 1B). In contrast, estrogen treatment of the 13-, 24-, or 75-week steroid-withdrawn cells stimulated only modest ($\sim 200\%$) increases in cell number. The minimal stimulation of the growth of long-term, steroid-deprived sublines by estradiol is presumably due to the near-maximal rate of cell division obtained by these cells under steroid-deprived conditions. The growth-inhibitory effect of antiestrogen was evident, however, at all stages of steroid deprivation. The antiestrogen, ICI 164,384 (ICI), suppressed markedly the growth rate of all sublines and was able to substantially reverse the growth stimulatory effect of the estradiol treatment.

Maintenance of Functional Estrogen Receptor and Markers of Estrogen Responsiveness in Steroid-withdrawn MCF-7 Cells. ER levels of 60 fmol/ 10^6 cells at 2 weeks of steroid deprivation increased 177% at 8 weeks and approximately 300% at 21 and 75 weeks (184 and 163 fmol/ 10^6 cells, respectively) of steroid withdrawal (Table 1). This increase in ER level determined by hormone binding was confirmed by Western blot analysis (Table 1). We have reported previously that the increase in ER levels in response to long-term estrogen deprivation was not accompanied by alterations in ER binding affinity or specificity (1, 20).

Two markers of estrogen responsiveness were used in order to determine whether ER remained functional in steroid-deprived MCF-7 cells. First, we monitored induction of PgR by estrogen (10^{-14} M- 10^{-10} M E₂) and observed similar dose-response curves and comparable levels of estradiol-induced PgR in short- and long-term, steroid-deprived cells (Fig. 2). Second, we studied the ability of estrogen (10^{-13} M- 10^{-7} M E₂) to induce CAT activity from an estrogen response element-containing reporter gene construct (ERE-

tk-CAT; Table 2). Both short- and long-term, steroid-deprived cells showed similar dose-responses and magnitudes of response. The antiestrogens ICI 164,384 and LY 117,018 (10^{-6} M) evoked little if any reporter gene activity in the 2-week steroid-deprived MCF-7 cells, as expected, as well as in the 100-week steroid-deprived MCF-7 cells (Table 2). The partial agonist, *trans*-hydroxytamoxifen (10^{-6} M), showed weak stimulation of CAT activity in both short- and long-term steroid-deprived MCF-7 cells. The three antiestrogens substantially reduced the stimulation by E₂ (Table 2).

Expression of TGF- α in MCF-7 Cells in Response to Steroid Withdrawal. To characterize the potential role of TGF- α in cell progression, expression of TGF- α mRNA in short- and long-term steroid-deprived MCF-7 cells was evaluated. TGF- α mRNA levels decreased by 66% in response to growth in short-term (2-week), steroid-depleted conditions (Fig. 3A). TGF- α mRNA levels then

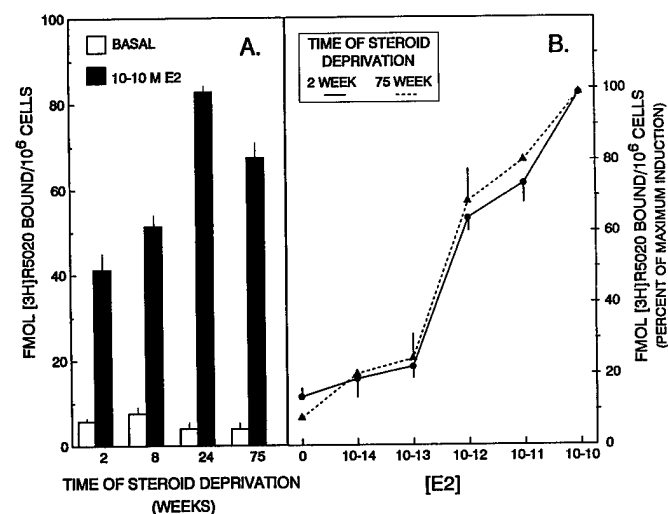


Fig. 2. Induction of PgR content in parental and steroid-deprived MCF-7 cells as determined by whole-cell hormone binding assay. A, MCF-7 cells steroid-deprived for 0, 2, 8, 24, or 75 weeks were treated with 10^{-10} M E₂ for 4 days. B, MCF-7 cells steroid-deprived for 2 or 75 weeks were treated with ethanol vehicle or E₂ at the concentrations indicated for 4 days. Values in (A) and (B) are the mean of triplicate flasks from two experiments. Bars, SEM.

Table 2 Transactivation of ERE-tk-CAT, a reporter plasmid containing a consensus estrogen response element linked to the Herpes simplex virus thymidine kinase promoter and the CAT reporter gene

ERE-tk-CAT was cotransfected into 2- or 100-week steroid-deprived MCF-7 cells, along with an internal control plasmid containing the lac-Z gene; and cells were treated with the ligands indicated for 24 h. The calculated fold increase in the CAT activity of each group was normalized for the β -galactosidase activity. Values are the mean \pm SEM of at least three experiments.

Treatment	Fold increase in CAT activity (control = 1)	
	Time of steroid deprivation	
	2 weeks	100 weeks
10^{-13} M E ₂	7.9 \pm 0.5	4.9 \pm 3.0
10^{-11} M E ₂	13.2 \pm 3.1	11.3 \pm 0.3
10^{-10} M E ₂	40.6 \pm 2.4	36.6 \pm 5.4
10^{-9} M E ₂	52.1 \pm 1.7	47.0 \pm 2.0
10^{-7} M E ₂	53.5 \pm 2.0	58.1 \pm 3.2
10^{-6} M ICI 164,384	0.9 \pm 0.1	1.8 \pm 0.6
10^{-6} M LY 117,018	2.6 \pm 1.0	5.0 \pm 1.0
10^{-6} M TOT ^a	6.1 \pm 1.7	15.8 \pm 5.1
10^{-9} M E ₂ + 10^{-6} ICI	13.0 \pm 1.0	20.5 \pm 5.5
10^{-9} M E ₂ + 10^{-6} LY	14.0 \pm 1.0	13.2 \pm 5.4
10^{-9} M E ₂ + 10^{-6} TOT	9.8 \pm 0.1	23.9 \pm 11.0

^a TOT, *trans*-hydroxytamoxifen; ICI, ICI 164,384; LY, LY 117,018.

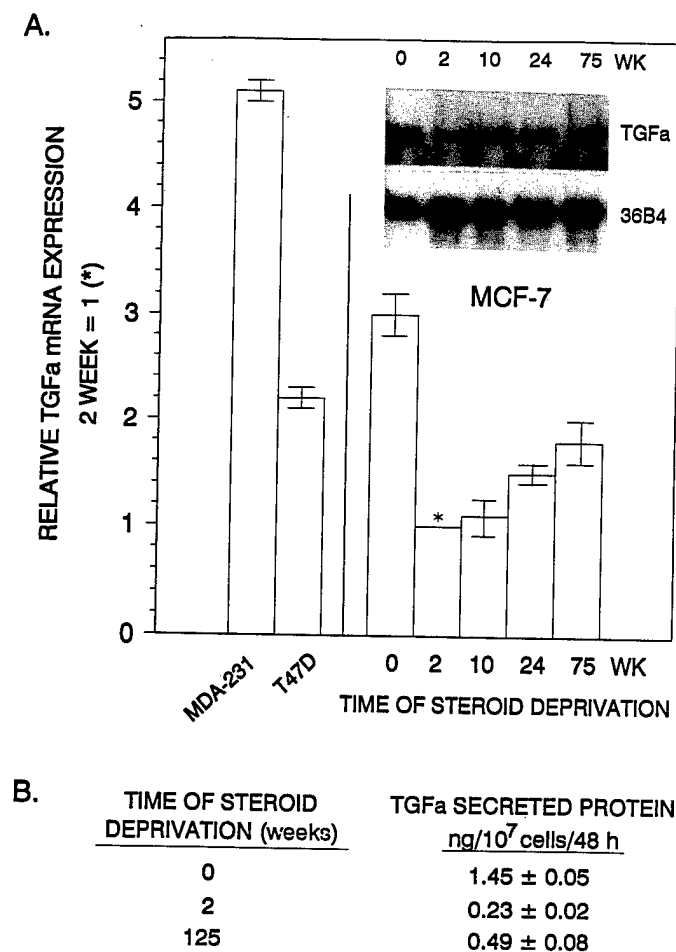


Fig. 3. A, expression of TGF- α in MCF-7 cells in response to steroid deprivation. RNA was isolated from parent MCF-7 cells (Time 0) and from steroid-withdrawn sublines, all grown to subconfluence. Ten to 20 μ g of total RNA were cohybridized with the probes for TGF- α and 36B4 in a RNase protection assay. Autoradiograms at several exposures were quantified on a densitometer and normalized against the internal control, 36B4; shown is one such autoradiogram. Values obtained were the mean from three experiments; bars, SEM. The expression of TGF- α mRNA in 2-week steroid-deprived MCF-7 cells is arbitrarily designated as having a value of 1.0 (*). The relative expression of TGF- α mRNA in 2-, 10-, 24-, and 75-week steroid-deprived cells differed significantly from the 0-week steroid-deprived cells ($P < 0.05$); only the relative expression of TGF- α mRNA in 75-week steroid-deprived cells differed significantly from the 2-week steroid-deprived cells at $P < 0.05$. For comparison, the levels of TGF- α mRNA in MDA-231 and T47D breast cancer cells are shown. B, conditioned media were collected over a period of 48 h, dialyzed, and lyophilized; and resuspensions were assayed for TGF- α protein in a radioimmunoassay. Values represent the mean \pm SEM.

progressively increased over time. At 75 weeks of steroid deprivation, the level of TGF- α mRNA expression was approximately 2-fold higher than that in the short-term, steroid-deprived cells but was still significantly below that of the parental MCF-7 cells. These alterations in TGF- α mRNA levels were closely mirrored by the amount of TGF- α secreted protein produced by these cells, i.e., an early marked decrease (from 1.45 \pm 0.05 ng/10⁷ cells/48 h to 0.23 \pm 0.02 ng/10⁷ cells/48 h) followed by a partial restoration (0.49 \pm 0.08 ng/10⁷ cells/48 h) of the TGF- α secretion rate of the steroid-maintained parental MCF-7 cells (Fig. 3B).

We also examined the magnitude of up-regulation of TGF- α mRNA by estradiol and its suppression by the antiestrogen, ICI 164,384 after brief (4-day) exposure to these agents in 2-, 10-, and 75-week steroid-deprived cells. Dramatic changes in the level of TGF- α mRNA by these agents were not found over the 4-day treatment period, but the modest modulation by estradiol and antiestrogen was similar in the 2-, 10-, and 75-week steroid-deprived cells (Fig. 4). E₂ exposure stimulated 120 to 145% increases in TGF- α mRNA

(which correlated closely with secreted TGF- α protein levels, in conditioned media; data not shown), and antiestrogen treatment reduced both the control and the E₂-stimulated TGF- α mRNA level in the three cell sublines, indicating little change in these cells in short-term estrogen and antiestrogen regulation of TGF- α over the 75 weeks of steroid deprivation.

Mitogenic Response of Steroid-deprived MCF-7 Cells to Exogenous TGF- α , Anti-TGF- α Antibodies, or Estrogen. As shown in Table 3 (Lines 1-3), exogenous TGF- α induced dose-dependent increases in [³H]thymidine incorporation in 2-week steroid-deprived MCF-7 cells; a maximal stimulation of 248 \pm 5% of control values was observed. Correspondingly, anti-TGF- α antibodies (which do not cross-react with EGF) induced dose-dependent decreases in [³H]thymidine incorporation in the 2-week steroid-deprived cells (Table 3, Lines 4-7). Cotreatment with TGF- α and anti-TGF- α antibody (Table 3, Line 8) demonstrated that the antibody was able to partially block the TGF- α -stimulated increase in [³H]thymidine incorporation. Treatment with E₂ (Table 3, Line 9) stimulated a marked increase in [³H]thymidine incorporation (2512 \pm 9% of control) in the 2-week steroid-deprived cells.

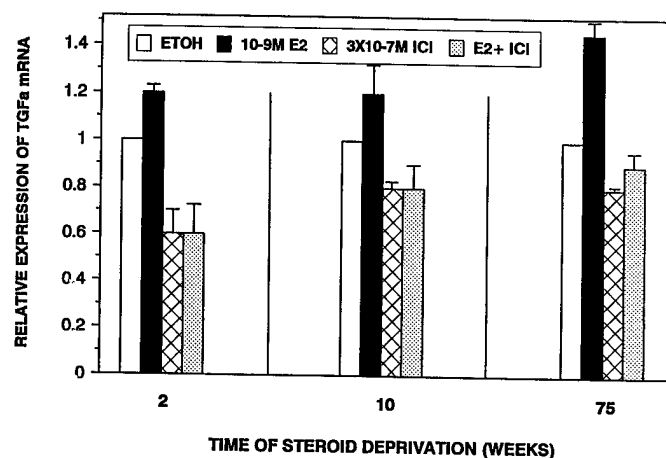


Fig. 4. Regulation of TGF- α mRNA by estrogen and antiestrogen in steroid-deprived MCF-7 cells. Subconfluent cultures were grown in 5% CDFCS IMEM medium without phenol red and were treated with the compounds indicated for 4 days. Details of the methods are described in the legend for Fig. 3A and in "Materials and Methods." Values represent the mean and range of two determinations.

Table 3 [³H]Thymidine incorporation in steroid-deprived MCF-7 cells in response to E₂, exogenous TGF- α , or anti-TGF- α antibodies

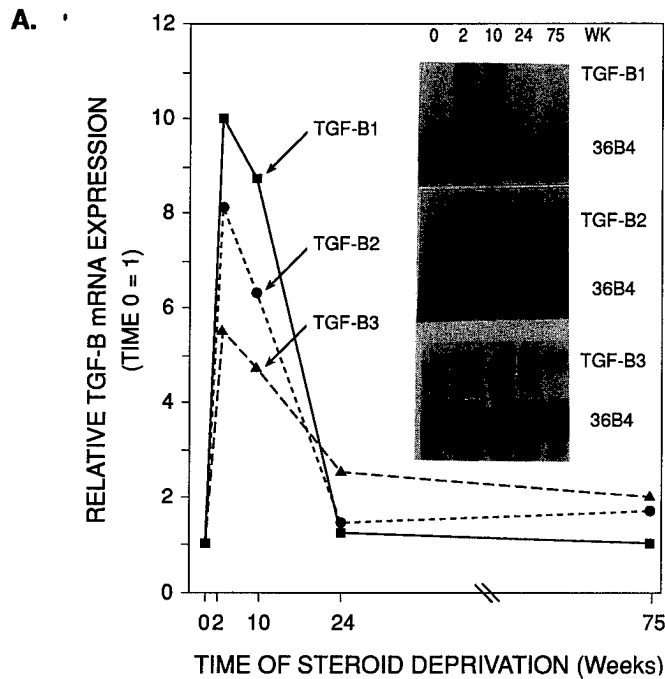
Two days after seeding the cells at 2000 cells/well in 24-well dishes, treatments were added in serum-free IMEM. On Day 4, without media changes, the cells were incubated with 0.5 mCi [³H]thymidine for 2 h and processed as described in "Materials and Methods."

Treatment			[³ H]Thymidine incorporation (% of control) ^a	
			Time of steroid deprivation	
TGF- α	Anti-TGF- α	E ₂	2 Weeks	75 Weeks
(1) 10 ng/ml			121 \pm 4 ^b	98 \pm 2
(2) 25			133 \pm 8 ^b	116 \pm 4
(3) 100			248 \pm 5 ^b	104 \pm 3
(4)	10 ng/ml		93 \pm 4	95 \pm 5
(5)	50		68 \pm 3 ^b	92 \pm 5
(6)	150		52 \pm 2 ^b	98 \pm 3
(7)	1000		ND ^c	91 \pm 11
(8) 100	100		169 \pm 2 ^b	104 \pm 6
(9)		10 ⁻⁹ M	2512 \pm 9 ^b	366 \pm 9 ^b
(10)	100	10 ⁻⁹ M	2388 \pm 6 ^b	447 \pm 16 ^b

^a n = 4; mean \pm SEM.

^b Values significantly different from the control at $P < 0.05$ by Student's *t* test.

^c ND, not determined.



TIME OF STEROID DEPRIVATION (Weeks)	TGF- β BIOACTIVITY	
	pg/10 ⁶ cells/48 h	% Active
0	51 ± 10	30 ± 13
2	153 ± 31	43 ± 5
125	54 ± 3	9 ± 2

Fig. 5. A, basal expression of TGF- β 1 (■), TGF- β 2 (●), and TGF- β 3 (▲) mRNA in steroid-deprived MCF-7 cells. RNA was isolated and cohybridized with TGF- β 1, - β 2, or - β 3 cDNA probe and with 36B4 internal control cDNA probe. RNase protection assays were quantitated by densitometric analyses of autoradiograms as described in "Materials and Methods." Values represent the average of two experiments. Shown is the autoradiogram of one such analysis. B, total and percent active TGF- β protein in duplicate conditioned media collections from two separate experiments were determined by inhibition of [³H]thymidine incorporation by Mv 1 Lu cells. Values represent the mean and range from the two separate experiments.

In comparison to the short-term (2-week), steroid-deprived cells, [³H]thymidine incorporation in long-term (75-week), steroid-deprived cells was not significantly affected by either TGF- α or anti-TGF- α antibodies (Table 3, Lines 1–8), and E₂ evoked a much more modest stimulation (366 ± 9%; Table 3, Line 9). Coincubation with anti-TGF- α antibody and E₂ (Table 3, Line 10) failed to inhibit the estrogen-induced increases in [³H]thymidine incorporation in either short- or long-term, steroid-deprived cells, suggesting that estrogen did not use the TGF- α pathway for its mitogenic effect in these cells.

Expression of TGF- β 1, - β 2, and - β 3 in MCF-7 Cells in Response to Steroid Withdrawal. We analyzed expression of TGF- β mRNAs and TGF- β bioactive protein and found the temporal patterns in response to growth in steroid-depleted conditions to be the inverse of the pattern we observed for TGF- α . At 2 weeks of steroid withdrawal, the mRNA for all three TGF- β s increased 5- to 10-fold from the low basal levels observed for parent cells cultured in estrogen-containing conditions (Fig. 5A). The levels of TGF- β 1, - β 2, and - β 3 mRNA remained high at 10 weeks of steroid deprivation but, by 24 weeks, the levels of these three mRNAs returned to the low levels observed in the parental (0-week) cells.

The increased levels of TGF- β mRNAs in response to short-term (2-week), steroid deprivation were reflected in 3-fold increases in TGF- β protein (from 51 ± 10 pg/10⁶ cells/48 h to 153 ± 31 pg/10⁶

cells/48 h; Fig. 5B). The reestablishment of low basal levels of TGF- β mRNAs at later times (≥24 weeks) was reflected in low secreted TGF- β protein levels (54 ± 3 pg/10⁶ cells/48 h in 125-week steroid-deprived cells) similar to levels observed in parental cells (Fig. 5B). No dramatic change was observed in the proportion of TGF- β in the active *versus* latent form in long-term, steroid-deprived MCF-7 cells as compared to the parent MCF-7 cells. Since the data in Fig. 6 (see below) show that the TGF- β s are negatively regulated by estrogen in these cells, it seems plausible that short-term growth in steroid-depleted serum released the TGF- β genes from suppression by estrogen, and the corresponding sharp rise of these TGF- β mRNAs and proteins contributed to the slow growth of these cells observed during the early time period after estrogen withdrawal.

We found, as shown in Fig. 6, that 2- and 75-week steroid-deprived cells showed generally similar patterns of response to brief (4-day) exposure to estradiol and/or antiestrogen (ICI). Thus, E₂ decreased and ICI increased the levels of TGF- β 1, - β 2, and - β 3 mRNA. The only difference between 2- and 75-week cells was in the more marked suppression of TGF- β 2 mRNA by E₂ in the 2-week steroid-deprived cells. It is important to note, however, that the control levels of the TGF- β mRNAs are 5- to 10-fold higher in the 2-week steroid-deprived cells, as reported in Fig. 5.

Growth Inhibition of Short- and Long-term, Steroid-deprived MCF-7 Cells by TGF- β 1. As shown in Fig. 7, 2- and 12-week steroid-deprived cells were highly sensitive to the growth inhibitory effect of TGF- β 1; near-maximal inhibition of [³H]thymidine incorporation was achieved with a low concentration (1–5 ng/ml) of TGF- β 1. Increasing concentrations of TGF- β 1 further decreased [³H]thymidine incorporation to a maximal suppression that was approximately 35–40% of control values. Interestingly, the long-term (140-week) steroid-deprived subline was substantially less sensitive to the growth inhibitory effects of low concentrations of TGF- β 1. In contrast to an approximate 50% inhibition of [³H]thymidine incorporation in short-term, steroid-deprived cells by 1 ng/ml TGF- β 1, cells which had been steroid-deprived for 140 weeks exhibited only 10% growth inhibition. An equivalent 50% growth inhibition in the long-term steroid-deprived subline required treatment with 10 ng/ml TGF- β 1, corresponding to an approximate 10-fold shift in the dose-response curve.

To examine the possibility that the long-term steroid-deprived cells lost sensitivity to TGF- β 1 due to changes in the content of TGF- β receptors, we performed binding studies with [¹²⁵I]TGF- β 1. We did not find a change in the binding capacity of short- and long-term, steroid-deprived MCF-7 cells (Table 4).

DISCUSSION

Several groups, including our own, have generated sublines of ER-positive human breast cancer cells by maintenance in steroid-depleted growth conditions both *in vivo* and *in vitro* in order to study the development of hormone-autonomous growth. These sublines have been characterized in terms of steroid-responsive growth, altered gene expression, and tumorigenic potential (1, 20, 28, 33–35). Little attention, however, has been focused on the time period during which the sublines accomplish this transition, including changes in growth factor production and sensitivity that may supplant steroid-dependent pathways. Consequently, we turned our attention to the role of potentially important growth factor mediators, namely TGF- α and the TGF- β s, during the progression to steroid-autonomous growth in MCF-7 cells.

Our studies reveal substantial changes in growth rates and in production of, and responsiveness to, TGF- α and TGF- β within 3 to 4 months of steroid deprivation. This phenotype was maintained for

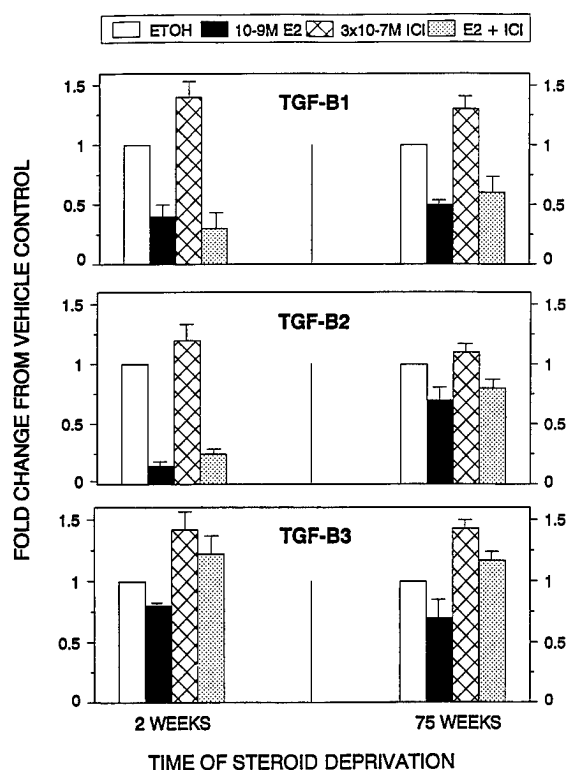


Fig. 6. Regulation of TGF- β 1, - β 2, and - β 3 mRNA by estrogen and anti-estrogen in short-term (2-week) and long-term (75-week) steroid-deprived MCF-7 cells. Hormone treatments were for 4 days. RNA was isolated and probed for TGF- β 1, - β 2, or - β 3 mRNA as described in "Materials and Methods." Autoradiograms were normalized for 36B4 RNA content and quantitated. The level of each TGF- β mRNA in the vehicle control-treated cells is arbitrarily designated as having a relative expression of 1.0. Values represent the mean and range from two experiments.

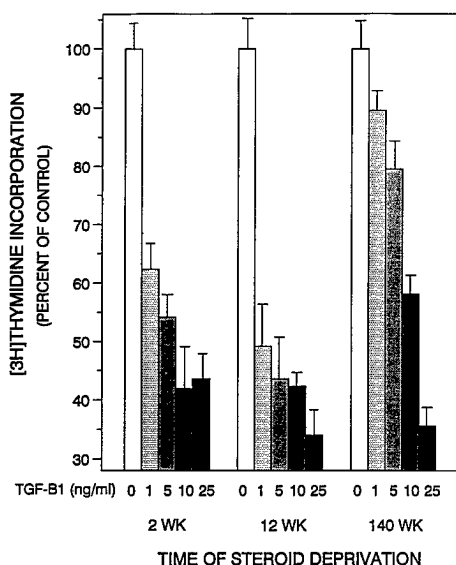


Fig. 7. Inhibition of [3 H]thymidine incorporation by TGF- β 1 in steroid-deprived MCF-7 cells. Cells were seeded in triplicate at 2000 cells/well in a 24-well plate. After 3 days, the medium (5% CDFCS IMEM) was replenished, and the indicated concentrations of TGF- β 1 were added. [3 H]Thymidine incorporation was measured during a 2-h incubation on Day 3 of treatment with TGF- β 1. Values represent the mean from three experiments; bars, SEM.

periods of at least 3 years of steroid deprivation, the longest time we studied. These long-term, steroid-deprived cells contain high levels of ER and show good estrogen and antiestrogen responsiveness, as monitored by PgR regulation and transcriptional activation of an

estrogen responsive element-containing reporter gene. Thus, estrogen-autonomous growth occurred in the presence of significant levels of functional estrogen receptor. Daly and Darbre (34) also reported 2- to 3-fold increases in ER levels and the maintenance of PgR and *pS2* gene responsiveness to estrogen in long-term, steroid-deprived ZR-75-1 and T47D human breast cancer cells. Apparently because our long-term, steroid-deprived MCF-7 cells exhibit rapid rates of cell division in the absence of estrogen, estradiol stimulates only moderate increases in [3 H]thymidine incorporation or cell division rates, while suppression of proliferation by antiestrogen is maintained. Long-term androgen deprivation of LNCaP prostate cancer cells was reported to give rise to cells similar to our MCF-7 long-term, steroid-deprived cells in that they proliferate rapidly without additional androgen, and they exhibit increased androgen receptor levels (36). The elevated steroid hormone receptor levels observed in these studies are consistent with the known suppressive effects of estrogen and androgen on their own receptor levels (22, 28, 37, 38).

We observed the reestablishment of rapid growth rates in the absence of steroids by 13 weeks of steroid deprivation. This rather short time frame suggests that an adaptational or cell selection mechanism, rather than mutation of the estrogen receptor gene or another gene, is most likely involved. This view is also supported by the normal dose-response sensitivity to estradiol for stimulation of the PgR, an estrogen-induced protein, and for transcriptional activation of a transiently transfected estrogen-responsive gene (ERE-TK-CAT) in the parental and long-term steroid-deprived cells. In contrast to our findings showing a maintenance of target gene responsiveness to estrogen in steroid-deprived MCF-7 cells, a loss of androgen sensitivity was reported in Shionogi 115 mouse mammary tumor cells in response to androgen withdrawal (39, 40).

Estrogens are known to regulate the production of growth factors and growth factor receptors in breast cancer (41) and uterine cells (42, 43). We were eager, therefore, to investigate whether changes in the production of growth stimulatory and growth inhibitory factors might be associated with the progression of breast cancer cells from an estrogen-responsive growth state to one in which growth factor regulation would occur largely independent of the requirement for estrogen. Our findings in MCF-7 cells are consistent with this notion and highlight that such changes occur within only weeks of steroid deprivation, suggesting that cellular events may allow regulation of cell proliferation to bypass the requirement for estrogen. We observed that the levels of TGF- α and the TGF- β s changed, being initially decreased or increased, respectively, and then returned to near the parental level in rapidly proliferating, long-term, steroid-deprived sublines.

Despite the partial reestablishment of the parental pattern of TGF- α mRNA and protein expression in long-term, steroid-deprived MCF-7

Table 4 Binding of 125 I-TGF- β 1 to parent and long-term, steroid-deprived MCF-7 cells

Cells at 75–90% confluency in 24-well plates were washed three times over 1 h with serum-free media supplemented with 0.1% BSA and incubated with 10^{-10} M 125 I-TGF- β 1 with or without a 100-fold excess of radioinert TGF- β 1 for 45 min. Cells were then washed, and solubilized fractions were counted in a gamma counter. Binding of 125 I-TGF- β 1 in ER-negative MDA-MB-231 human breast cancer cells is shown for comparison. Values are the mean \pm SEM of at least three determinations.

Cell line	Weeks of steroid deprivation	125 I-TGF- β 1 binding sites/cell
MCF-7	0	282 \pm 30
	1	320 \pm 36
	5	271 \pm 58
	75	183 \pm 34
	135	362 \pm 9
MDA-MB-231		1554 \pm 110

cells, these sublines lost growth responsiveness to exogenous TGF- α and anti-TGF- α antibodies, as monitored by [3 H]thymidine incorporation. Therefore, the cell division rates in the long-term, steroid-deprived cells were likely being maintained largely by factors other than TGF- α or estrogen. We have shown previously that long-term, steroid-deprived MCF-7 cells, which have elevated ER levels, express somewhat lower levels (~60%) of EGF receptor relative to parent MCF-7 cells (1), consistent with the well-known inverse relationship between ER and EGF receptor levels (44, 45). However, the complete lack of response to TGF- α as a stimulator of DNA synthesis in long-term, steroid-deprived cells is unlikely to be due to this partial reduction in EGF receptor level and suggests alterations at a subsequent step in the TGF- α response pathway. Furthermore, the TGF- α /EGF receptor pathway has been implicated as a mediator of estrogen action (46, 47). Our results suggest that the loss of growth responsiveness to TGF- α in our cell system may be unrelated to ER action, since induction of PgR and activation of a transiently transfected estrogen-responsive gene by estradiol were completely normal and as observed in the parental MCF-7 cells. We also failed to observe a decrease in estrogen-stimulated [3 H]thymidine incorporation by cotreatment with anti-TGF- α antibodies. Arteaga *et al.* (48) reported that antibody blockade of the TGF- α /EGF receptor did not alter estrogen-induced growth in hormone-dependent MCF-7 cells, and Clarke *et al.* (49) found that stably transfected MCF-7 cells producing high levels of TGF- α remained estrogen-responsive in terms of growth, PgR induction, and the formation of tumors in nude mice.

One of the initial effects of steroid depletion may have been a release of TGF- β transcription from an estrogen-mediated suppression. High levels of secreted, bioactive TGF- β resulted (Fig. 5) and may have contributed to the slow growth observed in these cells during the period of adaptation to steroid-depleted conditions. Indeed, we found that short-term, steroid-deprived MCF-7 cells were highly sensitive to the growth-inhibitory effect of TGF- β 1 (Fig. 7). This hypothesis is in agreement with the finding that removal of estradiol implants from MCF-7 tumors in nude mice resulted in three-fold increases in the level of tumor TGF- β 1 mRNA (50) and that treatment with estrogens down-regulates TGF- β mRNAs (26, 51). Interestingly, antiestrogen-stimulated levels of TGF- β have been implicated as the mediators of tamoxifen-induced growth suppression in breast cancer cells (52).

Of note, the long-term steroid-deprived sublines exhibited a 10-fold reduced sensitivity to the growth-inhibitory effects of exogenous TGF- β 1. One of the mechanisms by which TGF- β 1 insensitivity can be accomplished is through decreased TGF- β receptor binding (53). However, we found that there were no substantial differences in the number of TGF- β 1 binding sites in the parent and long-term, steroid-deprived MCF-7 cells. While the cells may have alterations in binding affinities for TGF- β 2 or TGF- β 3 (which we did not test) or in TGF- β receptor homodimer or heterodimer species, it appears most likely that postreceptor aberrations in TGF- β signaling account for the reduced TGF- β 1 sensitivity we observed in the long-term, steroid-deprived MCF-7 cells.

Our findings suggest a role for TGF- α and the TGF- β s in the progression of MCF-7 breast cancer cells from estrogen-responsive to estrogen-autonomous growth. Changes in the production of and/or sensitivity to other growth factors may also be occurring. Adaptations to a low steroid environment, involving alterations in growth factor production such as we report herein, may allow ER-positive breast cancer cells to proliferate rapidly, bypassing the requirement for full estrogen stimulation via the ER.

ACKNOWLEDGMENTS

We gratefully acknowledge Rik Derynck of the University of California at San Francisco and Genentech for the TGF- α , - β 1, - β 2, and - β 3 cDNAs; Pierre Chambon of INSERM (Strasbourg, France) for the 36B4 cDNA; Alan Wakeling of ICI Pharmaceuticals for the anti-estrogen, ICI 164,384; Geoffrey L. Greene of the University of Chicago for the ER-specific antibody H222; and Mary Jane Norman for technical assistance. Special thanks to Anita Roberts and Nan Roche of the National Cancer Institute for performing the TGF- β bioassay.

REFERENCES

- Clarke, R., Brunner, N., Katzenellenbogen, B. S., Thompson, E. W., Norman, M. J., Koppi, C., Paik, S., Lippman, M. E., and Dickson, R. B. Progression of human breast cancer cells from hormone-dependent to hormone-independent growth both *in vitro* and *in vivo*. *Proc. Natl. Acad. Sci. USA*, 86: 3649-3653, 1989.
- Arteaga, C. L., and Osborne, C. K. Growth factors as mediators of estrogen/antiestrogen action in human breast cancer cells. In: M. E. Lippman and R. B. Dickson (eds.), *Regulatory Mechanisms in Breast Cancer*, p. 289. Boston: Kluwer Academic Press, 1991.
- Morishige, K., Kuriche, H., Amemiya, K., Fujita, Y., Yamamoto, T., Miyake, A., and Tanizawa, O. Evidence for the involvement of transforming growth factor α and epidermal growth factor receptor in autocrine growth mechanism in primary human ovarian cancer *in vitro*. *Cancer Res.*, 51: 5322-5328, 1991.
- Murphy, L. C., and Dotzlaw, H. Endogenous growth factor expression in T-47D, human breast cancer cells, associated with reduced sensitivity to antiproliferative effects of progestins and antiestrogens. *Cancer Res.*, 49: 599-604, 1989.
- Ciardiello, F., McGeady, M. L., Kim, N., Basolo, F., Hynes, N., Langton, B. C., Yokosaki, H., Saeki, T., Elliot, J. W., Masui, H., Mendelsohn, J., Soule, H., Russo, J., and Salomon, D. S. Transforming growth factor α expression is enhanced in human mammary epithelial cells transformed by an activated c-Ha-ras protooncogene but not by the c-neu protooncogene and overexpression of the transforming growth factor α cDNA leads to transformation. *Cell Growth & Differ.*, 1: 407-420, 1990.
- McGeady, M. L., Kerby, S., Shankar, V., Ciardiello, F., Salomon, D. S., and Seidman, M. Infection with a TGF α retroviral vector transforms normal mouse mammary epithelial cells but not normal rat fibroblasts. *Oncogene*, 4: 1375-1382, 1989.
- Sandgren, E. P., Luetke, N. C., Palmiter, R. D., Brinster, R. L., and Lee, D. C. Overexpression of TGF α in transgenic mice: induction of epithelial hyperplasia, pancreatic metaplasia, and carcinoma of the breast. *Cell*, 61: 1121-1135, 1990.
- Matsui, Y., Halter, S. A., Holt, J. T., Hogan, B. L. M., and Coffey, R. J. Development of mammary hyperplasia and neoplasia in MMTB-TGF α transgenic mice. *Cell*, 61: 1147-1155, 1990.
- Raymond, V. W., Lee, D. C., Grisham, J. W., and Earp, H. S. Regulation of transforming growth factor α messenger RNA expression in a chemically transformed rat hepatic epithelial cell line by phorbol ester and hormones. *Cancer Res.*, 49: 3608-3612, 1989.
- Bjorge, J. D., Paterson, A. J., and Kudlow, J. E. Phorbol ester or epidermal growth factor stimulates the concurrent accumulation of mRNA for the EGF receptor and its ligand transforming growth factor- α in a breast cancer cell line. *J. Biol. Chem.*, 264: 4021-4027, 1989.
- Gong, Y., Ballejo, G., Murphy, L. C., and Murphy, L. J. Differential effects of estrogen and antiestrogen on transforming growth factor gene expression in endometrial adenocarcinoma cells. *Cancer Res.*, 52: 1704-1709, 1992.
- Zugmaier, G., Ennis, B. W., Deschauer, B., Katz, D., Knabbe, C., Wilding, G., Daly, P., Lippman, M. E., and Dickson, R. B. Transforming growth factors type B1 and B2 are equipotent growth inhibitors of human breast cancer cells. *J. Cell. Physiol.*, 141: 353-361, 1989.
- Roberts, A. B., and Sporn, M. B. The transforming growth factor- β s. In: M. B. Sporn and A. B. Roberts (eds.), *Handbook of Experimental Pharmacology*, Vol. 95, pp. 419-472. New York: Springer-Verlag, 1990.
- Gorsch, S. M., Memoli, V. A., Stukel, T. A., Gold, L. I., and Arriack, B. A. Immunohistochemical staining for transforming growth factor β 1 associates with disease progression in human breast cancer. *Cancer Res.*, 52: 6949-6952, 1992.
- Kawamata, H., Azuma, M., Kameyama, S., Nan, L., and Oyasu, R. Effect of epidermal growth factor/transforming growth factor α and transforming growth factor β 1 on growth *in vitro* of rat urinary bladder carcinoma cells. *Cell Growth & Differ.*, 3: 819-825, 1992.
- Arteaga, C. L., Carty-Dugger, T., Moses, H. L., Hurd, S. D., and Pietenpol, J. A. Transforming growth factor β 1 can induce estrogen-independent tumorigenicity of human breast cancer cells in athymic mice. *Cell Growth & Differ.*, 4: 193-201, 1993.
- Knabbe, C., Lippman, M. E., Wakefield, L. M., Flanders, K. C., Derynck, R., Dickson, R. B., and Kasid, A. Evidence that transforming growth factor β is a hormonally regulated negative growth factor in human breast cancer cells. *Cell*, 48: 417-428, 1987.
- Rodeck, U., Bossler, A., Graeven, U., Fox, F. E., Nowell, P. C., Knabbe, C., and Kari, C. Transforming growth factor β production and responsiveness in normal human melanocytes and melanoma cells. *Cancer Res.*, 54: 575-581, 1994.
- MacIndoe, J. H., Woods, G. R., and Etre, C. A. The specific binding of estradiol and estrone and the subsequent distribution of estrogen receptor complexes in MCF-7 human breast cancer cells. *Steroids*, 39: 245-258, 1982.
- Katzenellenbogen, B. S., Kendra, K. L., Norman, M. J., and Berthois, Y. Prolifera-

- tion, hormonal responsiveness, and estrogen receptor content of MCF-7 human breast cancer cell lines grown in the short-term and long-term absence of estrogens. *Cancer Res.*, 47: 4355-4360, 1987.
21. Read, L. D., Greene, G. L., and Katzenellenbogen, B. S. Regulation of estrogen receptor messenger ribonucleic acid and protein levels in human breast cancer cell lines by sex steroid hormones, their antagonists and growth factors. *Mol. Endocrinol.*, 3: 295-304, 1989.
 22. Chomczynski, P., and Sacchi, N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal. Biochem.*, 162: 156-159, 1987.
 23. Derynck, R., Roberts, A. B., Winkler, M. E., Chen, E. Y., and Goeddel, D. V. Human transforming growth factor- α : precursor structure and expression in *E. coli*. *Cell*, 38: 287-297, 1984.
 24. Derynck, R., Jarrett, J. A., Chen, E. Y., Eaton, D. H., Bell, J. R., Assoian, R. K., Roberts, A. B., Sporn, M. B., and Goeddel, D. V. Human transforming growth factor β complementary sequence and expression in normal and transformed cells. *Nature (Lond.)*, 316: 701-705, 1985.
 25. Martin, R., Haendler, B., Hofer-Warbinek, R., Gaugitsch, H., Wrann, M., Schusener, H., Seifert, J. M., Bodmer, S., Fontana, A., and Hofer, E. Complementary DNA for human glioblastoma-derived T cell suppressor factor, a novel member of the transforming growth- β gene family. *EMBO J.*, 6: 3673-3677, 1987.
 26. Arrick, B. A., Korc, M., and Derynck, R. Differential regulation of expression of three transforming growth factor β species in human breast cancer cell lines by estradiol. *Cancer Res.*, 50: 299-303, 1990.
 27. Laborda, J. 36B4 cDNA used as an estradiol-independent mRNA control is the cDNA for human acidic ribosomal phosphoprotein PO. *Nucleic Acids Res.*, 19: 3998, 1991.
 28. Cho, H., Ng, P. A., and Katzenellenbogen, B. S. Differential regulation of gene expression by estrogen in estrogen-growth independent and -dependent MCF-7 human breast cancer cell sublines. *Mol. Endocrinol.*, 5: 1323-1330, 1991.
 29. Kingston, R. E. Introduction of DNA into mammalian cells. In: F. M. Ausubel, R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith, and K. Struhl (eds.), *Current Protocols in Molecular Biology*, 9.1.1-9.1.7, 1993.
 30. Nielsen, D. A., Chang, T. C., and Shapiro, D. J. A highly sensitive, mixed-phase assay for chloramphenicol acetyl transferase activity in transfected cells. *Anal. Biochem.*, 179: 19-23, 1989.
 31. Herbolm, P., Bourachot, B., and Yaniv, M. Two distinct enhancers with different cell specificities coexist in the regulatory region of polyoma. *Cell*, 39: 653-662, 1984.
 32. Reese, J. R., and Katzenellenbogen, B. S. Mutagenesis of cysteines in the hormone binding domain of the human estrogen receptor. *J. Biol. Chem.*, 266: 10880-10887, 1991.
 33. Murphy, C. S., Pink, J. J., and Jordan, V. C. Characterization of a receptor-negative, hormone-unresponsive clone derived from a T47D human breast cancer cell line kept under estrogen-free conditions. *Cancer Res.*, 50: 7285-7292, 1990.
 34. Daly, R. J., and Darbre, P. D. Cellular and molecular events in loss of estrogen sensitivity in ZR-75-1 and T-47-D human breast cancer cells. *Cancer Res.*, 50: 5868-5875, 1990.
 35. Brunner, N., Boulay, V., Fojo, A., Freter, C. E., Lippman, M. E., and Clarke, R. Acquisition of hormone-independent growth in MCF-7 cells is accompanied by increased expression of estrogen-regulated genes but without detectable DNA amplifications. *Cancer Res.*, 53: 283-290, 1993.
 36. Kokontis, J., Takakura, K., Hay, N., and Liao, S. Increased androgen receptor activity and altered c-myc expression in prostate cancer cells after long-term androgen deprivation. *Cancer Res.*, 54: 1566-1573, 1994.
 37. Quarby, V. E., Yarbrough, W. G., Lubahn, D. B., French, F. S., and Wilson, E. M. Autologous down-regulation of androgen receptor messenger ribonucleic acid. *Mol. Endocrinol.*, 4: 22-28, 1990.
 38. Wolf, D., Herzinger, T., Herminkink, H., Blaschke, D., and Hörz, W. Transcriptional and posttranscriptional regulation of human androgen receptor by androgen. *Mol. Endocrinol.*, 7: 924-936, 1993.
 39. Darbre, P., and King, R. J. B. Progression to steroid autonomy in S115 mouse mammary tumor cells: role of DNA methylation. *J. Cell Biol.*, 99: 1410-1415, 1984.
 40. Darbre, P. D., and King, R. J. B. Progression to steroid insensitivity can occur irrespective of the presence of functional steroid receptors. *Cell*, 51: 521-528, 1987.
 41. Read, L. D., and Katzenellenbogen, B. S. Characterization and regulation of estrogen and progesterone receptors in breast cancer. In: R. B. Dickson and M. E. Lippman (eds.), *Genes, Oncogenes, and Hormones: Advances in Cellular and Molecular Biology of Breast Cancer*, pp. 277-299. Boston: Kluwer Academic Publishers, 1991.
 42. Murphy, L. J., Murphy, L. C., and Friesen, H. G. Estrogen induces insulin-like growth factor-I expression in the rat uterus. *Mol. Endocrinol.*, 1: 445-450, 1987.
 43. Mukku, V. R., and Stancel, G. M. Regulation of epidermal growth factor receptor by estrogen. *J. Biol. Chem.*, 260: 9820-9826, 1985.
 44. Iwase, H., Kobayashi, S., Itoh, Y., Kuzushima, T., Yamashita, H., Iwata, H., Naito, A., Yamashita, T., Itoh, K., and Masaoka, A. Clinical value of enzyme immunoassay of epidermal growth factor receptor in human breast cancer. *Breast Cancer Res. Treat.*, 28: 215-221, 1993.
 45. Kienhuis, C. B. M., Sluys, M., de Goeij, C. C. J., Koenders, P. G., and Benraad, T. J. Epidermal growth factor receptor levels increase but epidermal growth factor receptor ligand levels decrease in mouse mammary tumors during progression from hormone dependence to hormone independence. *Breast Cancer Res. Treat.*, 26: 289-295, 1993.
 46. Lippman, M. E., Dickson, R. B., Bates, S., Knabbe, C., Huff, K., Swain, S., McManaway, M., Bronzert, D., Kasid, A., and Gelmann, E. P. Autocrine and paracrine growth regulation of human breast cancer. *Breast Cancer Res. Treat.*, 7: 59-70, 1986.
 47. Kenney, N. J., Saeki, T., Gottardis, M., Kim, N., Garcia-Morales, P., Martin, M. B., Normanno, N., Ciardiello, F., Day, A., Cutler, M. L., and Salomon, D. S. Expression of transforming growth factor α antisense mRNA inhibits the estrogen-induced production of TGF α and estrogen-induced proliferation of estrogen-responsive human breast cancer cells. *J. Cell. Physiol.*, 156: 497-514, 1993.
 48. Arteaga, C. L., Coronado, E., and Osborne, C. K. Blockade of the epidermal growth factor receptor inhibits transforming growth factor α -induced but not estrogen-induced growth of hormone-dependent human breast cancer. *Mol. Endocrinol.*, 2: 1064-1069, 1988.
 49. Clarke, R., Brunner, N., Katz, D., Glanz, P., Dickson, R. B., Lippman, M. E., and Kern, F. G. The effects of a constitutive expression of transforming growth factor- α on the growth of MCF-7 human breast cancer cells *in vitro* and *in vivo*. *Mol. Endocrinol.*, 3: 372-380, 1989.
 50. Kyprianou, N., English, H. F., Davidson, N. E., and Isaacs, J. T. Programmed cell death during regression of the MCF-7 human breast cancer following estrogen ablation. *Cancer Res.*, 51: 162-166, 1991.
 51. Komm, B. S., Terpening, C. M., Benz, D. J., Graeme, K. A., Gallegros, A., Korc, M., Greene, G. L., O'Malley, B. W., and Haussler, M. R. Estrogen binding, receptor mRNA, and biological response in osteoblast-like osteosarcoma cells. *Science (Washington DC)*, 241: 81-84, 1988.
 52. Thompson, A. M., Kerr, D. J., and Steel, C. M. Transforming growth factor B1 is implicated in the failure of tamoxifen therapy in human breast cancer. *Br. J. Cancer*, 63: 609-614, 1991.
 53. Hugget, A. C., Hampton, L. L., Ford, C. P., Wirth, P. J., and Thorgeirsson, S. S. Altered responsiveness of rat liver epithelial cells to transforming growth factor β 1 following their transformation with v-*raf*. *Cancer Res.*, 50: 7468-7475, 1990.

Observations arising from the use of pure antioestrogens on oestrogen-responsive (MCF-7) and oestrogen growth-independent (K3) human breast cancer cells

by R I Nicholson, J M W Gee, A B Francis, D L Manning,
A E Wakeling and B S Katzenellenbogen

INTRODUCTION

During the last 7 years the Breast Cancer Group within the Tenovus Cancer Research Centre has maintained an involvement in the use of pure antioestrogens in two important areas of breast cancer research. First, their development as clinical agents, where we hoped to induce total oestrogen deprivation and thereby improve the effectiveness of first-line endocrine therapy (Nicholson *et al.* 1992, Nicholson 1993, Nicholson *et al.* 1993a, DeFriend *et al.* 1994, Nicholson *et al.* 1994c). Second, as pharmacological probes to investigate the cellular and molecular actions of oestrogens and tamoxifen (Nicholson *et al.* 1988, Weatherill *et al.* 1988, Wilson *et al.* 1990). Implicit in each of these areas of research are questions associated with the impact which pure antioestrogens might have on the therapy of endocrine-resistant states and whether resistance develops as a consequence of incomplete oestrogen withdrawal, with tumour cells more efficiently utilising either a reduced oestrogenic pool or the agonistic activity of an antioestrogen, or whether the resistant cells have completely circumvented the need for oestrogen receptor (ER)-mediated growth and hence sensitivity to the antitumour properties of pure antioestrogens (Nicholson *et al.* 1994c).

On this basis, in the current article we seek to describe a number of the properties exhibited by pure antioestrogens in oestrogen-responsive MCF-7 human breast cancer cells (Nicholson *et al.* 1990,

Nicholson *et al.* 1995) and in the oestrogen growth-independent variant K3 (Katzenellenbogen *et al.* 1987, Clarke *et al.* 1989, Cho *et al.* 1991, Reese & Katzenellenbogen 1992) of this tumour cell line. Limited data will also be presented on the growth-inhibitory properties of 4-(3-methylanilino)quinazoline (aniloquinazoline), a tyrosine kinase inhibitor which shows specificity for epidermal growth factor (EGF)-receptor signalling (Wakeling *et al.* 1994). The data presented are consistent with ER-mediated growth being important not only in MCF-7 cells, but also in their oestrogen-resistant variant, with transforming growth factor α (TGF α) possibly playing a supportive growth-regulatory role.

COMPARATIVE GROWTH EFFECTS OF OESTRADIOL AND ANTIOESTROGENS ON WILD-TYPE AND K3 MCF-7 CELLS

K3 cells were originally isolated by the exposure of MCF-7 human breast cancer cells to culture conditions low in oestrogenic substances (Katzenellenbogen *et al.* 1987). Thus, by growing MCF-7 cells in phenol red-free media and 5% dextran-coated charcoal-treated (DCC-stripped) foetal calf serum (FCS) for prolonged periods, a stable cell variant (K3) was obtained which showed a markedly increased basal rate of proliferation where added oestrogen was unable to increase this rate of

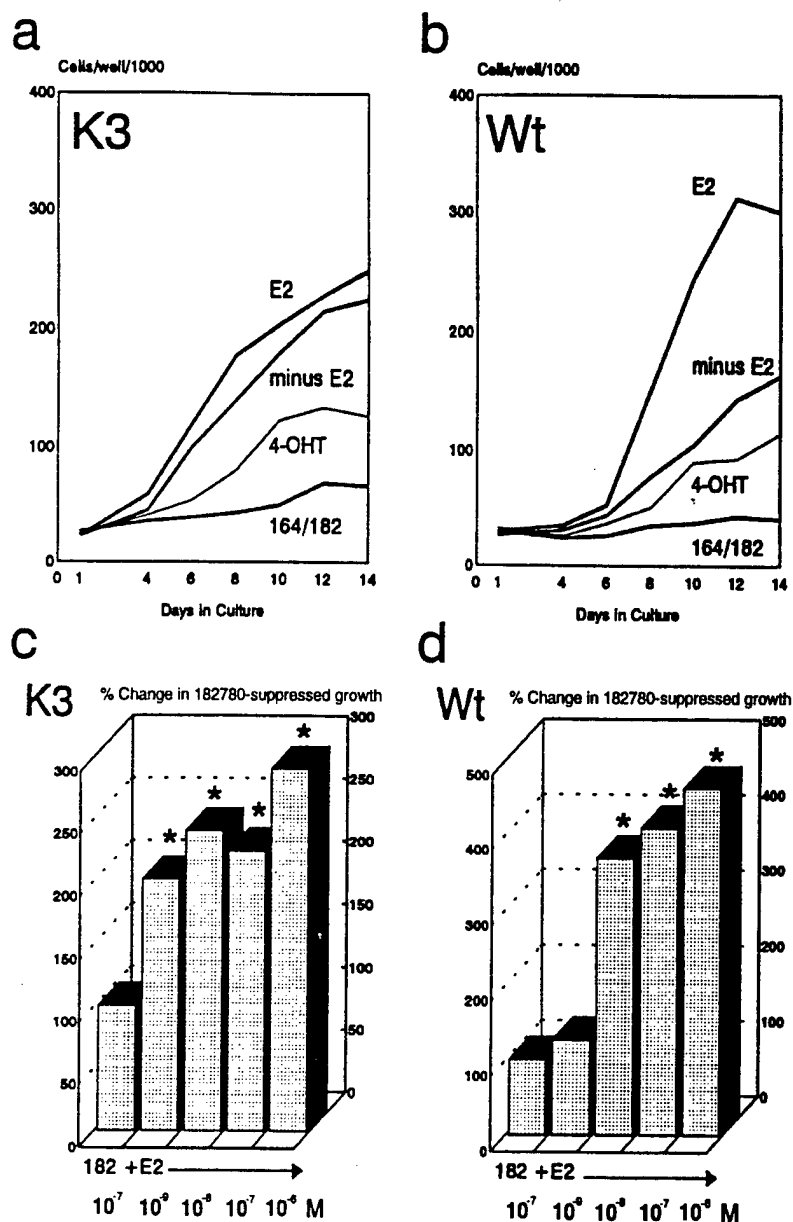


Figure 1 Characterisation of the growth of K3 and Wt MCF-7 cells in monolayer culture. (a and b) The cells were grown in multiwell dishes in white RPMI tissue culture medium with 5% DCC-stripped FCS (medium A); without additives (minus E2), and medium A containing 10⁻⁹M oestradiol (E2), 10⁻⁷M 4-hydroxytamoxifen (4-OHT), and 10⁻⁷M ICI 182780 (164/182) for up to 14 days. (c and d) The cells were grown in medium A containing 10⁻⁷M ICI 182780 for 8 days prior to the addition of various doses of oestradiol (182+E2). These cultures were harvested on day 14 after the addition of oestradiol. Cell numbers were assessed by the use of a Coulter counter and are the mean of 3 replicate cultures counted in triplicate. **P* v 182<0.05; statistical analysis performed using a Mann-Whitney U test.

proliferation further duplicated in Figure 1. The effect of ad growth of our W endogenous oestro

Despite their dependence, early st K3 cells could be tamoxifen (Katzner *al.* 1989). This eff In the present stud trogen ICI 182780 establish whether can achieve a gre: use of antioestrog oestrogen-like act ure 1a shows the 182780 exceeding allowing at maxim number. Over seve the tumour cell do K3 and wild-type This contrasts wit oestrogen-withdra *al.* 1987, Clarke *et* tamoxifen-treated 1987).

Importantly, th tion shown by pu: tumour cell lines : in that their action cer cells and concentrations (10 sociation constant the actions of an: oestradiol (see ref property is demc both in K3 and . Indeed, ICI 182 show an increase: parison with wil 10⁻⁷M ICI 182780

THE PARADOX SOLUTION

These data represe Wt cells, each of v

proliferation further. These results are essentially duplicated in Figure 1 and contrast with the stimulatory effect of added oestradiol (10^{-9} M) on the growth of our Wt-MCF-7 cells in media lacking endogenous oestrogens.

Despite their apparent oestrogen growth-independence, early studies established that the growth of K3 cells could be inhibited by 10^{-7} M 4-hydroxy-tamoxifen (Katzenellenbogen *et al.* 1987, Clarke *et al.* 1989). This effect is also illustrated in Figure 1a. In the present study we have used the pure antioestrogen ICI 182780 (10^{-7} M) (Wakeling *et al.* 1991) to establish whether complete oestrogen deprivation can achieve a greater antitumour effect than can the use of antioestrogens, like tamoxifen, with partial oestrogen-like activity (Nicholson *et al.* 1995). Figure 1a shows the growth-inhibitory activity of ICI 182780 exceeding that of 4-hydroxytamoxifen, allowing at maximum 2 doublings of the initial cell number. Over several experiments we have estimated the tumour cell doubling time for ICI 182780-treated K3 and wild-type (Wt) cells to be in excess of 150h. This contrasts with 32-35h for oestrogen-treated and oestrogen-withdrawn K3 cells (Katzenellenbogen *et al.* 1987, Clarke *et al.* 1989) and >80h for 4-hydroxytamoxifen-treated cells (Katzenellenbogen *et al.* 1987).

Importantly, the improved level of growth inhibition shown by pure antioestrogens in several breast tumour cell lines appears specific for ER signalling, in that their actions are restricted to ER-positive cancer cells and they are achieved at molar concentrations (10^{-10} to 10^{-9}) equivalent to the dissociation constant for their binding to ER. Moreover, the actions of antioestrogens may be reversed by oestradiol (see refs in Nicholson *et al.* 1994a). This property is demonstrated for pure antioestrogens both in K3 and in Wt cells in Figure 1c and d. Indeed, ICI 182780 growth-suppressed K3 cells show an increased sensitivity to oestradiol in comparison with wild-type cells, with the effects of 10^{-7} M ICI 182780 reversed by 10^{-9} M oestradiol.

THE PARADOX AND A POTENTIAL SOLUTION

These data represent a paradox both for K3 and for Wt cells, each of which are capable of growth in the

apparent absence of oestradiol (K3>Wt), yet are growth inhibited by a pure antioestrogen whose perceived mechanism of action is to antagonise the cellular actions of oestrogens at the ER. Indeed, their inhibitory actions may be reversed (K3>Wt) by oestradiol. A potential solution to this paradox arises from the observation that the cellular actions of the ER, in either an occupied (Wakeling *et al.* 1991, refs in Nicholson *et al.* 1994a) or unoccupied (Ignar-Trowbridge *et al.* 1992) form, may be potentiated by the presence of growth factors. ER-induced growth responses, therefore, may require only limited amounts of steroid, with differences between K3 and Wt cells reflecting altered regulation of growth factor production or cellular sensitivity to their actions.

AN INVOLVEMENT OF TGF α ?

As may be seen in Figure 2, when grown in an oestrogen-depleted environment K3 cells show a higher basal expression of the mitogenic growth factor TGF α than do Wt cells. Furthermore, in K3 cells the intracellular level of this protein is only poorly induced by oestradiol compared with a twofold increase seen in Wt cells. This parallels the lack of activity of the steroid on K3 growth. In each instance, ICI 182780 reduced the basal expression of TGF α . Importantly, the reduction in TGF α levels in pure antioestrogen-treated cells accompanies a substantial fall in their ER content (Fig. 2c and d; Reese & Katzenellenbogen 1992). This action would minimise the opportunity for cross talk between ER signalling and TGF α signalling pathways. Interestingly, K3 cells also show an elevated basal expression of pS2 (Cho *et al.* 1991), a protein whose gene promoter contains response elements both for oestradiol and for TGF α (Nunez *et al.* 1989). Once again, the expression of this protein is efficiently reduced by the presence of the pure antioestrogen (Nicholson *et al.* 1995).

Finally, we have examined the effects of 4-(3-methylanilino)quinazoline (ZM163613), a tyrosine kinase inhibitor reported to show specificity for EGF-receptor signalling (Wakeling *et al.* 1994, Ward *et al.* 1994), on K3 and Wt cells in order to determine whether TGF α is directly involved in growth signalling and oestrogen-regulated gene expression. The data shown in Figure 3a and b show that the Wt cells

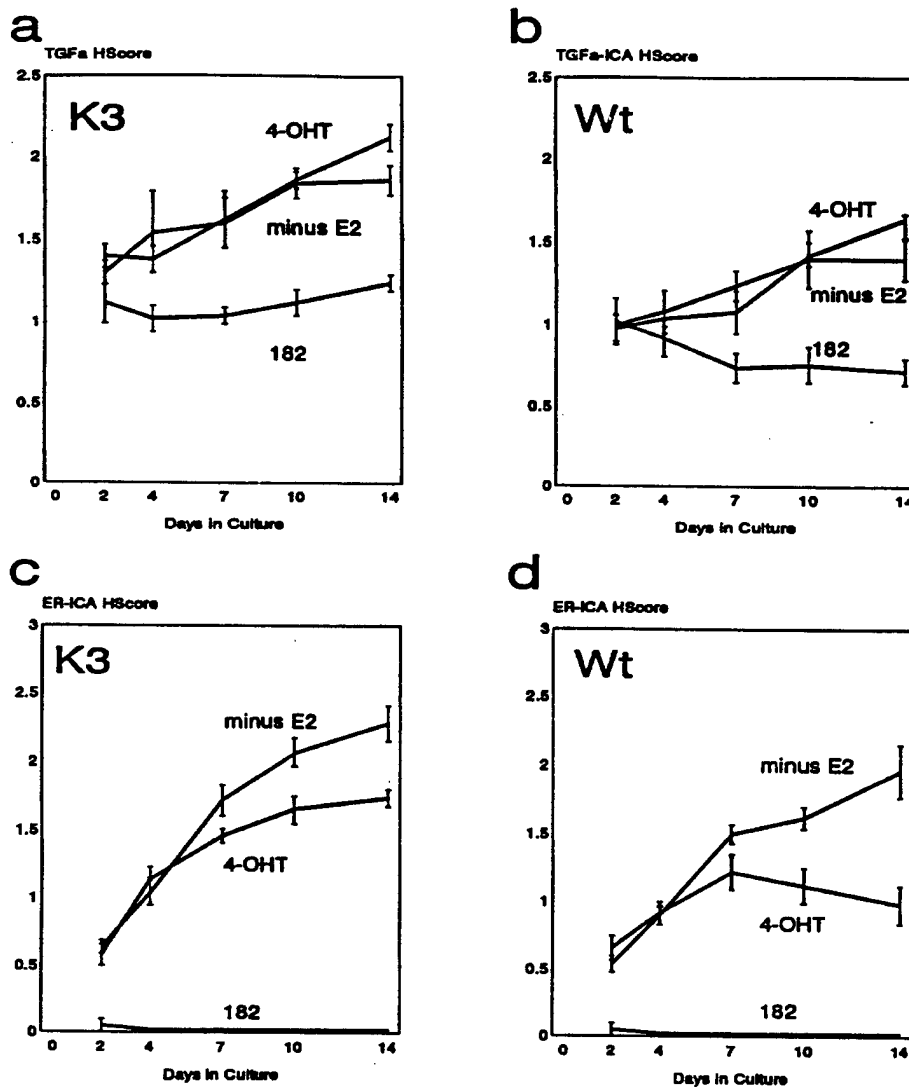


Figure 2 Immunohistochemical characterisation of K3 and Wt MCF-7 cells. The cells were cultured on 3-aminopropyltriethoxysilane-coated glass coverslips in medium A containing no additions (minus E2), 10^{-9} M oestradiol (E2), 10^{-7} M 4-hydroxytamoxifen (4-OHT), and 10^{-7} M ICI 182780 (182) for up to 14 days. TGFα (a and b) and ER (c and d) assays were performed according to the methods of Nicholson *et al.* (1991, 1993b) and Walker *et al.* (1988), respectively. The results are shown as mean values \pm S.D. of 5 replicates from a minimum of 2 coverslips. H scores were calculated according to the method of Gee *et al.* (1994).

are strongly growth inhibited by the drug at a concentration of $10\mu\text{M}$. At this concentration, the cells show reduced basal progesterone-receptor and pS2 levels whilst maintaining ER and TGFα cellular con-

centrations (Nicholson *et al.* 1995). However, an identical dose of ZM163613 is less growth inhibitory to K3 cells (Fig. 3a) and does not alter oestrogen-regulated gene expression, although some growth

inhibition of K3 cell (RI Nicholson, unpublished, achieved by $50\mu\text{M}$ Z

CONCLUSIONS

Several conclusions the results presented

(1) The importance retained in the basic growth-independent observations made which are sensitised drug (Osborne *et al.*

(2) TGFα signalling growth and circumv levels. This response potentially decreases ZM163613.

(3) Pure antioestrogen effects, in Wt and ER and TGFα level between these growth

inhibition of K3 cells (Fig. 3a) and decrease in pS2 (RI Nicholson, unpublished observations) may be achieved by 50 μ M ZM163613.

CONCLUSIONS

Several conclusions may be arrived at on the basis of the results presented.

- (1) The importance of ER-mediated signalling is retained in the basal growth responses of oestrogen growth-independent K3 cells and is in parallel with observations made on tamoxifen-resistant tumours which are sensitised to the agonistic activity of the drug (Osborne *et al.* 1994).
- (2) TGF α signalling may impinge on ER-mediated growth and circumvent the need for high oestrogen levels. This response may be exaggerated in K3 cells, potentially decreasing the cellular sensitivity to ZM163613.
- (3) Pure antioestrogens antagonise ER-mediated effects, in Wt and K3 cells, possibly by decreasing ER and TGF α levels and thereby reducing cross talk between these growth-signalling pathways.

Finally, it is interesting that we have also observed that a failure of ER-positive advanced breast cancer to respond to antihormones correlates with elevated TGF α levels (Nicholson *et al.* 1994b) and elevated cell-proliferation rates, evidenced by an increased Ki67 immunostaining (Nicholson *et al.* 1991, Nicholson *et al.* 1993b); factors which in K3 cells are associated with acquired oestrogen growth-independence. If these factors are causative in the loss of oestrogen growth-responsiveness, then primary and acquired endocrine resistance may occur on a similar developmental pathway and be equally vulnerable to pure antioestrogens. Trials to examine these possibilities are awaited.

ACKNOWLEDGEMENTS

The authors gratefully acknowledge the financial support of the Tenovus Organisation (R I N, J M W G, A B F, and D L M), The Association for International Cancer Research (R I N), The National Institutes of Health, and Susan G Komen Foundation (B S K).

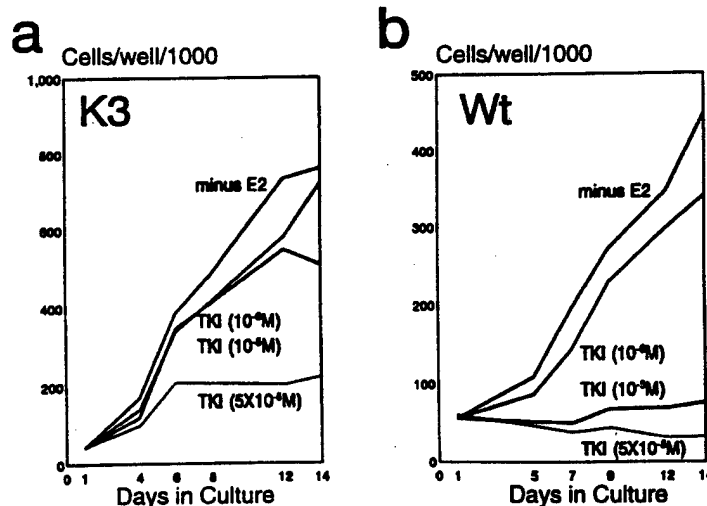


Figure 3 Effect of 4-(3-methylamino)quinazoline on the growth of K3 and Wt cells. The cells were grown as described in Figure 1, in medium A alone (minus E2) or containing the stated dose of the tyrosine kinase inhibitor (TKI). The results presented are the mean of 3 replicate cultures counted in triplicate.

REFERENCES

- Cho H, Peter ANG & Katzenellenbogen BS 1991 Differential regulation of gene expression by estrogen in estrogen growth-independent and -dependent MCF-7 human breast cancer cell sublines. *Molecular Endocrinology* **5** 1323-1330.
- Clarke R, Brunner N, Katzenellenbogen BS, Thompson EW, Norman MJ, Koppi C, Paik S, Lippman ME & Dickson RB 1989 Progression of human breast cancer cells from hormone-dependent to hormone-independent growth both *in vitro* and *in vivo*. *Proceedings of the National Academy of Sciences of the USA* **86** 3649-3653.
- DeFriend DL, Howell A, Nicholson RI, Anderson E, Dowsett M, Mansel RE, Bundred NJ, Robertson JFR, Saunders C, Baum M, Walton P, Sutcliffe F & Wakeling AE 1994 Investigation of a new pure antioestrogen (ICI 182780) in women with primary breast cancer. *Cancer Research* **54** 1-7.
- Gee JMW, Robertson JFR, Hoyle HB, Kyme SR, McClelland RA, Ellis IO, Blamey RW & Nicholson RI 1994 Immunocytochemical localisation of bcl-2 protein in human breast cancer and its relationship to a series of prognostic markers and response to endocrine therapy. *International Journal of Cancer* **59** 619-628.
- Ignar-Trowbridge DM, Nelson KG, Bidwell MC, Curtis SW, Washburn TF, McLachan JA & Korach, KS 1992 Coupling of dual signalling pathways: epidermal growth factor action involves the estrogen receptor. *Proceedings of the National Academy of Sciences of the USA* **89** 4658-4662.
- Katzenellenbogen BS, Kendra KL, Norman MJ & Berthois Y 1987 Proliferation, hormonal responsiveness, and estrogen receptor content of MCF-7 human breast cancer cells grown in the short-term and long-term absence of estrogens. *Cancer Research* **47** 4355-4360.
- Nicholson RI 1993 Recent advances in the antihormonal therapy of breast cancer. *Current Opinion in Investigational Drugs* **2** 1259-1268.
- Nicholson RI, Gotting KE, Gee JMW & Walker KJ 1988 Actions of oestrogens and antioestrogens on rat mammary gland development: Relevance to breast cancer prevention. *Journal of Steroid Biochemistry* **30** 95-103.
- Nicholson RI, Walker KJ, Bouzubar N, Gee JMW, Rushmere N & Davies P 1990 Oestrogen deprivation in breast cancer - Clinical, experimental and biological aspects. *Annals of the New York Academy of Sciences* **595** 316-327.
- Nicholson RI, Bouzubar N & Walker KJ 1991 Hormone sensitivity in breast cancer: Influence of heterogeneity of oestrogen receptor and cell proliferation. *European Journal of Cancer* **27** 908-913.
- Nicholson RI, Eaton CL & Manning DL 1992 New developments in the endocrine management of breast cancer. In *Recent Advances in Endocrinology and Metabolism*, vol 4, pp 151-165. Edinburgh: Churchill Livingstone.
- Nicholson RI, Manning DL & Gee JMW 1993a New antihormonal approaches to breast cancer therapy. *Drugs of Today* **29** 363-372.
- Nicholson RI, McClelland RA, Finlay P, Eaton CL, Gullick WJ, Dixon AR, Robertson JFR, Ellis IO & Blamey RW 1993b Relationship between EGF-R, c-erbB-2 protein expression and Ki67 immunostaining in breast cancer and hormone sensitivity. *European Journal of Cancer* **29A** 1018-1023.
- Nicholson RI, Gee JMW, Eaton CL, Manning DL, Mansell RE, Sharma AK, Douglas-Jones A, Price-Thomas M, Howell A, DeFriend DJ, Bundred NJ, Anderson E, Robertson JFR, Blamey RW, Dowsett M, Baum M, Walton P & Wakeling AW 1994a Pure antioestrogens in breast cancer: experimental and clinical observations. In *International Symposium on Sex Hormones and Antihormones in Endocrine Dependent Pathology: Basic and Clinical Aspects*. Eds M Motta & M Serio. Excerpta Medica International Congress Series 1064. Amsterdam: Elsevier Science BV.
- Nicholson RI, McClelland RA, Gee JMW, Manning DL, Cannon P, Robertson JFR, Ellis IO & Blamey RW 1994b Transforming growth factor α and endocrine sensitivity in breast cancer. *Cancer Research* **54** 1684-1689.
- Nicholson RI, Francis AB, McClelland RA, Manning DL & Gee JMW 1994c Pure anti-oestrogens (ICI 164384 and ICI 182780) and breast cancer: is the attainment of complete oestrogen withdrawal worthwhile? *Endocrine-Related Cancer* **1** (4) 5-17.
- Nicholson RI, Gee JMW, Manning DL, Wakeling AE & Katzenellenbogen BS 1995 Responses to pure antiestrogens (ICI 164384 and ICI 182780) in estrogen sensitive and resistant experimental and clinical breast cancer. *Annals of the New York Academy of Sciences* (In Press).

- Walker KJ 1991 Hormone influence of receptor and cell proliferation. *Journal of Cancer* 27
- Manning DL 1992 New management of breast cancer. *Endocrinology and Metabolism* 5. Edinburgh: Churchill Livingstone
- Gee JMW 1993a New breast cancer therapy. *Endocrinology and Metabolism* 5. Edinburgh: Churchill Livingstone
- Wakeling AE, Manning DL, Glas-Jones A, DeFriend DJ, Bundred R, Blamey RW, P & Wakeling AW 1993 Breast cancer: endocrine therapy. In *Endocrine and Metabolism: Independent Pathology*: eds M Motta & M Serio. Congress Series 1064. BV. Elsevier
- Gee JMW, Manning DL, Ellis IO & Blamey RW 1993 Receptor α and endocrine therapy. *Cancer Research* 54
- Land RA, Manning DL 1993 Estrogens (ICI 164384): is the attainment of a worthwhile goal worthwhile? *Journal of Cancer* 4) 5-17.
- Manning DL, Wakeling AE & Bundred R 1993 Responses to pure oestrogen (ICI 182780) in experimental and clinical breast cancer. *Journal of the New York Academy of Sciences* 714
- Nunez AM, Berry M, Imler JL & Chambon, P 1989 The 5'-flanking region of the pS2 gene contains a complex enhancer region responsive to oestrogens, epidermal growth factor, a tumour promoter (TPA), the c-Ha-ras oncoprotein and the c-jun protein. *EMBO Journal* 8 823-829.
- Osborne CK, Jarman M, McCague R, Coronado EB, Hilsenbeck SG & Wakeling AE 1994 The importance of tamoxifen metabolism in tamoxifen-stimulated growth. *Cancer Chemotherapy and Pharmacology* 34 89-95.
- Reese JC & Katzenellenbogen BS 1992 Examination of the DNA-binding of estrogen receptor in whole cells: Implications for hormone independent transactivation and the actions of antiestrogens. *Molecular and Cellular Biology* 12 4531-4538.
- Wakeling AE, Dukes M & Bowler J 1991 A potent specific pure antiestrogen with clinical potential. *Cancer Research* 51 3867-3873.
- Wakeling AE, Barker AJ, Davies DH, Brown DS, Green LR, Cartledge SA & Woodburn JR 1994 Inhibition of EGF-receptor tyrosine kinase activity by 4-anilinoquinazolines. *British Journal of Cancer* 69 Supplement XXI, p18, Abstract 6.6.
- Walker KJ, Bouzubar N, Robertson J, Ellis IO, Elston CW, Blamey RW, Wilson DW, Griffiths K & Nicholson RI 1988 Immunocytochemical localisation of estrogen receptor in human breast tissue. *Cancer Research* 48 6517-6522.
- Ward WHJ, Cook PN, Slater AM, Davies DH, Holdgate GA & Green LR 1994 Epidermal growth factor receptor tyrosine kinase. Investigation of catalytic mechanism, structure-based searching and discovery of a potent inhibitor. *Biochemical Pharmacology* 46 659-666.
- Weatherill PJ, Wilson APM, Nicholson RI, Davies P & Wakeling AE 1988 Interaction of the antioestrogen ICI 164384 with the oestrogen receptor. *Journal of Steroid Biochemistry* 30 263-266.
- Wilson APM, Weatherill PJ, Nicholson RI, Davies P & Wakeling AE 1990 A comparative study of the interaction of oestradiol and the steroidal pure antioestrogen, ICI 164384 with molybdate stabilised oestrogen receptor. *Journal of Steroid Biochemistry* 35 421-428.
- Nicholson RI, Manning DL, Wakeling AE, Francis AB, Manning DL, Manning AE, Wakeling¹ and Katzenellenbogen² 1993 Breast Cancer Unit, Tenovus Cancer Research Centre, UWCM, Cardiff CF4 4XX, UK, ¹Cancer Research Department, ZENECA Pharmaceuticals, Alderley Edge, Macclesfield, Cheshire SK10 4TG, UK, and ²Department of Physiology and Biophysics, University of Illinois, Urbana, IL, USA



Antiestrogens: Mechanisms and Actions in Target Cells

Benita S. Katzenellenbogen,* Monica M. Montano, Pascale Le Goff,
David J. Schodin, W. Lee Kraus, Bhavna Bhardwaj
and Nariaki Fujimoto

*Department of Physiology and Biophysics and Department of Cell and Structural Biology, University of Illinois and
University of Illinois College of Medicine, Urbana, IL 61801, U.S.A.*

Antiestrogens, acting via the estrogen receptor (ER) evoke conformational changes in the ER and inhibit the effects of estrogens as well as exerting anti-growth factor activities. Although the binding of estrogens and antiestrogens is mutually competitive, studies with ER mutants indicate that some of the contact sites of estrogens and antiestrogens are likely different. Some mutations in the hormone-binding domain of the ER and deletions of C-terminal regions result in ligand discrimination mutants, i.e. receptors that are differentially altered in their ability to bind and/or mediate the actions of estrogens vs antiestrogens. Studies in a variety of cell lines and with different promoters indicate marked cell context- and promoter-dependence in the actions of antiestrogens and variant ERs. In several cell systems, estrogens and protein kinase activators such as cAMP synergize to enhance the transcriptional activity of the ER in a promoter-specific manner. In addition, cAMP changes the agonist/antagonist balance of tamoxifen-like antiestrogens, increasing their agonistic activity and reducing their efficacy in reversing estrogen actions. Estrogens, and antiestrogens to a lesser extent, as well as protein kinase activators and growth factors increase phosphorylation of the ER and/or proteins involved in the ER-specific response pathway. These changes in phosphorylation alter the biological effectiveness of the ER. Multiple interactions among different cellular signal transduction systems are involved in the regulation of cell proliferation and gene expression by estrogens and antiestrogens.

J. Steroid Biochem. Molec. Biol., Vol. 53, No. 1-6, pp. 387-393, 1995

INTRODUCTION: ESTROGEN TARGET TISSUES AND ANTIESTROGEN EFFECTIVENESS

Estrogens influence the growth, differentiation and functioning of many target tissues. These include tissues of the reproductive system such as the mammary gland and uterus, cells in the hypothalamus and pituitary, as well as bone where estrogens play important roles in bone maintenance; and the liver and cardiovascular systems where estrogens influence liver metabolism, the production of plasma lipoproteins, and exert cardioprotective effects [1-3]. Estrogens, in addition to stimulating mammary gland growth and duct development, also increase proliferation and metastatic activity of breast cancer cells [4] and stimulate the proliferation

of uterine cells [1]. Antiestrogens, which antagonize the actions of estrogens, therefore have much potential as important therapeutic agents. Our studies have examined the effects of antiestrogens on a variety of target cells including liver [5] and hypothalamus and pituitary [6], but have primarily focused on their effects on breast cancer and uterine cells [7].

The actions of estrogens on breast cancer and uterine cells are antagonized by antiestrogens, which bind to the estrogen receptor (ER) in a manner that is competitive with estrogen but they fail to effectively activate gene transcription [7-9]. Two of the major challenges in studies on antiestrogens are to understand what accounts for their antagonistic effectiveness as well as the partial agonistic effects of some antiestrogens; and to understand how one can achieve tissue selective agonistic/antagonistic effects of these compounds. One of our approaches to addressing these issues has been to try to understand in detail how the ER discriminates between

*Proceedings of the IX International Congress on Hormonal Steroids,
Dallas, Texas, U.S.A., 24-29 September 1994.*

*Correspondence to B. S. Katzenellenbogen.

estrogen and antiestrogen ligands and between different categories of antiestrogens. This has involved the generation and analysis of variant human ERs with mutations throughout the ER hormone-binding domain and study of the activity of these receptors on different estrogen-responsive genes in several cell backgrounds when liganded with antiestrogen or estrogen. These studies and those of others have provided consistent evidence for the promoter-specific and cell-specific actions of the estrogen-occupied and antiestrogen-occupied ER. In addition, in the studies described below, we have observed that protein kinase activators enhance the transcriptional activity of the ER and alter the agonist/antagonist balance of some antiestrogens, suggesting that changes in cellular phosphorylation state should be important in determining the effectiveness of antiestrogens as estrogen antagonists.

ANALYSIS OF THE ER HORMONE BINDING DOMAIN AND LIGAND DISCRIMINATION

We have examined the interactions of estrogen and antiestrogens with the ER and the modulation of ER activity by phosphorylation and interaction with other proteins which result in changes in ER-mediated responses. Studies by us [10–17] have provided strong documentation that the response of genes to estrogen

and antiestrogen depend on four important factors: (1) the nature of the ER, i.e. whether it is wild-type or variant; (2) the promoter; (3) the cell context; and (4) the ligand. The gene response, in addition, can be modulated by cAMP, growth factors, and agents that affect protein kinases and cell phosphorylation [15, 18–21]. These may account for differences in the relative agonism/antagonism of antiestrogens like tamoxifen on different genes and in different target cells such as those in breast cancer cells, versus uterus, versus bone.

Antiestrogens are believed to exert their effects in large measure by blocking the actions of estrogens by competing for binding to the ER and altering ER conformation such that the receptor fails to effectively activate gene transcription. In addition, antiestrogens exert anti-growth factor activities, via a mechanism that requires ER but is still not fully understood [22].

Models of antiestrogen action at the molecular level are beginning to emerge, and recent biological studies as well indicate that antiestrogens fall into two distinct categories: antiestrogens, such as tamoxifen, that are mixed or partial agonists/antagonists (type I), and compounds, such as ICI 164,384, that are complete/pure antagonists (type II). The type I antihormone-ER complexes appear to bind as dimers to estrogen response elements (EREs); there, they block hormone-dependent transcription activation mediated by region E of the

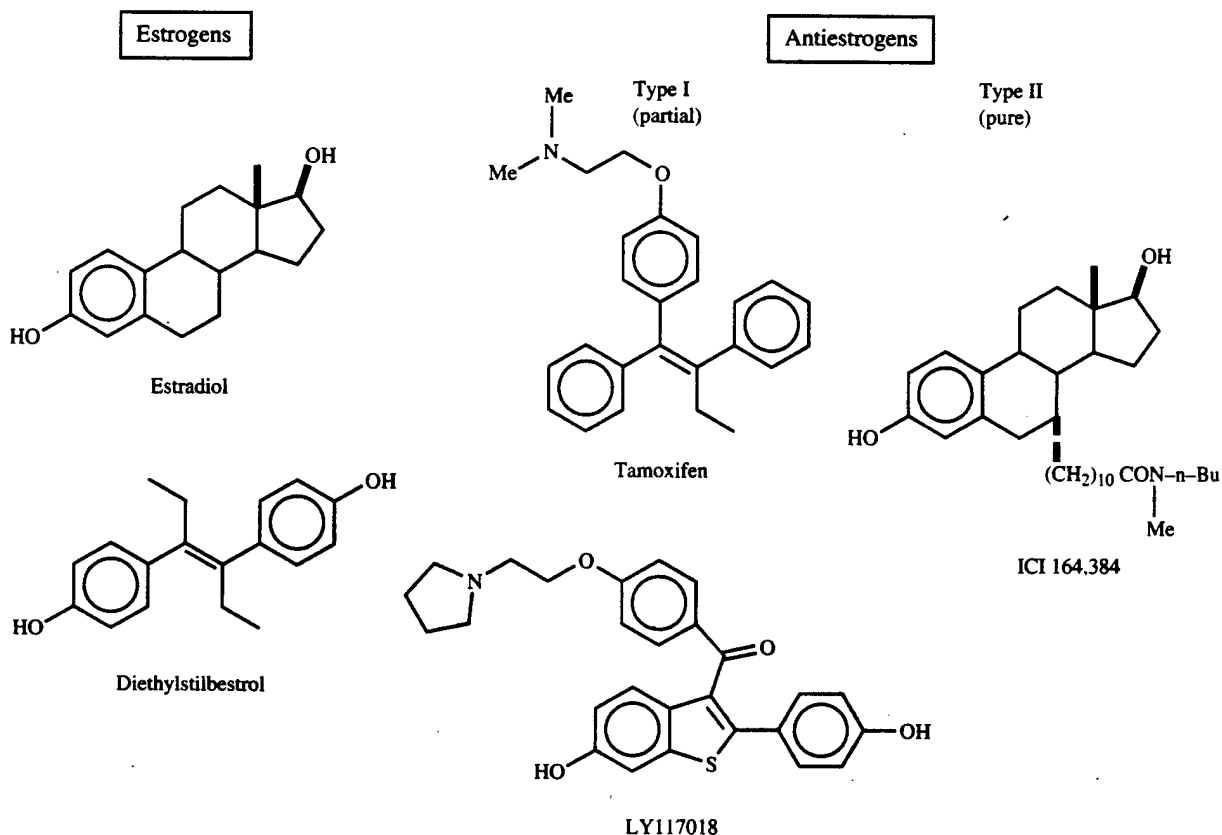


Fig. 1. Structures of several estrogenic and antiestrogenic ligands for the estrogen receptor used in our studies. The antiestrogens include the nonsteroidal compounds tamoxifen and LY117018 that often show partial agonist/antagonist activity (type I antiestrogens) and the steroidal, more pure antiestrogen ICI164,384 (type II antiestrogen).

receptor, but are believed to have little or no effect on the hormone-independent transcription activation function located in region A/B of the receptor [16]. Thus, they are generally partial or mixed agonist/antagonists, and their action must involve some subtle difference in ligand-receptor interaction, very likely associated with the basic or polar side chain that characterizes the antagonist members of this class. In the case of the more complete antagonists, such as ICI 164,384, obstruction of ER binding to DNA and reduction of the ER content of target cells appear to contribute to [23, 24], but may not fully explain, the pure antagonist character of this antiestrogen [25]. The structures of these antiestrogens, which can be both steroidal or non-steroidal in nature, are shown in Fig. 1, along with the structures of the naturally occurring estrogen estradiol, and the non-steroidal synthetic estrogen diethylstilbestrol. Of note, is the fact that antiestrogens typically have a bulky side chain which is basic or polar. This side chain is important for antiestrogenic activity; removal of this side chain results in a compound which is no longer an antiestrogen and, instead, has only estrogenic activity. Therefore we believe that interaction of this side chain with the ER must play an important role in the interpretation of the ligand as an antiestrogen.

In order to examine issues of ligand discrimination by the ER, we have used site-directed and random chemical mutagenesis to generate ERs with selected changes in the hormone binding domain. We have been particularly interested in identifying residues in the hormone binding domain important for the ligand binding and transactivation functions of the receptor, and in elucidating the mechanism by which the ER discriminates between agonistic and antagonistic ligands. Although both estrogens and antiestrogens bind within the HBD, the association must differ because estrogen binding activates a transcriptional enhancement function, whereas antiestrogens fully or partially fail in this role. Our studies have indicated that selective changes near amino acid 380, and amino acids 520-530, and changes at the C-terminus of the ER result in ER ligand discrimination mutants [10, 13, 26]. These data provide evidence that some contact sites of the receptor with estrogen and antiestrogen differ; and that the conformation of the receptor with estrogen and antiestrogen must also be different as a consequence [10, 27 and refs therein]. Our structure-function analysis of the hormone binding domain of the human ER has utilized region-specific mutagenesis of the ER cDNA and phenotypic screening in yeast, followed by the analysis of interesting receptor mutants in mammalian cells [14, 28]. Our observations, as well as very important studies by Malcolm Parker and colleagues [29, 30] have shown a separation of the transactivation and hormone-binding functions of the ER.

Since the basic or polar side chain is essential for antiestrogenic activity, and our previous studies identified cysteine 530 as the amino acid covalently labeled by

affinity labeling ligands [31], we introduced by site directed mutagenesis of the ER cDNA changes of specific charged residues close to C530 [10]. Interestingly, two mutants in which lysines at position 529 and 531 were changed to glutamines, so that the local charge was changed, resulted in receptors with an approx. 30-fold increased potency of antiestrogen in suppressing estradiol-stimulated reporter gene activity. Interestingly, these mutants receptors showed a reduced binding affinity for estrogens, but retained unaltered binding affinity for antiestrogen. These findings suggest that we are able to differentially alter estrogen and antiestrogen effectiveness by rather modest changes in the ER, and that the region near C530 is a critical one for sensing the fit of the side chain of the estrogen antagonist. Studies from the Parker Laboratory [27] have shown that nearby residues (i.e. G525 and M521 and/or S522 in the mouse ER) are also importantly involved in conferring differential sensitivity to these two categories of ligands.

We have also shown that if C530 is mutated, the covalent ligand tamoxifen aziridine binds to C381 instead, another cysteine in the hormone binding domain [32]. One interpretation of this result is that the 530 and 380 regions of the hormone-binding domain are close to one another in the three-dimensional ligand binding pocket of the ER, such that the ligand can label either site by alternative positioning of the reactive side chain [32]. We therefore investigated charged amino acids in the N-terminal portion of the hormone binding domain and showed the region around amino acid 380 to be important in transcriptional activity of the receptor [13]. As opposed to what was observed with charge changes in the region near C530, we observed that change of the charged residue E380 to E380Q resulted in a receptor more sensitive to estrogen, but less sensitive than wild-type ER to antiestrogen for suppression of transcriptional activity. Although estrogen and antiestrogen showed no alteration of their binding affinity for the wild-type or E380Q mutant, the E380 mutant showed greater transcriptional activity and enhanced binding to estrogen response element DNA, resulting in its increased sensitivity to estrogen. Our findings suggest that this region is important in influencing DNA binding and protein-protein interaction of the receptor that modulates transcriptional activity and provide additional evidence, suggesting that the conformation of the receptor with estrogen and antiestrogen results in differential transactivation activity. Our recent data [26] has also shown that tamoxifen-like antiestrogens are more pure antiestrogens with the ER missing the C-terminal F domain, approx. the last 40 amino acids of the receptor. The basis for the difference in the estrogenic activity of tamoxifen-like estrogens with wild-type ER versus ER missing this F domain is under investigation and should provide important information regarding the differential agonistic/antagonistic effects of this category of antiestrogens.

ALTERATION IN THE AGONIST/ANTAGONIST BALANCE OF ANTIESTROGENS BY ACTIVATION OF PROTEIN KINASE A SIGNALING PATHWAYS: ANTIESTROGEN SELECTIVITY AND PROMOTER DEPENDENCE

There is increasing evidence for ER interaction with other cell signaling pathways. We became interested in this cross-talk between cell signaling pathways in our studies of estrogen regulation of the progesterone-receptor and estrogen responsive promoter-reporter gene constructs in cells. These studies showed stimulation by growth factors (IGF-1, EGF) as well as stimulation by cAMP and estrogen. The observation that the stimulation by these agents could be suppressed by antiestrogens or protein kinase inhibitors implied the involvement of the ER and phosphorylation pathways in these responses [18–21, 33]. We therefore have undertaken studies to examine directly whether activators of protein kinases can modulate transcriptional activity of the ER.

We find that activators of protein kinase A and protein kinase C markedly synergize with estradiol in ER-mediated transcriptional activation and that this transcriptional synergism shows cell- and promoter-specificity [15, 21, 34]. The synergistic stimulation of ER-mediated transcription by estradiol and protein kinase activators did not appear to result from changes in ER content or in the binding affinity of ER for ligand or the ERE DNA, but, rather, may be a consequence of a stabilization or facilitation of interaction of target components of the transcriptional machinery, possibly either through changes in phosphorylation of ER or other proteins important in ER-mediated transcriptional activation [34].

Figure 2 shows a model indicating how we think the protein kinase-ER transcriptional synergism may occur. Agents influencing protein kinase pathways may enhance intracellular protein phosphorylation resulting in either phosphorylation of the ER itself or the phosphorylation of nuclear factors with which the receptor interacts in mediating transcription. Likewise, there is evidence that the steroid hormone itself can alter

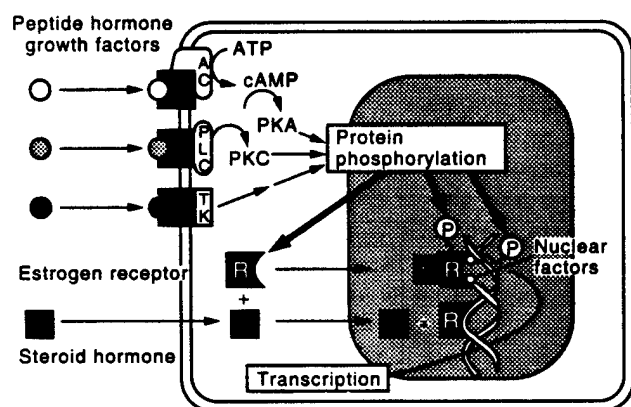


Fig. 2. Model depicting protein kinase-ER transcriptional synergism. See text for description.

Table 1. Levels of ligand-stimulated and protein kinase activator-stimulated phosphorylation of the human ER

Treatments	Phosphorylation level	
	mean \pm SE	n
Control	1	
10^{-9} M estradiol (E_2)	2.8 ± 0.3	3
10^{-8} M estradiol (E_2)	4.3 ± 0.7	6
10^{-8} M transhydroxytamoxifen (TOT)	2.9 ± 0.1	2
10^{-7} M ICI 164,384	3.6 ± 0.6	3
1 μ g/ml cholera toxin (CT) + 10^{-4} M isobutylmethylxanthine (IBMX)	1.9 ± 0.3	3
10^{-7} M TPA	2.6 ± 0.3	3

Human ER was expressed in COS-1 cells and transfected cells were incubated for 4 h with [32 P]orthophosphate in the presence of the indicated treatment. ER was immunoprecipitated with anti-receptor antibodies, resolved by SDS-PAGE and transferred to nitrocellulose. ER protein levels were determined by immunoblot and ER phosphorylation by autoradiography as described [35]. The levels of phosphorylation of the different samples were standardized according to ER protein levels and standard errors (SE) were calculated. 1 represents the basal level of phosphorylation (vehicle alone) in each experiment. n represents the number of experiments. (From Le Goff *et al.* ref. [35]).

receptor conformation increasing its susceptibility to serve as a substrate for protein kinases [19, 35–38 and Table 1]. Therefore, agents which increase the phosphorylation may, either through phosphorylation of the ER itself, or through phosphorylation of nuclear factors required for ER transcription, result in synergistic activation of ER-mediated transcription.

As shown in Fig. 3, we have compared the effects of cAMP on the transcriptional activity of the estradiol-liganded and antiestrogen-liganded ER complexes. We find that increasing the intracellular concentration of cAMP, or of protein kinase. A catalytic subunit of transfection [15], activates and/or enhances the transcriptional activity of type I but not type II antiestrogen-occupied ER complexes and reduces the estrogen antagonist activity of the type I transhydroxytamoxifen (TOT) antiestrogen. In Fig. 3(A and B), we have determined, in MCF-7 human breast cancer cells, the effect of cAMP on the activity of TOT, ICI 164,384 and E_2 on a simple TATA promoter with one consensus ERE upstream of the CAT gene and on the more complex pS2 gene promoter and 5'-flanking region (–3000 to +10) containing an imperfect ERE. The endogenous pS2 gene is regulated by E_2 in MCF-7 breast cancer cells. Estradiol increased the transcription of both of these gene constructs, and treatment with IBMX/CT and E_2 evoked a synergistic increase in transcription, with activity being ca 2.5 times that of E_2 alone. Both antiestrogens (TOT and ICI) failed to stimulate transactivation of these reporter gene constructs, but in the presence of IBMX/CT, TOT gave significant stimulation of transcription (85 or 60% that of E_2 alone). ICI failed to stimulate transactivation even in the presence of IBMX/CT, and ICI fully blocked E_2 stimulation in the presence or absence of cAMP. By contrast, treatment with IBMX/CT reduced the ability

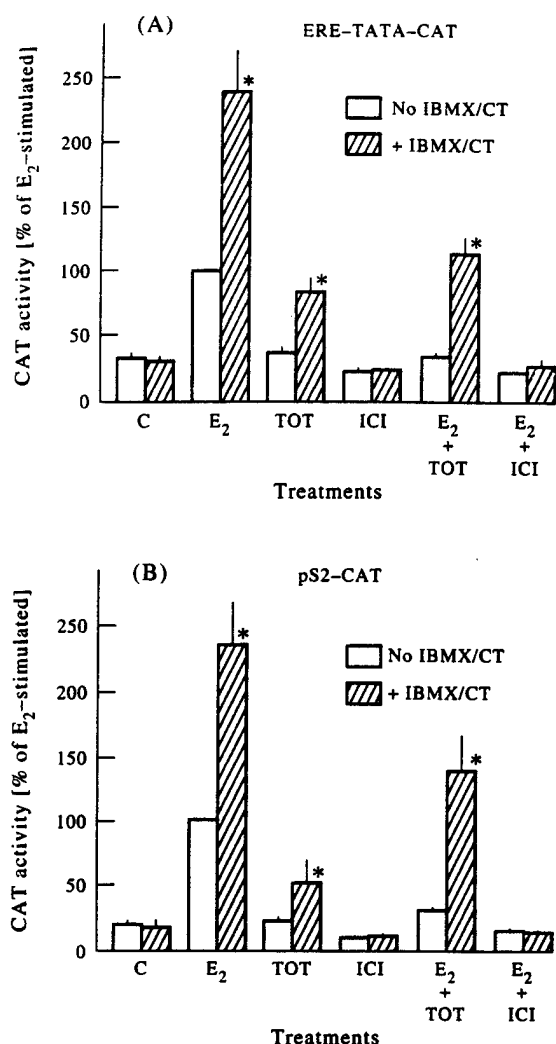


Fig. 3. Effect of IBMX/CT on the ability of E₂ and antiestrogens to stimulate transactivation of ERE-TATA-CAT (panel A) and pS2-CAT (panel B), and on the ability of antiestrogens to suppress E₂-stimulated transactivation. MCF-7 cells were transfected with the indicated reporter plasmid and an internal control plasmid that expresses β -galactosidase and were treated with the agents indicated for 24 h. Each bar represents the mean \pm SEM ($n = 3$ experiments). * Indicates significant difference from the no IBMX/CT cells ($P < 0.05$). C, control ethanol vehicle; E₂, 10^{-8} M; TOT (hydroxytamoxifen), 10^{-6} M; ICI (ICI 164,384), 10^{-6} M; IBMX (3-isobutyl-1-methyl-xanthine), 10^{-4} M; and CT (cholera toxin), 1 μ g/ml. (From Fujimoto and Katzenellenbogen, ref. [15]).

of TOT to inhibit E₂ transactivation. While TOT returned E₂ stimulation down to that of the control in the absence of IBMX/CT (compare open bars E₂ vs E₂ + TOT), TOT only partially suppressed the E₂ stimulation in the presence of IBMX/CT (compare stippled bars E₂ vs E₂ + TOT).

Although alteration in the agonist and antagonist activity of TOT was observed with promoter-reporter-constructs containing a simple TATA promoter and a more complex, pS2 promoter, elevation of cAMP did not enhance the transcription by either TOT or estradiol of the reporter plasmid ERE-thymidine kinase-CAT [15]. Thus, this phenomenon is promoter-specific.

Of note, cAMP and protein kinase A catalytic subunit transfection failed to evoke transcription by the more pure antiestrogen ICI 164,384 with any of the promoter-reporter constructs tested. These findings, which document that stimulation of the protein kinase A signaling pathway activates the agonist activity of tamoxifen-like antiestrogens, may in part explain the development of tamoxifen resistance by some ER-containing breast cancers. They also suggest that the use of antiestrogens like ICI 164,384, that fail to activate ER transcription in the presence of cAMP, may prove more effective for long-term antiestrogen therapy in breast cancer.

PHOSPHORYLATION OF THE ESTROGEN RECEPTOR

Since our data suggested that estrogens, and agents that activate protein kinases, might influence ER transcription by altering the state of phosphorylation of the ER and/or other factors required for ER regulation of transcription, we undertook studies to examine directly the effects of these agents on ER phosphorylation. In addition, we compared the effects of the type I and type II antiestrogens on phosphorylation of the ER (Table 1). Estradiol, each of the two antiestrogens, as well as protein kinase A and C activators enhanced overall ER phosphorylation, and in all cases, this phosphorylation occurred exclusively on serine residues [35]. Tryptic phosphopeptide patterns of wild-type and domain A/B-deleted receptors and site-directed mutagenesis of several serines involved in known protein kinase consensus sequences allowed us to identify serine 104 and/or serine 106 and serine 118, all three being part of a serine-proline motif, as major ER phosphorylation sites. Mutation of these serines to alanines so as to eliminate the possibility of their phosphorylation, resulted in an approx. 40% reduction in transactivation activity in response to estradiol while mutation of only one of these serines showed an approx. 15% decrease in activation [35]. Of note, estradiol and antiestrogen-occupied ERs showed virtually identical two-dimensional phosphopeptide patterns suggesting similar sites of phosphorylation. In contrast, the cAMP-stimulated phosphorylation likely occurs on different phosphorylation sites as indicated by some of our mutational studies [35] and this aspect remains under investigation in our laboratory.

cAMP-DEPENDENT SIGNALING PATHWAY INVOLVEMENT IN ACTIVATION OF THE TRANSCRIPTIONAL ACTIVITY OF ERs OCCUPIED BY TAMOXIFEN-LIKE BUT NOT ICI 164,384-LIKE ANTIESTROGENS

Our data provide strong evidence for the involvement of cAMP-dependent signaling pathways in the agonist actions of tamoxifen-like estrogen antagonists. The promoter-specificity of the transcriptional enhancement

phenomenon suggests that factors in addition to ER are probably being modulated by protein kinase A pathway stimulation. The findings imply that changes in the cAMP content of cells, which can result in activation of the agonist activity of tamoxifen-like antiestrogens, might account, at least in part, for the resistance to antiestrogen therapy that is observed in some breast cancer patients. Of interest, MCF-7 cells transplanted into nude mice fail to grow with tamoxifen treatment initially, but some hormone-resistant cells grow out into tumors after several months of tamoxifen exposure [8, 39, 40]. Studies have shown that this resistance to tamoxifen is, more correctly, a reflection of tamoxifen stimulation of proliferation, representing a change in the interpretation of the tamoxifen-ER complex and its agonist/antagonist balance. It is of interest that we found the pS2 gene, which is under estrogen and antiestrogen regulation in breast cancer [41], to be activated by tamoxifen in the presence of elevated cAMP. By contrast, however, antiestrogens such as ICI, shown in many systems to be more complete estrogen antagonists, are not changed in their agonist/antagonist balance by increasing intracellular concentrations of cAMP. Therefore, ICI-like compounds may prove to be more efficacious and less likely to result in antiestrogen-stimulated growth.

The transcriptional enhancement we have observed between protein kinase A activators and ER occupied by tamoxifen-like antiestrogens and estradiol provides further evidence for cross-talk between the ER and signal transduction pathways regulated by cAMP that are important in ER-dependent responses.

Acknowledgements—This work was supported by grants from the National Institutes of Health (CA18119), the U.S. Army Breast Cancer Research Program (DAMD17-94-J-4205), and the Susan G. Komen Foundation. We are especially grateful to several former associates of this laboratory for their important contributions to the studies reported here, in particular, Susan Aronica, Hyeseong Cho, Farzad Pakdel, Joseph Reese, and Carol Wrenn.

REFERENCES

- Katzenellenbogen B. S., Bhakoo H. S., Ferguson E. R., Lan N. C., Tatee T., Tsai T. L. and Katzenellenbogen J. A.: Estrogen and antiestrogen action in reproductive tissues and tumors. *Recent Prog. Horm. Res.* 35 (1979) 259–300.
- Katzenellenbogen B. S.: Dynamics of steroid hormone receptor action. *Ann. Rev. Physiol.* 42 (1980) 17–35.
- Sarrel P. M., Lufkin E. G., Oursler M. J., Keefe D.: Estrogen actions in arteries, bone and brain. *Sci. Am. Sci. Med.* 1 (1994) 44–53.
- Read L. D. and Katzenellenbogen B. S.: Characterization and regulation of estrogen and progesterone receptors in breast cancer. In *Genes, Oncogenes and Hormones: Advances in Cellular and Molecular Biology of Breast Cancer* (Edited by R. B. Dickson and M. E. Lippman). Kluwer-Nijhoff Publishers, Nowell, MA (1991) pp. 277–299.
- Kneifel M. A. and Katzenellenbogen B. S.: Comparative effects of estrogen and antiestrogen on plasma renin substrate levels and hepatic estrogen receptors in the rat. *Endocrinology* 108 (1981) 545–552.
- Toney T. W. and Katzenellenbogen B. S.: Antiestrogen action in the medial basal hypothalamus and pituitary of immature female rats: insights concerning relationships among estrogen, dopamine and prolactin. *Endocrinology* 119 (1986) 2661–2669.
- Katzenellenbogen B. S., Miller M. A., Mullick A. and Sheen Y. Y.: Antiestrogen action in breast cancer cells: modulation of proliferation and protein synthesis, and interaction with estrogen receptors and additional antiestrogen binding sites. *Breast Cancer Res. Treat.* 5 (1985) 231–243.
- Jordan V. C. and Murphy C. S.: Endocrine pharmacology of antiestrogens as antitumor agents. *Endocrine Res.* 11 (1990) 578–610.
- Santen R., Manni A., Harvey H. and Redmond C.: Endocrine treatment of breast cancer in women. *Endocrine Rev.* 11 (1990) 221–265.
- Pakdel F. and Katzenellenbogen B. S.: Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J. Biol. Chem.* 267 (1992) 3429–3437.
- Reese J. R. and Katzenellenbogen B. S.: Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor over-expressed in mammalian cells. *Nucl. Acids Res.* 19 (1991) 6595–6602.
- Reese J. C. and Katzenellenbogen B. S.: Characterization of a temperature-sensitive mutation in the hormone-binding domain of the human estrogen receptor: studies in cell extracts and intact cells and their implications for hormone-dependent transcriptional activation. *J. Biol. Chem.* 267 (1992) 9868–9873.
- Pakdel F., Reese J. C. and Katzenellenbogen B. S.: Identification of charged residues in an N-terminal portion of the hormone binding domain of the human estrogen receptor important in transcriptional activity of the receptor. *Molec. Endocr.* 7 (1993) 1408–1417.
- Wrenn C. K. and Katzenellenbogen B. S.: Structure-function analysis of the hormone binding domain of the human estrogen receptor by region-specific mutagenesis and phenotypic screening in yeast. *J. Biol. Chem.* 268 (1993) 24,089–24,098.
- Fujimoto N. and Katzenellenbogen B. S.: Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen-selectivity and promoter-dependence. *Molec. Endocr.* 8 (1994) 296–304.
- Berry M., Metzger D. and Chambon P.: Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the antioestrogen 4-hydroxytamoxifen. *EMBO J.* 9 (1990) 2811–2818.
- Tzukerman M. T., Esty A., Santiso-Mere D., Danielian P., Parker M. G., Stein R. B., Pike J. W. and McDonnell D. P.: Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Molec. Endocr.* 94 (1994) 21–30.
- Aronica S. M. and Katzenellenbogen B. S.: Progesterone receptor regulation in uterine cells: stimulation by estrogen, cyclic adenosine 3',5'-monophosphate and insulin-like growth factor I and suppression by antiestrogens and protein kinase inhibitors. *Endocrinology* 128 (1991) 2045–2052.
- Aronica M. and Katzenellenbogen B. S.: Stimulation of estrogen receptor-mediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic AMP and IGF-I. *Molec. Endocr.* 7 (1993) 743–752.
- Cho H., Aronica S. M. and Katzenellenbogen B. S.: Regulation of progesterone receptor gene expression in MCF-7 breast cancer cells: a comparison of the effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology* 134 (1994) 658–664.
- Kraus W. L., Montano M. M. and Katzenellenbogen B. S.: Cloning of the rat progesterone receptor gene 5' region and identification of two functionally distinct promoters. *Molec. Endocr.* 7 (1993) 1603–1616.
- Vignon F., Bouton M.-M. and Rochefort H.: Antiestrogens inhibit the mitogenic effect of growth factors on breast cancer cells in the total absence of estrogens. *Biochem. Biophys. Res. Commun.* 146 (1987) 1502–1508.
- Fawell S. E., White R., Hoare S., Sydenham M., Page M. and Parker M. G.: Inhibition of estrogen receptor-DNA binding by the "pure" antiestrogen ICI 164,384 appears to be mediated by impaired receptor dimerization. *Proc. Natn. Acad. Sci. U.S.A.* 87 (1990) 6883–6887.

24. Dauvois S., Danielian P. S., White R. and Parker M. G.: Antiestrogen ICI 164,384 reduces cellular estrogen receptor content by increasing its turnover. *Proc. Natn. Acad. Sci. U.S.A.* 89 (1992) 4037-4041.
25. Reese J. C. and Katzenellenbogen B. S.: Examination of the DNA binding abilities of estrogen receptor in whole cells: implications for hormone-independent transactivation and the action of the pure antiestrogen ICI 164,384. *Molec. Cell. Biol.* 12 (1992) 4531-4538.
26. Montano M. M., Muller V. and Katzenellenbogen B. S.: Role for the carboxy-terminal F domain of the estrogen receptor in transcriptional activity of the receptor and in the effectiveness of antiestrogens as estrogen antagonists. *Endocrinology* 134 (1994) 323.
27. Danielian P. S., White R., Hoare S. A., Fawell S. F. and Parker M. G.: Identification of residues in the estrogen receptor that confer differential sensitivity to estrogen and hydroxytamoxifen. *Molec. Endocr.* 7 (1993) 232-240.
28. Katzenellenbogen B. S., Bhardwaj B., Fang H., Ince B. A., Pakdel F., Reese J. C., Schodin D. J. and Wrenn C. K.: Hormone binding and transcription activation by estrogen receptors: analyses using mammalian and yeast systems. *J. Steroid Biochem. Molec. Biol.* 47 (1993) 39-48.
29. Danielian P. S., White R., Lees J. A. and Parker M. G.: Identification of a conserved region required for hormone-dependent transcriptional activation by steroid hormone receptors. *EMBO J.* 11 (1992) 1025-1033.
30. Fawell S. E., Lees J. A., White R. and Parker M. G.: Characterization and colocalization of steroid binding and dimerization activities in the mouse estrogen receptor. *Cell* 60 (1990) 953-962.
31. Harlow K. W., Smith D. N., Katzenellenbogen J. A., Greene G. L. and Katzenellenbogen B. S.: Identification of cysteine-530 as the covalent attachment site of an affinity labeling estrogen (ketononestrol aziridine) and antiestrogen (tamoxifen aziridine) in the human estrogen receptor. *J. Biol. Chem.* 264 (1989) 17,476-17,485.
32. Reese J. C., Wooge C. H. and Katzenellenbogen B. S.: Identification of two cysteines closely positioned in the ligand binding pocket of the human estrogen receptor: roles in ligand binding and transcriptional activation. *Molec. Endocr.* 6 (1992) 2160-2166.
33. Katzenellenbogen B. S. and Norman M. J.: Multihormonal regulation of the progesterone receptor in MCF-7 human breast cancer cells: interrelationships among insulin/IGF-I, serum and estrogen. *Endocrinology* 126 (1990) 891-898.
34. Cho H. and Katzenellenbogen B. S.: Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Molec. Endocr.* 7 (1993) 441-452.
35. Le Goff P., Montano M. M., Schodin D. J. and Katzenellenbogen B. S.: Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J. Biol. Chem.* 269 (1994) 4458-4466.
36. Ali S., Metzger D., Bornert J. and Chambon P.: Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J.* 12 (1993) 1153-1160.
37. Denton R. R., Koszewski N. J. and Notides A. C.: Estrogen receptor phosphorylation: hormonal dependence and consequence on specific DNA binding. *J. Biol. Chem.* 267 (1992) 7263-7268.
38. Lahooti H., White R., Danielian P. S. and Parker M. G.: Characterization of ligand-dependent phosphorylation of the estrogen receptor. *Molec. Endocr.* 8 (1994) 182-188.
39. Osborne C. K., Coronado E., Allred D. C., Wiebe V. and DeGregorio M.: Acquired tamoxifen resistance: correlation with reduced breast tumor levels of tamoxifen and isomerization of trans-4-hydroxytamoxifen. *J. Natn. Cancer Inst.* 83 (1991) 1477-1482.
40. Gottardis M. M. and Jordan V. C.: Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res.* 48 (1988) 5183-5187.
41. Brown A. M. C., Jeltsch J. M., Roberts M. and Chambon P.: Activation of pS2 gene transcription is a primary response to estrogen in the human breast cancer cell line MCF-7. *Proc. Natn. Acad. Sci. U.S.A.* 81 (1984) 6344-6348.

Repression of Endogenous Estrogen Receptor Activity in MCF-7 Human Breast Cancer Cells by Dominant Negative Estrogen Receptors*

B. AVERY INCE, DAVID J. SCHODIN, DAVID J. SHAPIRO, AND
BENITA S. KATZENELLENBOGEN

Departments of Cell and Structural Biology (B.A.I., B.S.K.), Physiology and Biophysics (D.J.Sc., B.S.K.), and Biochemistry (D.J.Sh.), University of Illinois, Urbana, Illinois 61801

ABSTRACT

We have investigated the ability of several transcriptionally inactive estrogen receptor (ER) mutants to block endogenous ER-mediated transcription in MCF-7 human breast cancer cells. In transient transfections of MCF-7 cells, two of the mutants, a frame-shifted ER (S554fs) and a point-mutated ER (L540Q), strongly inhibit the ability of endogenous wild-type ER to activate transcription of estrogen-regulated reporter plasmids. A third mutant, ER1-530, which is missing 65 residues from its carboxy-terminus, is a weaker repressor of estradiol-stimulated transcription. When an estrogen response element (ERE)-thymidine kinase-chloramphenicol acetyltransferase reporter gene is used, S554fs, L540Q, and ER1-530 suppress the transcriptional activity of endogenous MCF-7 ER by 87%, 97%, and 62%, respectively. The magnitude of dominant negative repression is promoter specific; when an ERE-pS2-chloramphenicol acetyltransferase reporter is employed, inhibition of endogenous ER activity by equivalent amounts of S554fs, L540Q, and ER1-530 ranges from 85–97%.

Dose-response studies show the S554fs mutant to be the most potent of the three ER mutants as a repressor of estrogen action in these cells. In addition, elevated levels of intracellular cAMP, achieved by the addition of 3-isobutyl-1-methylxanthine plus cholera toxin to cells, fail to compromise the effectiveness of these mutants as dominant negative ERs despite the cAMP-enhanced transcriptional activity of ER. The mutants are also powerful repressors of the agonist activity of *trans*-hydroxytamoxifen-stimulated ER transcription. The dominant negative activity of the three mutants is lost when the A/B domain of these receptors is deleted, implying an important role for this N-terminal region of the ER in the ability of these mutants to inhibit endogenous wild-type ER activity. All in all, the data suggest that S554fs in particular is a reasonable candidate for studies designed to use a dominant negative ER to inhibit the estrogen- and tamoxifen-stimulated growth of human breast cancer cells. (*Endocrinology* 136: 3194–3199, 1995)

THE GROWTH of nearly 40% of all human breast tumors is highly dependent upon the sex steroid hormone, estrogen (1–3). As the proliferative effect of estrogens on breast cancer cells is mediated by the estrogen receptor (ER), there is much interest in exploring the means by which this protein can be functionally inactivated. We are currently investigating the possibility of eventually employing dominant negative ER mutants to block wild-type ER-mediated transcription and growth stimulation in estrogen-dependent breast cancer cells.

The ER, which belongs to the conserved superfamily of steroid and thyroid receptors, is a nuclear regulatory protein that functions as a hormone-activated transcription factor in target cells (4, 5). Receptor activation is apparently a consequence of ligand-induced conformational changes in ER structure (6). The hormone-receptor complex binds with high affinity to a well defined palindromic nucleotide sequence, the estrogen response element (ERE), which is usually

located upstream of an estrogen-responsive gene (7, 8). It appears that activated receptors recruit transcription factors and establish transcriptionally productive protein-protein interactions with other components of the transcription machinery (9–11). Current attempts to functionally inactivate the ER in *in vivo* and *in vitro* experimental systems and in actual breast cancer therapy employ the antiestrogen, tamoxifen. Tamoxifen binds to the ER and is thought to induce a conformational change that renders the receptor virtually incapable of activating transcription of genes involved in cancer cell proliferation and tumorigenesis (12). Administered antiestrogens have been found, however, to retain estrogenic activity in certain tissues, including some cancerous mammary tissues (13). We wanted to explore the feasibility of employing dominant negative ER mutants to suppress ER-mediated transcription, whether 17 β -estradiol (E₂) or tamoxifen stimulated, in estrogen-responsive breast cancer cells.

Dominant negative mutants of a protein, when coexpressed with the wild-type version, block the action of the parent protein (14–16). Our group previously reported the successful generation of three dominant negative ER mutants and their characterization in ER-deficient Chinese hamster ovary (CHO) cells (17). In these experiments, we investigated the effectiveness of the reported mutants as inhibitors of endogenous ER in an E₂-stimulated human breast cancer cell line. We also examined the issue of dominant negative inhibition of tamoxifen-stimulated ER

Received December 13, 1994.

Address all correspondence and requests for reprints to: Dr. Benita S. Katzenellenbogen, Department of Physiology and Biophysics, University of Illinois, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, Illinois 61801.

* This work was supported by NIH Grant CA-60514 (to B.S.K. and D.J.S.), U.S. Army Grant DAMD17-94-J-4205 (to B.S.K.), a National Science Foundation Graduate Fellowship and a Ford Foundation Dissertation Fellowship (to B.A.I.), and in part by Postdoctoral Fellowship HD-7028 (to D.J.Sc.).

transcription, assessed the ability of the ER mutants to repress estrogen action in the presence of elevated levels of intracellular cAMP, and examined the role of the N-terminal portion of the ER in dominant negative ER activity. These studies should prove informative in efforts to identify ER mutants that can plausibly be employed in future efforts to antagonize the estrogen- and tamoxifen-stimulated growth of human breast cancer cells.

Materials and Methods

Chemicals and materials

Cell culture media and sera were purchased from Gibco (Grand Island, NY). Radioinert E_2 , 3-isobutyl-1-methylxanthine (IBMX), cholera toxin (CT), and chloramphenicol were obtained from Sigma Chemical Co. (St. Louis, MO). The antiestrogen *trans*-4-hydroxytamoxifen (TOT) was provided by ICI Pharmaceuticals (Macclesfield, UK). [3H]Acetyl coenzyme A (1 mCi/ml) was obtained from DuPont-New England Nuclear (Boston, MA).

Plasmids

For transcriptional activation studies, the estrogen-responsive plasmids ERE-tk-chloramphenicol acetyltransferase (CAT) (18) and (ERE) $_2$ -pS2-CAT were employed. (ERE) $_2$ -pS2-CAT was constructed by W. Lee Kraus of this laboratory by cloning two copies of a consensus estrogen-responsive element into the *Bam*HI site of pS2-CAT (19). Mutant human ER complementary DNAs subcloned into the eukaryotic expression vector pCMV5 (CMV = cytomegalovirus) (20) were used to express ER mutants in transfected cells. The plasmid pCH110 (Pharmacia LKB Biotechnology, Piscataway, NJ), which contains the β -galactosidase gene, was used as an internal control for transfection efficiency in all experiments. The plasmid pTZ19, used as carrier DNA, was provided by Dr. Byron Kemper of the University of Illinois.

ER mutagenesis and expression of mutant receptors in cells

S554fs, L540Q, and ER1-530 were generated as previously described (21). The M7 mutant K520D/G521V/E523R/H524L was described previously (17). Complementary DNAs encoding the N-terminal-truncated ($\Delta A/B$) versions of these mutants were generated by replacing the *Hind*III fragment of these full-length mutants with the *Hind*III fragment of CMV- $\Delta A/B$ hER [which deletes nucleotides from the CMV-5 polylinker (22) to codon 176]. The resultant expression vectors contain the human ER-coding region from amino acids 176-595 and produce human ER derivatives that are deleted of residues N-terminal to Met¹⁷⁶ in the ER primary sequence. Although we could not accurately determine levels of expression in MCF-7 cells for the mutant receptors ($\Delta A/B$ dominant negative ERs, S554fs, L540Q, ER1-530, and M7) because of the small percentage of cells transfected and because many of these receptors are indistinguishable on Western blots from endogenous MCF-7 ER, we did compare expression levels in CHO cells. We found comparable levels of these receptors made when equal amounts of expression plasmids were transfected (as reported in Refs. 17, 21, and 23, where expression levels for many of these mutants were determined). We also observed that the $\Delta A/B$ dominant negative ERs and $\Delta A/B$ wild-type ER were expressed at similar levels after transfection into MCF-7 cells.

Cell culture and transient transfections

MCF-7 human breast cancer cells were maintained in Eagle's Minimum Essential Medium (MEM; Gibco, Grand Island, NY) supplemented with 5% calf serum (Hyclone, Logan, UT), 25 μ g/ml gentamycin, 100 U/ml penicillin (Gibco), and 100 μ g/ml streptomycin (Gibco). Before the experiments, cells were maintained for 1 week in MEM containing the above antibiotics and 5% charcoal dextran-treated calf serum (CDCS); they were then cultured for 1 week in phenol red-free MEM with 5% CDCS and the same antibiotics. Transient transfections were performed as follows. Cells were plated at about 4×10^6 cells/100-mm dish, main-

tained at 37°C in a humidified 5% CO $_2$ atmosphere for roughly 48 h, and transfected by the CaPO $_4$ coprecipitation method (24). In transactivation assays, 100-mm plates were treated with 1.0 ml DNA precipitate containing 2.0 μ g reporter plasmid, 3.0 μ g pCH110 internal control plasmid, 0.2-10 μ g ER or ER mutant expression vector, and up to 9 μ g pTZ carrier DNA. In all cases, cells remained in contact with the precipitate for 4-6 h and were then subjected to a 3-min glycerol shock (25% in MEM plus 5% CDCS). Plates were rinsed, given fresh medium, and treated with E_2 , TOT, E_2 plus IBMX/CT, or ethanol vehicle as appropriate. Cells were harvested after 24 h, and extracts were prepared in 250 μ l 250 mM Tris, pH 7.5, using three freeze-thaw cycles. β -Galactosidase activity was measured (25) to normalize for transfection efficiency among plates. CAT assays were performed as previously described (26).

Results

ER mutants S554fs and L540Q are potent repressors of E_2 -stimulated endogenous ER activity

Three ER mutants were selected for study because they had previously exhibited strong dominant negative activity in transfected CHO cells (17). The mutants, generated by random chemical mutagenesis, include a frame shift (S554fs), a point mutation (L540Q), and a truncated receptor (ER1-530) (21). MCF-7 cells were transfected with either the ERE-tk-CAT or (ERE) $_2$ -pS2-CAT reporter plasmid in addition to expression vector for the ER mutant under examination. CAT activity in response to a saturating dose of E_2 (10^{-9} M) was then measured for each mutant studied. The data in Fig. 1 indicate dramatic differences in resultant CAT activity between MCF-7 cells into which no ER mutants were introduced and those transfected with dominant negative ERs. Whereas endogenous MCF-7 ER exhibited a 70-fold induction of transcriptional activity (set at 100%) from an ERE-tk-CAT reporter in response to 10^{-9} M E_2 , cells transfected with 10 μ g expression vector for S554fs, L540Q, and ER1-530 exhibited 87%, 97%, and 62% repressions of E_2 -induced transcription, respectively (Fig. 1). Lesser amounts of expression vector for each mutant were used in an attempt to gauge their relative potencies as dominant negative inhibitors. These studies showed S554fs to be the most potent of the three ER mutants in inhibiting E_2 -induced transcriptional activity in MCF-7 cells (Fig. 1). When a reporter gene containing the pS2 promoter, (ERE) $_2$ -pS2-CAT, was used in similar experiments, E_2 stimulated a 30-fold increase in MCF-7 ER transcriptional activity, and 10 μ g expression vector for S554fs, L540Q, and ER1-530 repressed ER-mediated transcription by 90%, 97%, and 85%, respectively (Fig. 2). Comparative studies with lesser amounts of the three mutants again showed S554fs to be the most potent of the three. Another ER mutant, K520D/G521V/E523R/H524L (M7), which was previously determined to be transcriptionally inactive and to show only modest ER inhibitory activity in CHO cells (17, 27), was assayed for dominant negative activity in the MCF-7 cell system. Consistent with its weak dominant negative activity in CHO cells, the M7 mutant failed to inhibit ER-mediated transcription from either reporter gene employed in this study when transfected at the 1.5 μ g expression plasmid level (Figs. 1 and 2), whereas it demonstrated some suppressive activity at the 10- μ g plasmid concentration, but always much less than that of the three dominant negative mutants. Transfection of 10 μ g of the empty vector pCMV5 had no effect on endogenous MCF-7 ER activity (data not shown).

FIG. 1. Dose-response analysis of the ability of ER mutants to block E_2 -stimulated transcriptional activity of endogenous ER. MCF-7 cells were cotransfected with the reporter plasmid ERE-tk-CAT; the indicated amounts of expression vector for the ER mutants S554fs, L540Q, ER1-530, and M7; and a β -galactosidase internal reporter to correct for transfection efficiency. Two tenths to 10 μ g mutant ER expression vector were employed. Cells were treated with 10^{-9} M E_2 for 24 h. Extracts were prepared and analyzed for β -galactosidase and CAT activity as described in *Materials and Methods*. The magnitude of wild-type (MCF-7) ER activation by E_2 alone was set at 100%. Error bars represent the range (n = 2 experiments) or SEM (n = 3–6 experiments). Each value from an experiment is the average of duplicate determinations from two plates of cells.

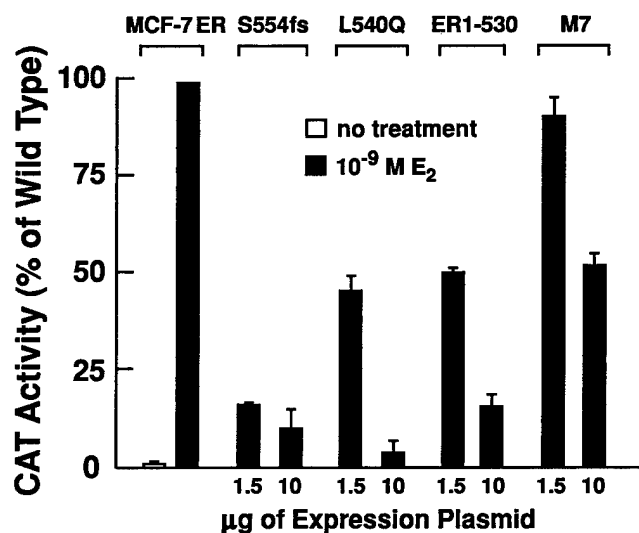
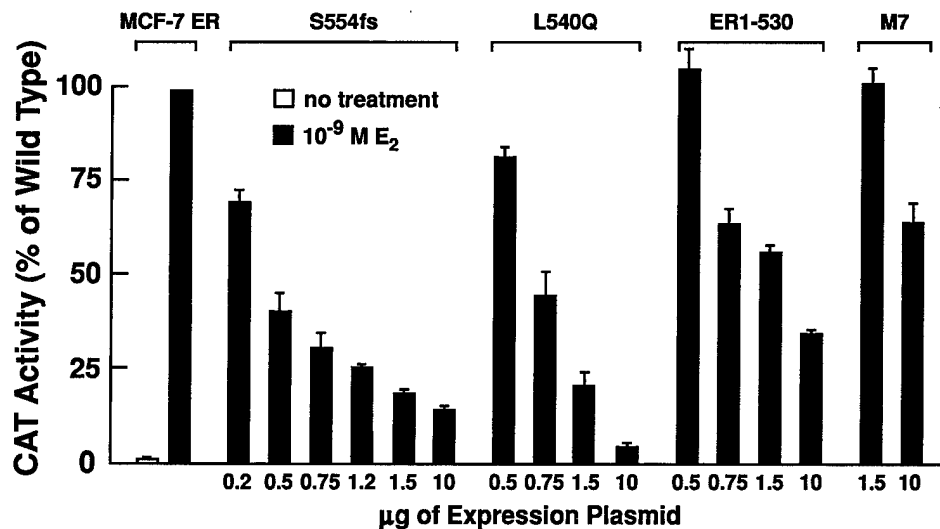


FIG. 2. Examination of the ability of ER mutants to block E_2 -stimulated endogenous ER transcriptional activity from a reporter plasmid containing the pS2 promoter. MCF-7 cells were cotransfected with the reporter plasmid (ERE)₂-pS2-CAT; 1.5 or 10 μ g expression vector for the ER mutants S554fs, L540Q, ER1-530, and M7; and a β -galactosidase internal reporter to correct for transfection efficiency. Cells were treated with 10^{-9} M E_2 for 24 h. Extracts were prepared and analyzed for β -galactosidase and CAT activity as described in *Materials and Methods*. The magnitude of wild-type ER activation by E_2 alone was set at 100%. Error bars represent the range (n = 2 experiments) or SEM (n = 3–6 experiments). Each value from an experiment is the average of duplicate determinations from two plates of cells.

Dominant negative mutants strongly antagonize tamoxifen-stimulated transcription

We next examined whether the ER mutants were capable of inhibiting TOT-stimulated transcription. TOT treatment of MCF-7 cells resulted in a 9-fold induction of ER-mediated transcription, i.e. a response about 30% of that elicited by E_2 (Fig. 3). This activity was almost completely eliminated in cells containing any of the transfected dom-

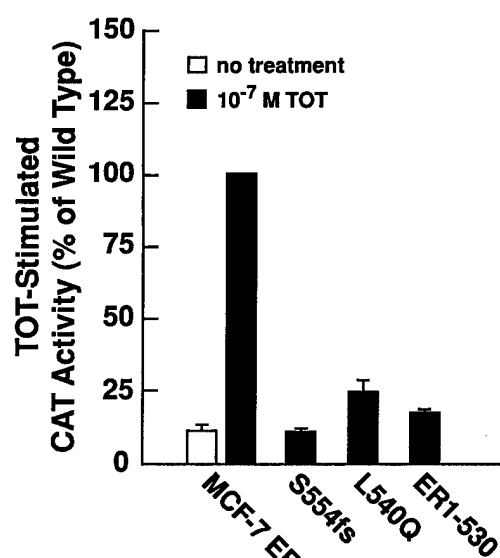


FIG. 3. Examination of the ability of ER mutants to block TOT-stimulated transcriptional activity of endogenous ER. MCF-7 cells were cotransfected with the (ERE)₂-pS2-CAT reporter plasmid; 0.2 μ g expression vector for the ER mutants S554fs, L540Q, and ER1-530; and a β -galactosidase internal reporter to correct for transfection efficiency. Cells were treated with 10^{-7} M TOT for 24 h. Extracts were prepared and analyzed for β -galactosidase and CAT activity as described in *Materials and Methods*. The magnitude of wild-type ER activation by TOT alone (8-fold) was set at 100%. Error bars represent the range (n = 2 experiments) or SEM (n = 3 experiments). Each value from an experiment is the average of duplicate determinations from two plates of cells.

inant negative mutants. A low amount (0.2 μ g) of expression vector for S554fs, L540Q, and ER1-530 suppressed 100%, 84%, and 93% of TOT-stimulated transcription, respectively (Fig. 3). Thus, the stimulatory activity of the TOT-occupied MCF-7 ERs appeared to be even more effectively suppressed by the dominant negative ER mutants than was that of the E_2 -occupied receptors.

S554fs and L540Q function well as dominant negative receptors in the presence of elevated intracellular cAMP

Recent reports have documented the ability of protein kinase A activators to increase ligand-stimulated transactivation by steroid receptors, including ER (18, 23, 28–32). As such, the ability of the mutant ERs to antagonize ER-mediated transcription in the presence of high levels of intracellular cAMP was assessed by treating transfected MCF-7 cells not only with E_2 , but also with IBMX/CT, agents that have been shown to elevate intracellular cAMP in these cells (33). Although there was a strong induction of ER-mediated transcriptional activity from the ERE-tk-CAT reporter gene in response to E_2 treatment (set at 100%), this was elevated consistently (~1.4-fold) when IBMX/CT was also administered to transfected cells. Exposure to IBMX/CT alone had little effect on MCF-7 ER activity. When 0.75 μ g expression plasmid for each of the dominant negative mutants was introduced into E_2 - plus IBMX/CT-treated MCF-7 cells, S554fs, L540Q, and ER1-530 achieved repressions of 87%, 88%, and 61%, respectively (Fig. 4A, ▨). These levels of inhibition compare favorably to those achieved in the absence of elevated intracellular cAMP and were, in fact, slightly greater. Similar experiments (Fig. 4B) were conducted using the (ERE)₂-pS2-CAT reporter gene; E_2 plus IBMX/CT exposure elicited a stimulation of MCF-7 ER CAT activity 2.2-fold that evoked by E_2 alone. Once again, repression of ER activity by the dominant negative mutants in the absence of increased levels of intracellular cAMP was almost identical to that in the presence of added IBMX/CT (Fig. 4B). The experiments suggest that the presence of high levels of cAMP does not impair the ability of these mutants to act as strong dominant negative inhibitors of ER action despite the cAMP-stimulated enhancement of ER transcriptional activity.

ER mutants deleted of their N-terminal transactivation function lose the dominant negative phenotype

The dominant negative ER mutants contain the entire A/B regions of the receptor and, therefore, have an intact N-terminal transactivation (AF-1) domain. These AF-1 regions, which are widely thought to be hormone independent (34), might confer upon the mutants some intrinsic ability to activate transcription, thereby reducing their dominant negative inhibitory action. In an attempt to further increase the potency of the ER mutants as dominant negative ER inhibitors, we deleted the first 175 residues at their N-terminals and, therefore, removed their AF-1 transactivation functions. We then transfected MCF-7 cells with these truncated ER mutants and compared their abilities to function as dominant negative ER repressors with those of the full-length dominant negative mutants. Although 0.5 μ g expression vector for S554fs and L540Q achieved 60% and 20% repression of transcriptional activity, and 1.5 μ g expression vector for S554fs and L540Q achieved 80–85% repression of transcriptional activity, equivalent amounts of Δ A/B S554fs and Δ A/B L540Q showed little ability to repress E_2 action (Fig. 5). The ER1-530 mutant, although the least effective of the three dominant negative receptors, also became less effective in suppressing endogenous ER activity when present in the

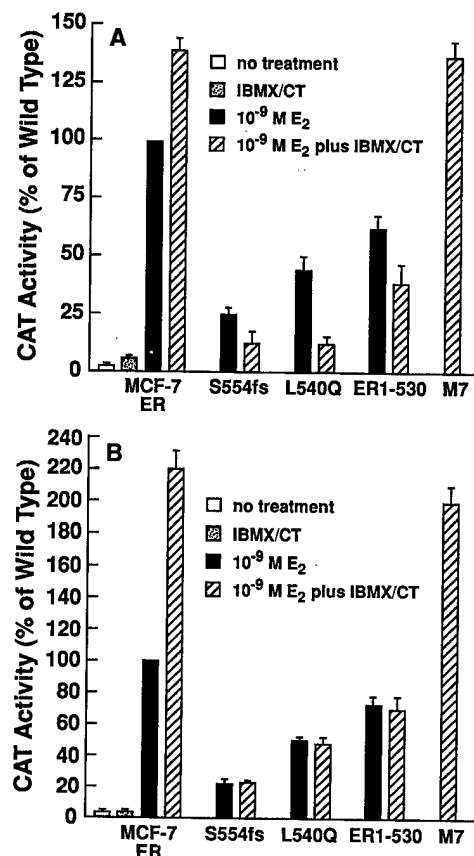


FIG. 4. Examination of the ability of ER mutants to block E_2 -stimulated transcriptional activity of endogenous ER in the presence of elevated intracellular cAMP. MCF-7 cells were cotransfected with the ERE-tk-CAT reporter plasmid; 0.75 μ g expression vector for the ER mutants S554fs, L540Q, ER1-530, and M7; and a β -galactosidase internal reporter to correct for transfection efficiency (A) or the (ERE)₂-pS2-CAT reporter plasmid, 1.5 μ g mutant ER expression vector, and a β -galactosidase internal reporter (B). Cells were treated with IBMX/CT alone, E_2 alone, or 10^{-9} M E_2 and 10^{-4} M IBMX plus 1 μ g/ml CT for 24 h. Extracts were prepared and analyzed for β -galactosidase and CAT activities as described in *Materials and Methods*. The magnitude of wild-type ER activation by E_2 alone was set at 100%, and all values (with and without IBMX/CT exposure) are expressed as a percentage of the value for wild-type ER plus E_2 alone. Error bars represent the range ($n = 2$ experiments) or SEM ($n = 3$ experiments). Each value from an experiment is the average of duplicate determinations from two plates of cells.

truncated (Δ A/B) form (Fig. 5). Using 1.5 μ g expression vector, the ER1-530 mutant achieved a 45% repression of endogenous ER activity; the repression was reduced to 15% for the Δ A/B ER1-530 mutant. As such, deletion of the AF-1 transactivation domain from these ER mutants not only failed to increase their potency as dominant negative ER repressors, but it also destroyed their ability to function as effective inhibitors of ER action.

Discussion

We report that two human ER mutants, S554fs and L540Q, are potent dominant negative inhibitors of endogenous ER transcriptional activity in MCF-7 human breast cancer cells. A third mutant, ER1-530, is a weaker repres-

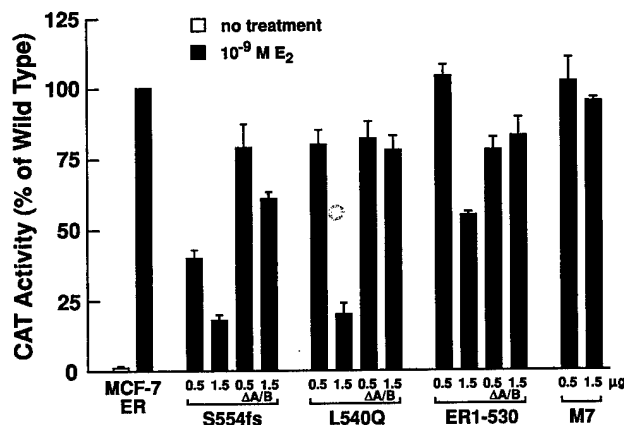


FIG. 5. Examination of the ability of $\Delta\Delta/B$ ER mutants to block E_2 -stimulated transcriptional activity of endogenous ER. MCF-7 cells were cotransfected with the ERE-tk-CAT reporter plasmid, a β -galactosidase internal reporter to correct for transfection efficiency, and 0.5 or 1.5 μ g expression vector for the ER mutants S554fs, $\Delta\Delta/B$ S554fs, L540Q, $\Delta\Delta/B$ L540Q, ER1-530, $\Delta\Delta/B$ ER1-530, and M7. Cells were treated with 10^{-9} M E_2 for 24 h. Extracts were prepared and analyzed for β -galactosidase and CAT activities as described in *Materials and Methods*. The magnitude of wild-type ER activation by E_2 alone was set at 100%. Error bars represent the range ($n = 2$ experiments) or SEM ($n = 3$ experiments). Each value from an experiment is the average of duplicate determinations from two plates of cells.

sor of ER action in this cell line. As S554fs has previously been shown to bind to ERE DNA with a lower affinity than that of wild-type ER (17), its relatively high potency as a dominant negative ER in MCF-7 cells may arise from an ability to form heterodimers with the wild-type ER, which are transcriptionally compromised. Alternatively, it could be the result of a greater ability on the part of S554fs to sequester cellular factors with which wild-type ER interacts to activate transcription. Transcriptional inactivity alone is not sufficient to confer a strong dominant negative phenotype, however, because the ER mutant M7 was not an effective repressor of MCF-7 ER activity at concentrations (0.5 or 1.5 μ g) at which the dominant negative ER mutants showed suppressive activity. At higher plasmid concentrations (10 μ g), M7 showed some suppressive activity, consistent with its ability to act as an ER-selective inhibitor at high concentrations (17, 27).

S554fs, L540Q, and ER1-530 all proved to be extremely effective inhibitors of TOT-stimulated ER activity. It is possible that the conformation of wild-type ER when bound by TOT (6, 10, 35) may lend the receptor to easy suppression not only by S554fs and L540Q, but also by ER1-530.

Given reports documenting the ability of protein kinase A activators to increase ligand-stimulated transactivation by ER (23, 28) as well as recent studies by us demonstrating the occasional transcriptional activation of the ER mutants S554fs and L540Q in some cell and promoter contexts by a combination of estrogen or antiestrogen ligands and agents that elevate intracellular cAMP (18), we assessed the ability of the mutant ERs to antagonize ER-mediated transcription in the presence of high levels of intracellular cAMP. When the dominant negative ER mutants were introduced into E_2 - and IBMX/CT-treated MCF-7 cells, S554fs, L540Q, and

ER1-530 achieved repressions of 87%, 88%, and 61%, respectively, which compare favorably with those achieved in the absence of elevated intracellular cAMP. As it is now clear that cell and promoter context markedly influence transcriptional activation by the ER (34, 36) and other steroid and thyroid hormone receptors (37, 38), it is possible that elevated levels of cAMP in MCF-7 cells modulate either the conformation or the activity of wild-type ER, the mutant ERs, or cellular factors with which they interact, so as to maintain or even enhance the dominant negative effects seen.

Of note, we observed that deletion of the N-terminal A/B domain of the dominant negative receptors, which contains the AF-1 transactivation region, rendered them ineffective. Therefore, it appears that the N-terminal region of the ER, which is known to interact with other cellular factors (34, 36), is necessary for the ER mutants to function as dominant negative inhibitors. This raises the distinct possibility that the mutants may need to interact with cellular factors other than the ER to achieve their inhibitory effects and is consistent with the promoter dependence of the dominant negative phenomenon. For example, the mutants, especially ER1-530, differed somewhat in their effectiveness in suppressing MCF-7 ER activity on the estrogen-responsive tk *vs.* pS2 promoter gene constructs studied. On the other hand, the possibility that the N-terminal-truncated ER mutants may be impaired in some other function, such as dimerization, cannot be formally discounted, and experiments exploring these issues are being undertaken.

Recent studies have revealed the presence of ER variants, some demonstrating dominant negative activity, in breast cancers (1). These naturally occurring variants are truncated receptors due to the deletion of exon 3 (39) or exon 7 (40). Their role in modulating the response of wild-type ER to endocrine therapies is an issue of great interest. Our studies indicate that potent dominant negative ER mutants can markedly suppress the activity of the endogenous wild-type ER in breast cancer cells.

In summary, ER mutants S554fs and L540Q seem to be potent repressors of ligand-stimulated transcriptional activity in MCF-7 cells. Although cAMP significantly elevates wild-type ER-mediated transcriptional activity, the presence of elevated levels of intracellular cAMP does not seem to thwart the ability of any of these mutants to function as dominant negative ER suppressors in MCF-7 cells; in fact, in these cells, it sometimes appeared to enhance their inhibitory function slightly. The results, taken as a whole, strongly suggest the suitability of these ER mutants for further experiments aimed at suppressing not only the ligand-induced transcriptional activity of ER in MCF-7 human breast cancer cells, but also the stimulation of cell growth and proliferation.

Acknowledgments

We thank Dr. W. Lee Kraus of this laboratory for the ERE-tk-CAT and (ERE)₂-pS2-CAT expression plasmids, Yao Zhuang for the M7 ER mutant, and Anne Keller, William Schelman, and Angela Trobaugh for their contributions to parts of this work.

References

- McGuire WL, Chamness GC, Fuqua SAW 1991 Estrogen receptor variants in clinical breast cancer. *Mol Endocrinol* 5:1571-1577
- Osborne CK, Yochmowitz MG, Knight III WA, McGuire WL 1980 The value of estrogen and progesterone receptors in the treatment of breast cancer. *Cancer* 46:2884-2888
- Santen R, Manni A, Harvey H, Redmond C 1990 Endocrine treatment of breast cancer in women. *Endocr Rev* 11:221-265
- Evans RM 1988 The steroid and thyroid hormone receptor superfamily. *Science* 240:889-895
- Beato M 1989 Gene regulation by steroid hormones. *Cell* 56:335-344
- Beekman JM, Allan GF, Tsai SY, Tsai M-J, O'Malley BW 1993 Transcriptional activation by the estrogen receptor requires a conformational change in the ligand binding domain. *Mol Endocrinol* 7:1266-1274
- Kumar V, Chambon P 1988 The estrogen receptor binds tightly to its responsive element as a ligand-induced homodimer. *Cell* 55:145-156
- Carson-Jurica MA, Schrader W, O'Malley B 1990 Steroid receptor family: structure and functions. *Endocr Rev* 11:201-220
- Tsai MJ, O'Malley BW 1994 Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 63:451-486
- Meyer ME, Gronemeyer H, Turcotte B, Bocquel M, Tasset D, Chambon P 1989 Steroid hormone receptors compete for factors that mediate their enhancer function. *Cell* 57:433-442
- Pham TA, Elliston JF, Nawaz Z, McDonnell DP, Tsai MJ, O'Malley BW 1991 Antiestrogen can establish nonproductive receptor complexes and alter chromatin structure at target enhancers. *Proc Natl Acad Sci USA* 88:3125-3129
- McDonnell DP, Nawaz Z, O'Malley BW 1991 *In situ* distinction between steroid receptor binding and transactivation at a target gene. *Mol Cell Biol* 11:4350-4355
- Katzenellenbogen BS 1991 Antiestrogen resistance: mechanisms by which breast cancer cells undermine the effectiveness of endocrine therapy. *J Natl Cancer Inst* 83:1434-1435
- Herskowitz I 1987 Functional inactivation of genes by dominant negative mutations. *Nature* 329:219-222
- Damm K, Thompson C, Evans R 1989 Protein encoded by v-erb A functions as a thyroid hormone receptor antagonist. *Nature* 339:593-597
- Privalsky M, Sharif M, Yamamoto K 1990 The viral erb A oncogene protein, a constitutive repressor in animal cells, is a hormone-regulated activator in yeast. *Cell* 63:1277-1286
- Ince BA, Zhuang Y, Wrenn CK, Shapiro DJ, Katzenellenbogen BS 1993 Powerful dominant negative mutants of the human estrogen receptor. *J Biol Chem* 268:14026-14032
- Ince BA, Montano MM, Katzenellenbogen BS 1994 Activation of transcriptionally inactive human estrogen receptors by cyclic adenosine 3',5'-monophosphate and ligands including antiestrogens. *Mol Endocrinol* 8:1397-1406
- Aronica SM, Kraus WL, Katzenellenbogen BS 1994 Estrogen action via the cAMP signaling pathway: stimulation of adenylate cyclase and cAMP-regulated gene transcription. *Proc Natl Acad Sci USA* 91:8517-8521
- Reese JC, Katzenellenbogen BS 1991 Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor overexpressed in mammalian cells. *Nucleic Acids Res* 19:6595-6602
- Wrenn CK, Katzenellenbogen BS 1993 Structure-function analysis of the hormone binding domain of the human estrogen receptor by region-specific mutagenesis and phenotypic screening in yeast. *J Biol Chem* 268:24089-24098
- Anderson S, Davis DN, Dahlback H, Jornvall H, Russell DW 1988 Cloning, structure, and expression of the mitochondrial cytochrome P-450 sterol 26-hydroxylase, a bile acid biosynthetic enzyme. *J Biol Chem* 264:8222-8229
- Cho H, Katzenellenbogen BS 1993 Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Mol Endocrinol* 7:441-452
- Chen C, Okayama H 1987 High efficiency transformation of mammalian cells by plasmid DNA. *Mol Cell Biol* 7:2745-2752
- Herbommel P, Bourachot B, Yaniv M 1984 Two distinct enhancers with different cell specificities coexist in the regulatory region of polyoma. *Cell* 39:653-662
- Nielsen DA, Chang TC, Shapiro DJ 1989 A highly sensitive mixed-phase assay for chloramphenicol acetyl transferase activity in cultured cells. *Anal Biochem* 179:19-23
- Zhuang Y, Katzenellenbogen BS, Shapiro DJ 1995 Estrogen receptor mutants which do not bind 17 β -estradiol dimerize and bind to the estrogen response element *in vivo*. *Mol Endocrinol* 9:457-466
- Fujimoto N, Katzenellenbogen BS 1994 Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen selectivity and promoter dependence. *Mol Endocrinol* 8:296-304
- Sartorius CA, Tung L, Takimoto GS, Horwitz K 1993 Antagonist-occupied human progesterone receptors bound to DNA are functionally switched to transcriptional agonists by cAMP. *J Biol Chem* 268:9262-9266
- Beck CA, Weigel NL, Moyer ML, Nordeen SK, Edwards DP 1993 The progesterone receptor antagonist RU486 acquires agonist activity upon stimulation of cAMP signaling pathways. *Proc Natl Acad Sci USA* 90:4441-4445
- Rangarajan PN, Umesono K, Evans RM 1992 Modulation of glucocorticoid receptor function by protein kinase A. *Mol Endocrinol* 6:1451-1457
- Nordeen SK, Bona BJ, Moyer ML 1993 Latent agonist activity of the steroid antagonist, RU486, is unmasked in cells treated with activators of protein kinase A. *Mol Endocrinol* 7:731-742
- Cho H, Aronica SM, Katzenellenbogen BS 1994 Regulation of progesterone gene expression in MCF-7 breast cancer cells: a comparison of the effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology* 134:658-664
- Berry M, Metzger D, Chambon P 1990 Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-2817
- Allan GF, Leng X, Tsai SY, Weigel NL, Edwards DP, Tsai M-J, O'Malley BW 1992 Hormone and antihormone induce distinct conformational changes which are central to steroid receptor activation. *J Biol Chem* 267:19513-19520
- Tzukerman MT, Esty A, Santiso-Mere D, Danielian P, Parker MG, Stein RB, Pike WJ, McDonnell DP 1994 Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol Endocrinol* 8:21-30
- Nagaya T, Jameson JL 1993 Distinct dimerization domains provide antagonist pathways for thyroid hormone receptor action. *J Biol Chem* 268:24278-24282
- Rentoumis A, Chatterjee VKK, Madison LD, Datta S, Gallagher GD, DeGroot LJ, Jameson JL 1990 Negative and positive transcriptional regulation by thyroid hormone receptor isoforms. *Mol Endocrinol* 4:1522-1531
- Wang Y, Miksicek RJ 1991 Identification of a dominant negative form of the human estrogen receptor. *Mol Endocrinol* 11:1707-1715
- Fuqua SAW, Fitzgerald SD, Allred DC, Elledge RM, Nawaz Z, McDonnell DP, O'Malley BW, Greene GL, McGuire WL 1992 Inhibition of estrogen receptor action by a naturally occurring variant in human breast tumors. *Cancer Res* 52:483-486

Responses to Pure Antiestrogens (ICI 164384, ICI 182780) in Estrogen-Sensitive and -Resistant Experimental and Clinical Breast Cancer^a

R. I. NICHOLSON,^b J. M. W. GEE,^b D. L. MANNING,^b
A. E. WAKELING,^c M. M. MONTANO,^d AND
B. S. KATZENELLENBOGEN^d

^bTenovus Cancer Research Centre
University of Wales College of Medicine
Tenovus Building
Heath Park

Cardiff CF4 4XX, United Kingdom

^cZeneca Pharmaceuticals
Macclesfield, United Kingdom

^dDepartment of Physiology and Biophysics
University of Illinois
Urbana, Illinois

INTRODUCTION

The last ten years has seen the emergence of a new class of pharmacological agents termed pure antiestrogens (reviewed in Refs. 1, 2). These compounds, which were originally discovered by ICI Pharmaceuticals Division (now Zeneca Pharmaceuticals) in the UK, have the unique property of binding to the estrogen receptor (ER),³ producing a receptor complex which lacks estrogenic activity.^{4,5} They are of use in two important areas of breast cancer research. Firstly, as clinical agents, where it is hoped that their ability to induce total estrogen deprivation will improve the effectiveness of endocrine therapy. Secondly, as pharmacological probes to investigate the cellular and molecular actions of estrogens and tamoxifen. Inherent in each of these areas of research are questions associated with the impact pure antiestrogens may have on the therapy of endocrine-resistant states and whether resistance develops as a consequence of incomplete estrogen withdrawal; with tumor cells more efficiently utilizing either a reduced estrogenic pool or the agonistic activity of an antiestrogen,

^aThe authors gratefully acknowledge the financial support of the Tenovus Organisation (RIN, JMWG, DLM), the Association for International Breast Cancer Research (RIN), the National Institutes of Health, and the Susan G. Komen Foundation (BSK).

or whether the resistant cells have completely circumvented the need for ER-mediated growth and hence sensitivity to pure antiestrogens.²

Since pure antiestrogens are now entering clinical development, the current paper seeks to outline some of their basic cellular and antitumor properties on estrogen-responsive (MCF-7) human breast cancer cells *in vitro*, primarily using the lead compound ICI 164384. This information will be briefly compared with the properties exhibited by pure antiestrogens in endocrine-resistant variants of human breast cancer cells (see refs. in Ref. 2) and phase I and II trials of ICI 182780 in primary⁶ and advanced⁷ breast cancer patients. Where possible examples will be given from immunohistochemical studies, since this technique is most readily applicable to clinical material and ultimately should facilitate an assessment of the degree to which pure antiestrogens are fulfilling their potential as complete antagonists of estrogen action in clinical breast cancer and thereby aid in defining the importance of estrogens in the regulation of breast cancer growth and development.

Experimental Studies with Pure Antiestrogens

Biological Consequences of Exposure of Breast Cancer Cells *In Vitro* to Pure Antiestrogens

Evidence from breast cancer cells grown in culture suggests that pure antiestrogens may be highly efficient in counteracting the stimulatory effects exhibited by estrogens both on cell proliferation and on steroid hormone-regulated gene expression.

One of the most important early observations arising from the functional disablement of ER signalling by pure antiestrogens in estrogen-sensitive human breast cancer cell lines was that, in contrast to the stimulatory activity of estradiol, treated cells became efficiently growth arrested (Fig. 1A).⁸⁻¹⁰ This action is reflected in the growth dynamics of the tumor cells, with several groups showing that while estradiol increases the tumor cell growth fraction and acts to stimulate the passage of cells through the cell cycle, pure antiestrogens promote a highly effective restriction of the cell cycle approximately 5 hours into G1 and hence cause a reduction of the proportion of cells undergoing DNA synthesis.^{8,9} On continuous exposure to pure antiestrogens there is almost a complete loss of those nuclear antigens which mark cell proliferation (Fig. 1B,C)¹⁰ as a large proportion of the cells pass into a noncycling population.⁹ Such cells show a reduced RNA/DNA ratio, characteristic of G0 (Nicholson, Francis and Hoy, unpublished results). It is noteworthy that the growth-inhibitory activity of pure antiestrogens is not solely restricted to cytostatic activity; on continuous exposure there also appears to be a limited cytotoxic component.⁵

The growth-inhibitory activity of pure antiestrogens on human breast cancer cell lines is characteristically preceded by changes in the expression of several estrogen-regulated genes,^{5,10-14} with, for example, the high levels of nuclear immunodetectable PR induced in estradiol-treated MCF-7 cells being rapidly reversed by a 100-fold molar excess of ICI 164384 (Fig. 1D).⁵ Indeed, examination of the percentage of PR-positive cells throughout estradiol and ICI 164384 treatment shows that not only does the pure antiestrogen block estradiol-induced PR levels, but that it also obliterates all PR signalling after 4 days of culture. Such cells are as a consequence no longer responsive to progesterone.

Similarly, the substantial increase in cytoplasmic pS2 immunostaining (a protein of unknown function in the breast) that is induced by estradiol in human breast cancer cells is largely absent following ICI 164384 treatment.⁵ Any residual pS2 staining tends to be present towards the outer cell membrane in small secretory vesicles. Once secreted, however, the cells remain negative, with no evidence of further pS2 production within the endoplasmic reticulum.

Predictably, the decrease in estrogen-regulated proteins often parallels a highly significant fall in their mRNA levels,^{3,11,13,14} with, for example, pS2 mRNA levels being undetectable following 5 days of ICI 164384 treatment.⁵ Indeed, even after reverse transcription PCR (30 cycles), the pS2 mRNA has been shown to be barely detectable in ICI 164384- and ICI 182780-treated cells,⁵ indicating that pure antiestrogens can produce a rapid and complete shutdown of estrogen-regulated gene function. These actions contrast with the effect of both ICI 164384 and ICI 182780 on the estrogen-suppressed gene sequence pMGT-1, the expression of which is very significantly upregulated in their presence.¹⁵

A number of the changes in gene expression may directly contribute to the mechanism of action of the drugs, with ICI 182780 promoting decreases in immunodetectable TGF α , an estrogen-inducible mitogenic growth factor,¹⁶ and the bcl-2 protein, a factor which has been implicated in the protection of cells against programmed cell death.¹⁷ In each instance, while these proteins are readily detectable in a high proportion of cells treated by estradiol, their levels are lowered by estrogen withdrawal and further reduced by the pure antiestrogen.⁵ This is especially evident for the bcl-2 protein, with estradiol-related immunostaining being largely abolished by ICI 182780. Indeed, bcl-2 positivity is a relatively rare event following the administration of pure antiestrogens and, in line with its role in cell survival, its absence is often associated with the presence of pyknotic tumor cell nuclei.⁵

Comparison with Antiestrogens Exhibiting Partial Estrogen-like Activity In Vitro

The inhibitory actions of pure antiestrogens, initially on estrogen-induced transcriptional events and subsequently on cell proliferation and survival, consistently exceed those effects which may be achieved by established antiestrogens with partial estrogen-like activity.

A comparison of the potency and efficacy of tamoxifen, ICI 164384 and ICI 182780 as inhibitors of the growth of MCF-7 cells showed that the pure antiestrogens are up to two orders of magnitude more potent,^{12,21,22} reflecting, in part, their higher

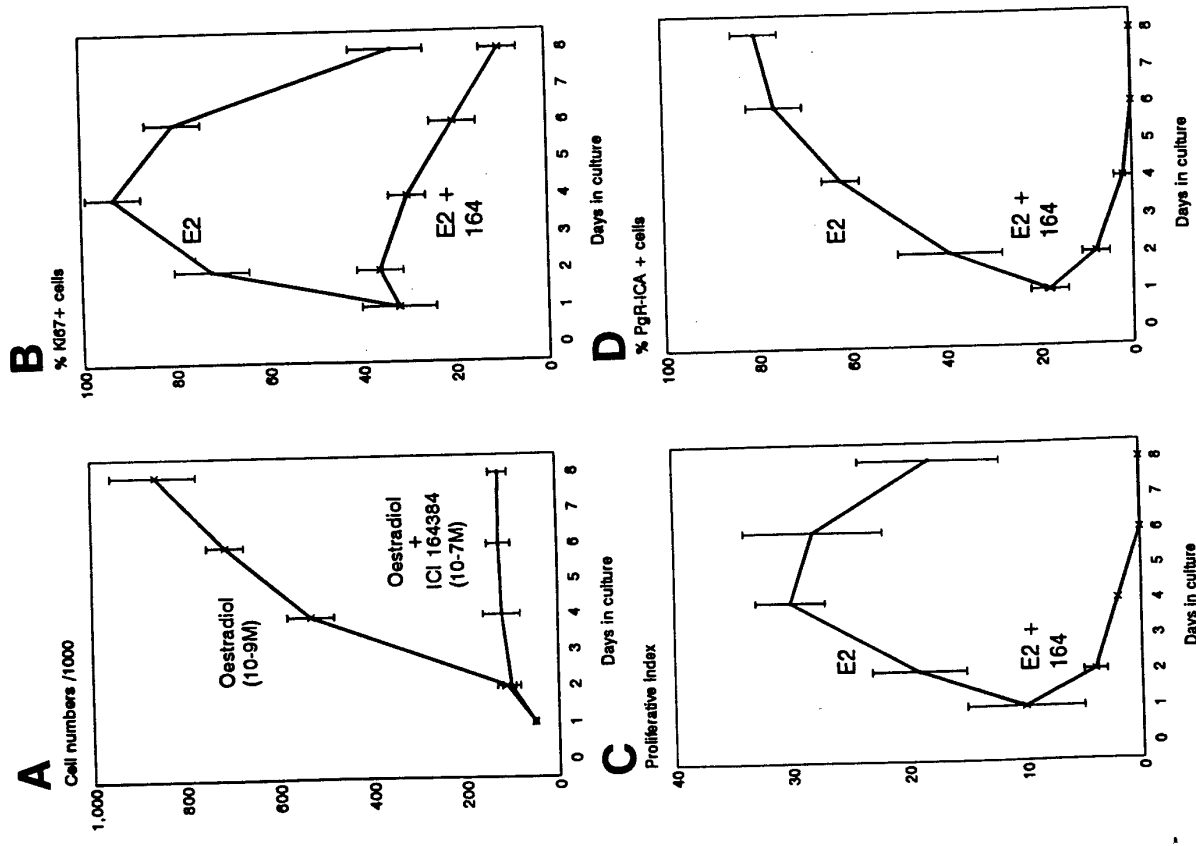


FIGURE 1. Growth and immunohistochemical characterization of MCF-7 cells. The cells were grown in multiwell dishes in white RPMI tissue culture medium with 5% DCC stripped FCS (medium A) containing estradiol \pm ICI 164384. (A) Cell numbers were assessed using a Coulter Counter; (B,C) Ki67 and (D) PR assays were performed according to the methods of Bouzubar *et al.*¹⁸ and Press & Greene,¹⁹ respectively. The Ki67 proliferative index (c) was calculated as the proportion of cells showing intense nucleoplasmic and nucleolar staining patterns.²⁰ The results are shown as the mean \pm SD of six replicates.

produced steroids, through the estrogen receptor, would remain vulnerable to the antagonistic activity of pure antiestrogens.

Thus, in charcoal-stripped serum it is possible that some estradiol is formed from polar precursors that are inefficiently adsorbed from the fetal calf serum, the most obvious candidate being estrone sulphate. Indeed, several groups have shown sulphatase activity in estrogen-sensitive human breast cancer cell lines which can initiate the conversion of estrone sulphate to estradiol.²⁸⁻³⁰ Such conversions can occur at physiological concentrations of estrone sulphate, generating sufficient quantities of estradiol to stimulate growth in estrogen-withdrawn cells (0.1 nM estrone sulphate significantly increases S-phase fraction). Under these conditions, the nuclear steroids are unconjugated estrone and estradiol.²⁹ This sulphatase activity is, however, lowered by the presence of ICI 164384,^{29,31} with basal PR and pS2 immunostaining abrogated,⁵ an effect which appears not to be due to a direct inhibition of the sulphatase enzyme, but rather to be an indirect ER-mediated response.²⁹

Furthermore, interactions with other growth signalling pathways potentially amplify the importance of low estradiol concentrations, with several studies demonstrating that the stimulatory actions of estrogens on tumor cell growth and gene expression can be potentiated by EGF/TGF α , IGF-1 and the FGF family.^{22,32-34} In the case of EGF/TGF α signalling, this may occur through the activation of EGF-receptor tyrosine kinase activity, which, in its turn, influences key intermediates that interact with ER in the nucleus.³⁵ This concept is supported by the recent observations that an EGF-specific tyrosine kinase inhibitor³⁶ can inhibit the expression of PR and pS2 (Fig. 2A,B) in estrogen-withdrawn MCF-7 cells in the presence of maintained amounts of the estrogen receptor and TGF α (Fig. 2C,D), and that similar compounds can inhibit the growth of MCF-7 cells.³⁷ Importantly, EGF³⁵ and dopamine³⁸ have been shown to activate estrogen receptor signalling in the complete absence of its ligand and thus may operate to enhance the constitutive activity of the estrogen receptor in breast cancer cells *in vitro*. The existence of such mechanisms would normally act to protect cells against complete estrogen withdrawal. Importantly, pure antiestrogens have been shown to reduce the above interactions between estrogen receptor and growth factor signalling, generating cells which are desensitized to estrogens, partial antiestrogens, growth factors and dopamine.^{23,35,38} Pure antiestrogens diminish estrogen receptor levels and thus reduce the impact of residual estrogens, and also secondary growth factor signalling. Additionally, ICI 182780 has been shown to reduce the intracellular levels of TGF α (Fig. 4B), thereby reducing its potential autocrine stimulation of breast cancer cells. Elevated TGF α immunostaining in estrogen receptor-positive breast cancer has been recently shown to be associated with a decreased sensitivity of the disease to tamoxifen therapy (39). Its reduction therefore in clinical samples might further improve the sensitivity of breast cancer to estrogen withdrawal.

Biological Consequences of Exposure of Tamoxifen or Estrogen-Resistant Breast Cancer Cell Model Systems to Pure Antiestrogens

Initial clinical responses in breast cancer patients to endocrine-type therapies are all too often overtaken by the development of tumor resistance. This is also the case *in vitro*, where cultured human breast cancer cells gradually gain a comparable

affinity for estrogen receptors.^{3,22} More significantly, flow cytometric analysis of the growth dynamics of the cultured cells showed that, although both classes of agent share the ability to block cell division in the G1 phase of the cell cycle, both ICI 164384 and ICI 182780 were more effective than tamoxifen^{9,22} or hydroxychlorophene⁸ in reducing the proportion of cells which remain able to synthesize DNA after prolonged exposure. These activities, which are specific for estrogen receptor signalling,¹⁰ are reflected in the tumor cell growth fraction^{5,10} with pure antiestrogens abrogating growth responses to tamoxifen.

These differences observed between pure and partial antiestrogens and the control of tumor cell growth have been ascribed to their interactions with other signalling pathways, with the partial agonistic activity of tamoxifen being amplified by the presence of growth factors.⁹ This appears particularly evident for the interaction of tamoxifen and insulin/IGF-1, where a modest growth response to the antiestrogen is considerably increased by the presence of these factors.^{9,23} Such activity is much weaker for ICI 164384, with the compound being more effective than tamoxifen in inhibiting the stimulatory activities of IGF-1 and TGF α .

A further feature of the cellular actions of pure antiestrogens which may relate to their improved antitumor activity has recently been revealed by studying their effects on the expression of estrogen receptors.^{5,10,24} It has been observed that they are associated with a rapid loss of the receptor protein in estrogen receptor-positive cells, producing after relatively short periods of time cellular estrogen-receptor negativity.^{5,10,24,25} This property contrasts with the increases in ER levels that are seen on either estrogen withdrawal or tamoxifen treatment.^{5,10} Recent studies by Fawell *et al.*²⁶ with ICI 164384 have shown that dimerization of the receptor is impaired by the pure antagonist and this may result in the pure antiestrogen receptor complex becoming more fragile and perhaps more sensitive to the normal processes involved in receptor degradation. Certainly, the half-life of the ICI 164384 receptor complex appears substantially shorter²⁵ than the half-lives of the estrogen receptor and tamoxifen receptor complexes.^{26,27}

Comparison with Estrogen Withdrawal In Vitro

Encouragingly and somewhat surprisingly, the effects of pure antiestrogens on the growth of estrogen receptor-positive breast cancer cells substantially surpass the effects of estrogen withdrawal.^{10,21} This property has been demonstrated by several groups, with, for example, ICI 164384 severely impairing the growth of MCF-7 cells in phenol red-free medium where the 5% fetal calf serum has been extensively stripped of its endogenous estrogens by charcoal absorption, a procedure which reduces the level of endogenous estradiol to below 10^{-12} M.^{5,10} Once again changes in cell numbers correspond to their recorded growth dynamics, with pure antiestrogens decreasing tumor cell S-phase fraction and increasing the proportion of cells in G0/G1 relative to estrogen withdrawn cells.²²

The local production of hormones by breast cancer cells may play an important role in promoting some cell growth and gene expression in estrogen-withdrawn cells. The cells would potentially be highly sensitive to these, since estrogen receptor levels are elevated following oestrogen withdrawal.^{5,10} However, the actions of locally

endocrine resistance following their prolonged exposure to either estrogen withdrawal or antiestrogens. This phenomenon tends to be associated with an increase in cell growth rate, while retaining the capacity to express estrogen receptors and thus potentially to demonstrate estrogen receptor-mediated responses. Studies with T47D and MCF-7 estrogen-responsive human breast cancer cells suggest that the mechanism underlying the development of tamoxifen resistance^{2,40-43} is the outgrowth of subpopulations of cells whose growth is stimulated by tamoxifen,^{40,41} rather than the selection of cells which are unaffected by tamoxifen.⁴⁴

Fortunately, it is likely that such tamoxifen-resistant cells retain a sensitivity to the growth-inhibitory actions of pure antiestrogens *in vitro*,^{12,13,45} and thus it is feasible that these new compounds may be of clinical value in patients who relapse on tamoxifen therapy. In contrast, cross-resistance to several structurally diverse partial antiestrogens has been demonstrated in the antiestrogen-resistant LY2 cell line.^{46,47} A similar resistance mechanism has been identified *in vivo* in animal model systems,^{40,47} with tamoxifen-resistant cells again exhibiting growth inhibition by pure antiestrogens.^{48,49} This is associated with a parallel decrease in the intracellular levels of estrogen receptors and an associated fall in estrogen-regulated gene products.⁴⁹

Furthermore, it is likely that estrogen-resistant breast cancer cells also retain a sensitivity to the inhibitory actions of pure antiestrogens.^{52,53} FIGURE 3A illustrates this phenomenon for a MCF-7 variant cell line, K3.⁵³⁻⁵⁵ This cell line was derived from estrogen-responsive MCF-7 cells by estrogen withdrawal and no longer shows a growth stimulation by estradiol (cmf. response of wild-type cells to oestradiol; FIG. 3B). Despite this, it is growth inhibited by antiestrogens, with ICI 182780 (10^{-7} M) being more effective than 4-hydroxytamoxifen (10^{-7} M). Indeed, only two doublings of the initial cell number occurred throughout ICI 182780 treatment and contrasts with the 7 to 8 doublings recorded under estrogen-treated or -withdrawn conditions. Over several experiments the estimated tumor cell doubling time for ICI 182780-treated K3 cells is in excess of 150 hours. This compares favorably with the value that may be estimated following the treatment of wild-type MCF-7 cells with pure antiestrogens (FIG. 3B).

The actions of ICI 164384 and ICI 182780 cells appear specific for ER signalling and may be reversed by the presence of estradiol.⁵³ Indeed, K3 cells appear more sensitive to the growth-promoting effects of the steroid than do wild-type cells, with 10^{-9} M and 10^{-8} M estradiol reversing the effects of 10^{-7} M ICI 182780, respectively. As a possible contributor to the increased sensitivity of K3 cells to estradiol, the cells also show a higher basal expression of TGF α (FIG. 4A),⁵³ which, in contrast to wild-type cells, is not substantially increased by the presence of estradiol.⁵³ The intracellular concentration of TGF α is, however, lowered by the presence of ICI 182780, but not 4-OHT. Importantly, the reduction in TGF α levels in pure antiestrogen-treated cells accompanies a substantial fall in their ER content (FIG. 4C,D).^{53,55} This action would minimize the opportunity for cross-talk between ER and TGF α signalling pathways. Interestingly, K3 cells also show an elevated basal expression of pS2 (FIG. 3C,D),⁵² a protein whose gene promoter contains a response element for both estrogens and TGF α .⁵⁶ Once again, the expression of this protein is efficiently reduced by the presence of the pure antiestrogen (FIG. 3C,D).⁵³

Although less is known about the emergence of breast cancer cells resistant to pure antiestrogens, breast cancer cell xenograft studies have indicated longer remission

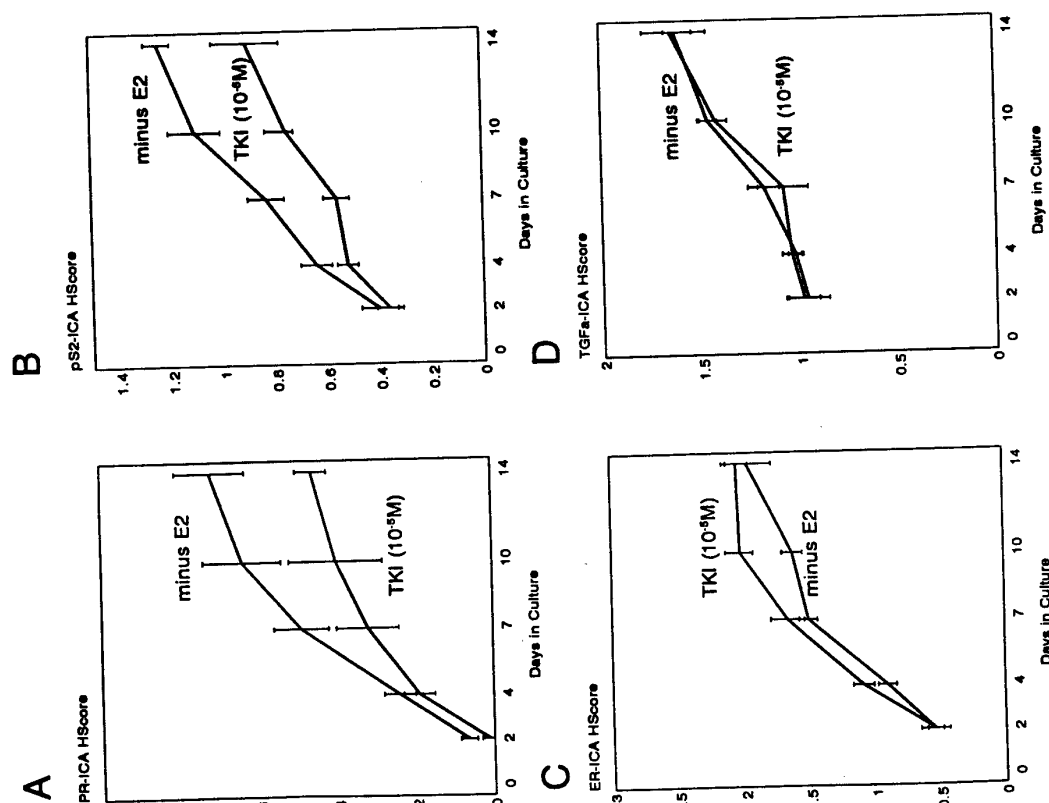


FIGURE 2. Effect of 4-(3-methylamino)quinazoline (ZM 163613) on MCF-7 cells. The cells were grown on TESPA-coated coverslips in medium A (minus E2) with and without the tyrosine kinase inhibitor (TKI). Assays for PR (A), pS2 (B), ER (C) and TGF α (D) were performed according to the methods of Press & Greene¹⁹ (A), Charpin *et al.*⁵⁰ (B), Walker *et al.*⁵¹ (C) and Nicholson *et al.*³⁹ (D). The results are the mean values \pm SD of 5 replicates from a minimum of 2 coverslips.

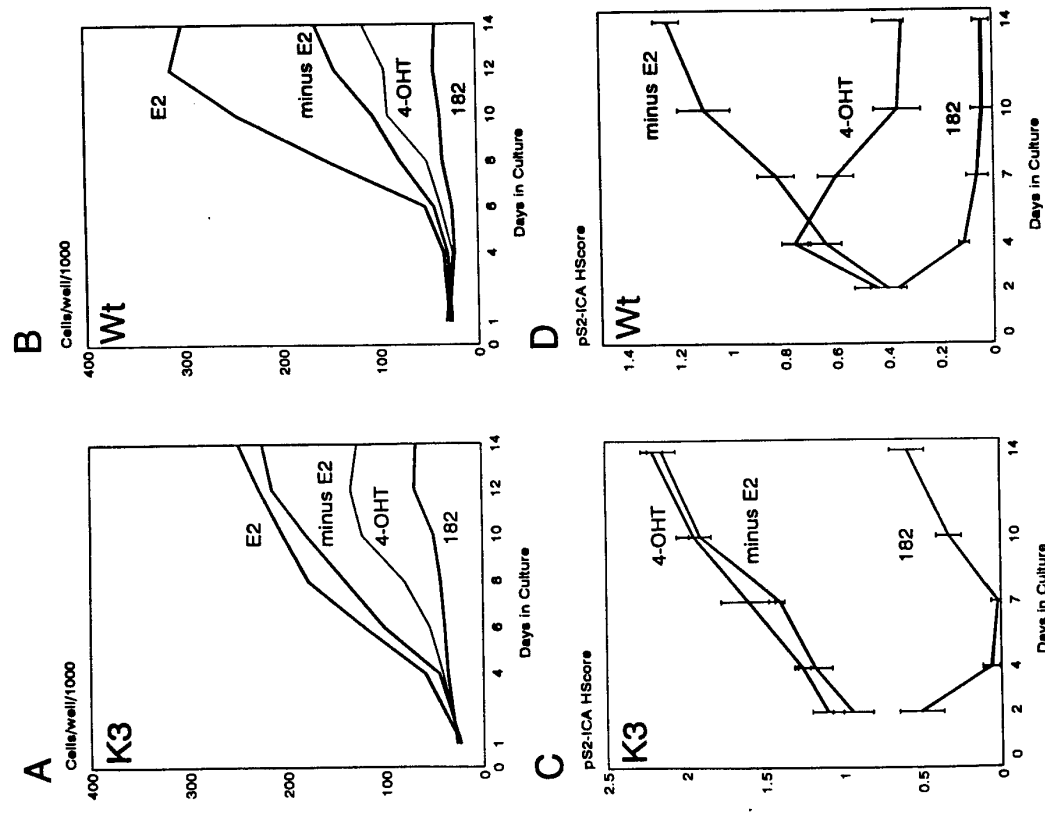


FIGURE 3. Growth and immunohistochemical characterisation of K3 and wild type (Wt) MCF-7 cells. K3 (A) and Wt (B) cells were grown in multiwell dishes in medium A containing no additives (minus E2), estradiol (10^{-9} M; E2), 4 hydroxytamoxifen (10^{-7} M; 4-OHT) and ICI 182780 (10^{-7} M; 182). Cell numbers were assessed using a Coulter Counter. K3 (C) and Wt (D) cells were grown on TESPA-coated coverslips in medium A containing no additives, 4-OHT (10^{-7} M) and 182 (10^{-7} M) and assayed for pS2.⁵⁰ The results are the mean value \pm SD of 5 replicates from a minimum of 2 coverslips.

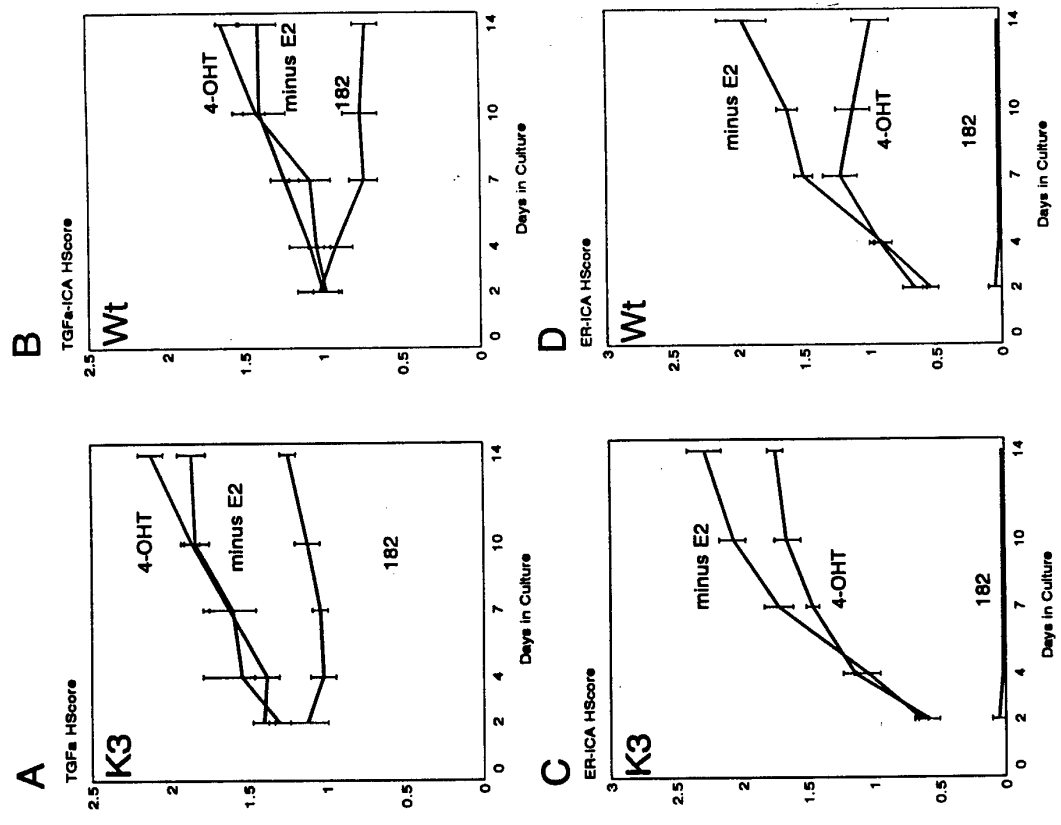


FIGURE 4. Immunohistochemical characterization of K3 and Wt MCF-7 cells. K3 (A,C) and Wt (B,D) cells were grown on TESPA-coated coverslips under conditions as described in Figure 3C,D and assayed for TGF α (A,B) and ER (C,D). The results are the mean value \pm SD of 5 replicates from a minimum of 2 coverslips.

times with pure antiestrogens than with either tamoxifen or estrogen withdrawal.⁴⁹ Moreover, where ICI 182780-resistant tumors have been transplanted into castrated mice, their growth has been shown to be slightly impeded by tamoxifen alone or in combination with ICI 182780. Such data imply that true cross-resistance to these agents has not fully developed in this animal model, despite the fact that pure antiestrogens abrogate ER levels.⁴⁹ Interestingly, this may not be the case *in vitro*,⁴⁵ since cultured MCF-7/LCC9 ICI 182780 resistant cells are also tamoxifen resistant.⁴⁵

Properties of Pure Antiestrogens in Clinical Breast Cancer

Antitumor Activity

Although clinical trials with pure antiestrogens are in their infancy and consequently little is known about their clinical properties, in late 1991 a Phase I study of ICI 182780 (6 or 18 mg/day in a short acting propylene glycol-based formulation) was initiated.^{5,6} The purpose of this study was to assess the safety and pharmacokinetic properties of the drug and to begin to investigate its biological effects on tumor tissue. The latter was achieved by measuring a number of immunohistochemical end points on pretreatment needle core biopsies from newly diagnosed primary breast cancer patients, comparing results with identical measurements performed on the posttreatment specimen removed at the time of primary surgery seven days later. Analysis of these data has shown that, as observed *in vitro* and within animal model systems, the pure antiestrogen is capable of reducing both tumor cell proliferation and estrogen-regulated gene expression.

In the Phase I study, ICI 182780 produced a highly significant decrease in ER expression in patients with tumors initially classified as ER positive, mirroring those effects observed in experimental systems. Comparison of these data with similar measurements derived from short-term tamoxifen-treated patients showed that although tamoxifen also produced a suppression of ER levels, the effects were not as great as those induced by the highest dose level of ICI 182780 (18 mg/day). Furthermore, examination of PR levels in these samples revealed a divergence of response to the antiestrogens, with the pure antiestrogen inhibiting PR expression while tamoxifen promoted some increase in its tissue concentration. Thus, in initially ER-positive and PR-positive tumors, ICI 182780 caused a fall in PR levels in the majority of patients. Indeed, in several patients the tumors became negative for both of these steroid receptors after the seven-day treatment period. The estrogen-regulated protein pS2 showed similar responses to PR, although the inhibitory effects of ICI 182780 appeared blunted. No short-term effects of ICI 182780 or tamoxifen were recorded on the mammotrophic growth factor TGF α or the bcl-2 protein. However, ICI 182780 (18 mg/day) produced a significant reduction in the expression of the tumor cell proliferation marker Ki-67, with staining values decreasing in the majority of ER-positive tumors examined.

A more recent second study directly attempted to address the issues of whether pure antiestrogens can promote clinical tumor remissions of worthwhile duration and also if these drugs remain effective in tamoxifen-resistant patients.⁷ Initial results using an oil-based monthly depot again appear promising. Thus, approximately two-

thirds of women who had either received adjuvant tamoxifen (minimum of two years) and subsequently recurred, or who had relapsed following an initial favorable response to the antiestrogenic drug, gained further benefit from ICI 182780 treatment (7/19 partial responses, 6/19 no change at 6 months). Within responding patients, 10/13 women were still in remission at 9 months. However, although these results are better than would have been expected following tamoxifen withdrawal^{57,58} or second line endocrine therapy,⁵⁹ the study numbers were small and no direct randomized comparisons were made with other endocrine measures. It is noteworthy, however, that in clinical studies where tamoxifen-resistant tumors have been treated with another triphenylethylene antiestrogen, toremifene, cross-resistance was demonstrated.⁵⁶ This result parallels experimental studies on antiestrogen-resistant human breast cancer cells.

Side Effects

To date, no serious drug-related side effects have been reported in either of the above clinical studies. In particular, there has been no evidence of altered coagulation or thrombogenicity after treatment with ICI 182780. On long-term therapy a rise in serum gonadotrophin levels has been recorded, suggesting that ICI 182780 has an antiestrogenic effect on the pituitary gland. There were no significant changes in serum levels of sex hormone binding globulin, implying no estrogenic effect of the drug on the liver.

CONCLUSIONS

It is highly encouraging that the majority of experimental studies to date have shown the recently developed pure antiestrogens to be effective antitumor agents, certainly with regard to inhibition of tumor cell growth and proliferation, and estrogen-regulated gene expression. Indeed, in many instances their effects substantially surpass those observed following estrogen withdrawal or tamoxifen therapy. Treatment of breast cancer cells with pure antiestrogens appears to promote an efficient growth arrest *in vitro* and *in vivo* by their induction of a state of strict estrogen deprivation. Such estrogen withdrawal is likely to be induced primarily by the compounds antagonizing the cellular actions of estrogens and, possibly, reducing other growth signalling activities through the estrogen receptor. Importantly, these mechanisms appear relevant to the treatment of endocrine-resistant states.

In clinical breast cancer it is too early to judge the final value of these compounds. They have, however, passed the first major hurdle by successfully promoting tumor remissions in previously treated patients generating minimal adverse side-effects. The compounds seem capable of reducing cell proliferation and expression of both the estrogen receptor and several estrogen-regulated genes. However, their ultimate success or failure will depend on many factors, most notably the importance of small amounts of estrogen to the tumor cell. If we have not, as yet, passed the threshold of response to estrogen withdrawal, a potential exists for pure antiestrogens to improve the outcome of endocrine therapy^{61,62} in such important areas as the rate and duration of remission and the prevention and treatment of resistant states.² Over the next few

years, analysis of the clinical actions of pure antiestrogens will establish many unknowns in breast cancer. Let us hope that one of them is that the inhibition of all ER-mediated signalling is a worthwhile goal.²

REFERENCES

1. WAKELING, A. E. 1993. The future of pure antiestrogens in clinical breast cancer. *Br. Cancer Res. Treat.* 25: 1-9.
2. NICHOLSON, R. I., A. B. FRANCIS, D. L. MANNING & J. M. W. GEE. 1994. Pure antiestrogens (ICI 164384 and ICI 182780) and breast cancer: Is the attainment of complete oestrogen withdrawal worthwhile? *Endocrine-Related Cancer* 1(4): 5-17.
3. WILSON, A. P. M., P. J. WEATHERILL, R. I. NICHOLSON, P. DAVIES & A. E. WAKELING. 1990. A comparative study of the interaction of oestradiol and the steroidal pure antiestrogen, ICI 164384 with molybdate stabilized oestrogen receptor. *J. Steroid Biochem.* 35: 421-428.
4. WAKELING, A. E. & J. BOWLER. 1987. Steroidal pure antiestrogens. *J. Endocrinol.* 112: R7-R10.
5. NICHOLSON, R. I., J. M. W. GEE, C. L. EATON, D. L. MANNING, R. E. MANSEL, A. K. SHARMA, A. DOUGLAS-JONES, M. PRICE-THOMAS, A. HOWELL, D. J. DEFRIEND, N. J. BUNDRED, E. ANDERSON, J. F. R. ROBERTSON, R. W. BLAMEY, M. DOWSETT, P. WALTON & A. E. WAKELING. 1994. Pure antiestrogens in breast cancer: experimental and clinical observations. In *Sex Hormones and Antihormones in Endocrine Dependent Pathology: Basic and Clinical Aspects*. Excerpta Medica International Congress Series. M. Motta & M. Serio, Eds. 347-360. Elsevier Science B. V. Amsterdam.
6. DEFRIEND, D. L., A. HOWELL, R. I. NICHOLSON, E. ANDERSON, M. DOWSETT, R. E. MANSEL, R. W. BLAMEY, N. J. BUNDRED, J. F. R. ROBERTSON, C. SAUNDERS, M. BAUM, P. WALTON, F. SUTCLIFFE & A. E. WAKELING. 1994. Investigation of a new pure antiestrogen (ICI 182780) in women with primary breast cancer. *Cancer Res.* 54: 1-7.
7. HOWELL, A., D. L. DEFRIEND, R. W. BLAMEY, J. F. ROBERTSON & P. WALTON. Response to a pure antiestrogen (ZM 182780) in tamoxifen resistant breast cancer. *Lancet*. In press.
8. MUSGROVE, E. A., A. E. WAKELING & R. L. SUTHERLAND. 1989. Points of action of estrogen antagonists and a calmodulin antagonist within the MCF-7 human breast cancer cell cycle. *Cancer Res.* 49: 2398-2404.
9. WAKELING, A. E., E. NEWBOULT & S. W. PETERS. 1989. Effects of antiestrogens on the proliferation of MCF-7 human breast cancer cells. *J. Mol. Endocrinol.* 2: 225-234.
10. NICHOLSON, R. I., K. J. WALKER, N. BOUZUBAR, R. WILLS, J. M. W. GEE, N. K. RUSHMERE & P. DAVIES. 1990. Estrogen deprivation in breast cancer: clinical, experimental and biological aspects. *Ann. N. Y. Acad. Sci.* 595: 316-327.
11. WISEMAN, L. R., A. E. WAKELING, F. E. MAY & B. R. WESTLEY. 1989. Effects of the antiestrogen, ICI 164384, on oestrogen induced mRNAs in MCF-7 cells. *J. Steroid Biochem.* 31: 1-6.
12. LYKESFELDT, A. E. & E. K. SORESENSEN. 1992. Effect of estrogen and antiestrogens on cell proliferation and synthesis of secreted proteins in the human breast cancer cell line MCF-7 and a tamoxifen resistant variant subline, AL-1. *Acta Oncol.* 31: 131-138.
13. LYKESFELDT, A. E., M. W. MADSEN & P. BRIAND. 1994. Altered expression of estrogen-regulated genes in a tamoxifen-resistant and ICI 164384 and ICI 182780 sensitive human breast cancer cell line, MCF-7/TAM^R-1. *Cancer Res.* 54: 1587-1595.
14. MANNING, D. L., R. A. MCCLELLAND, S. BRYANT, J. KNOWLDEN, J. M. W. GEE, A. B. FRANCIS, J. F. R. ROBERTSON, R. W. BLAMEY, R. L. SUTHERLAND, C. J. ORMANDY, R. E. MANSEL & R. I. NICHOLSON. Oestrogen regulated genes in breast cancer: role in prognosis and endocrine sensitivity. *Acta Oncol.* In press.
15. MANNING, D. L. & R. I. NICHOLSON. 1993. Isolation of pMGT1: a gene that is repressed by oestrogen and increased by antiestrogens and antiestrogens. *Eur. J. Cancer* 29A: 759-762.
16. BATES, S. E., N. E. DAVIDSON, E. V. VALVERIUS, C. E. FRETHER, R. B. DICKSON, J. P. TAM, J. E. KUDLOW, M. E. LPPMAN & D. S. SOLOMON. 1988. Expression of transforming growth factor alpha and its messenger ribonucleic acid in human breast cancer: its regulation by oestradiol and its possible functional significance. *Mol. Endocrinol.* 2: 543-555.
17. GEE, J. M. W., J. F. R. ROBERTSON, H. B. HOYLE, S. R. KYME, R. A. MCCLELLAND, I. O. ELLIS, R. W. BLAMEY & R. I. NICHOLSON. 1994. Immunocytochemical localisation of BCL-2 protein in human breast cancer and its relationship to a series of prognostic markers and response to endocrine therapy. *Int. J. Cancer.* 59: 619-628.
18. BOUZUBAR, N., K. J. WALKER, K. GRIFFITHS, I. O. ELLIS, C. W. ELSTON, J. ROBERTSON, R. W. BLAMEY & R. I. NICHOLSON. 1989. Ki67 immunostaining in primary breast cancer: pathological and clinical associations. *Br. J. Cancer* 59: 943-947.
19. PRESS, M. F. & G. L. GREENE. 1988. Immunocytochemical localisation of oestrogen and progesterone receptors. In *Advances in Immunohistochemistry*. R. A. DeLellis, Ed. 341-361. Raven Press. New York.
20. BOUZUBAR, N. F. H. 1991. Studies on hormone and antihormone action in early breast cancer using immunocytochemical techniques. PhD Thesis. University of Wales College of Medicine. Cardiff.
21. WAKELING, A. E. & J. BOWLER. 1988. Novel antiestrogens without partial agonistic activity. *J. Steroid Biochem.* 31: 645-653.
22. WAKELING, A. E., M. DUKES & J. BOWLER. 1991. A potent specific pure antiestrogen with clinical potential. *Cancer Res.* 51: 3867-3873.
23. NICHOLSON, R. I. 1992. Pure antiestrogens and breast cancer. In *Current Directions in Cancer Chemotherapy*. IBC Technical Services. London.
24. GIBSON, M. K., L. A. NEMMERS, L. A. BECKMAN, W. C. BECKMAN, V. L. DAVIS, S. W. CURTIS & K. S. KORACH. 1991. The mechanism of ICI 164384 antiestrogenicity involves rapid loss of estrogen receptor in uterine tissue. *Endocrinology* 129: 2000-2010.
25. DAUVOIS, S., P. S. DANIELIAN, R. WHITE & M. G. PARKER. 1992. Antiestrogen ICI 164384 reduces cellular estrogen receptor content by increasing its turnover. *Proc. Natl. Acad. Sci. USA* 89: 4037-4041.
26. FAWELL, S. E., R. WHITE, S. HOARE, M. SYDENHAM, M. PAGE & M. G. PARKER. 1990. Inhibition of estrogen receptor-DNA binding by the pure antiestrogen ICI 164384 appears to be mediated by impaired receptor dimerisation. *Proc. Natl. Acad. Sci. USA* 87: 6883-6887.
27. ECKERT, R. L., A. MULLICK, E. A. RORKE & B. S. KATZENELLENBOGEN. 1984. Estrogen receptor synthesis and turnover in MCF-7 breast cancer cells measured by a density shift technique. *Endocrinology* 114: 629-637.
28. PASQUALINI, J. R., B. SCHATZ, C. VARIN & B. L. NGUYEN. 1992. Recent data on estrogen sulfatases and sulphotransferases activities in human breast cancer. *J. Steroid Biochem. Mol. Biol.* 41: 323-329.
29. SANTNER, S. J., B. OHLSSON-WILHELM & R. J. SANTEN. 1993. Estrone sulfate promotes human breast cancer cell replication and nuclear uptake of estradiol in MCF-7 cell cultures. *Int. J. Cancer* 54: 119-124.
30. HOWARTH, N. M., A. PUROHIT, M. J. REED & B. V. L. POTTER. 1994. Estrone sulfamates: potential inhibitors of estrone sulfatase with therapeutic potential. *J. Med. Chem.* 37: 219-221.
31. CHETRITE, G., C. VARIN, L. DELALONDE & J. R. PASQUALINI. 1993. Effect of promegestone, tamoxifen, 4-hydroxytamoxifen and ICI 164384 on the oestrogen sulphatase activity of human breast cancer cells. *Anticancer Res.* 13: 931-934.
32. STEWART, A. J., M. D. JOHNSON, F. E. MAY & B. R. WESTLEY. 1990. Role of insulin-like growth factors and type I insulin-like growth factor receptor in the estrogen-

- stimulated proliferation of human breast cancer cells. *J. Biol. Chem.* **265**: 21172-21178.
33. STEWART, A. J., B. R. WESTLEY, & F. E. B. MAY. 1992. Modulation of proliferative response of breast cancer cells to growth factors by oestrogen. *Br. J. Cancer* **66**: 640-648.
34. WESTLEY, B. R. & F. E. B. MAY. 1991. IGFs and control of cell proliferation in breast and other cancers. *Rev. Endocr. Relat. Cancer* **39**: 29-34.
35. IGNAR-TROWBRIDGE, D. M., K. G. NELSON, M. C. BIDWELL, S. W. CURTIS, T. F. WASHBURN, J. A. McLACHLAN & K. S. KORACH. 1992. Coupling of dual signalling pathways: epidermal growth factor action involves the estrogen receptor. *Proc. Natl. Acad. Sci. USA* **89**: 4658-4662.
36. WAKELING, A. E., 1994. Inhibition of EGF-receptor tyrosine kinase activity by 4-aminoliquorizolines. *Br. J. Cancer* **69** (Suppl. XXI): 18. Abstract 6.6.
37. REDDY, K. B., G. L. MANGOLD, A. K. TANDON, T. YONEDA, G. R. MUNDY, A. ZILBERSTEIN & C. K. OSBORNE. 1992. Inhibition of breast cancer cell growth *in vitro* by a tyrosine kinase inhibitor. *Cancer Res.* **52**: 3636-3641.
38. POWER, R. F., S. K. MANI, J. CODINA, O. M. CONNEELLY & B. W. O'MALLEY. 1991. Dopaminergic and ligand-independent activation of steroid hormone receptors. *Science* **254**: 1636-1639.
39. NICHOLSON, R. I., R. A. McCLELLAND, J. M. W. GEE, D. L. MANNING, P. CANNON, J. F. R. ROBERTSON, I. O. ELLIS & R. W. BLAMEY. 1994. Transforming growth factor α and endocrine sensitivity in breast cancer. *Cancer Res.* **54**: 1684-1689.
40. OSBORNE, C. K., E. B. CORONADO & J. P. ROBINSON. 1987. Human breast cancer in the athymic mouse: cytostatic effects of long-term antiestrogen therapy. *Eur. J. Cancer Clin. Oncol.* **23**: 1189-1196.
41. OSBORNE, C. K., M. JARMAN, R. McCAGUE, E. B. CORONADO, S. G. HILSENBECK & A. E. WAKELING. 1994. The importance of tamoxifen metabolism in tamoxifen-stimulated growth. *Cancer Chemother. Pharmacol.* **34**: 89-95.
42. GOTTARDIS, M. M. & V. C. JORDAN. 1988. Development of tamoxifen-stimulated growth of MCF-7 tumours in athymic mice after long-term antioestrogen administration. *Cancer Res.* **48**: 5183-5187.
43. CLARKE R., N. BRUNNER, B. S. KATZENELLENBOGEN, E. W. THOMPSON, M. J. NORMAN, C. KORPI, S. PAIK, M. E. LIPPMAN & R. B. DICKSON. 1989. Progression of breast cancer cells from hormone dependent to hormone independent growth both *in vitro* and *in vivo*. *Proc. Natl. Acad. Sci. USA* **86**: 3649-3653.
44. GRAHAM, M. L., J. A. SMITH, P. B. JEWETT & K. B. HORWITZ. 1992. Heterogeneity of progesterone receptor content and remodelling by tamoxifen characterise subpopulations of cultured human breast cancer cells: analysis by quantitative dual parameter flow cytometry. *Cancer Res.* **52**: 593-602.
45. BRUNNER, N., B. BOYSEN, J. F. KILGAARD, T. L. FRANSSEN, S. JURUS & R. CLARKE. 1993a. Resistance to 4OH-tamoxifen does not confer resistance to the steroidal antioestrogen ICI 182780, while acquired resistance to ICI 182780 results in cross-resistance to 4OH-tamoxifen. *Breast Cancer Res. Treat.* **27**: 135. Abstract 19.
46. BRONZERT, D. A., G. L. GREENE & M. E. LIPPMAN. 1985. Selection and characterisation of breast cancer cell line resistant to the antioestrogen LY 117018. *Endocrinology* **117**: 1409-1417.
47. CLARKE, R., E. W. THOMPSON, F. LEONESSA, J. LIPPMAN, M. MCGARVEY, T. L. FRANSSEN & N. BRUNNER. 1993. Hormone resistance, invasiveness, and metastatic potential in breast cancer. *Breast Cancer Res. Treat.* **24**: 227-239.
48. GOTTARDIS, M. M., S.-Y. JIANG, M.-H. JENG & V. C. JORDAN. 1989. Inhibition of tamoxifen-stimulated growth of an MCF-7 tumour variant in athymic mice by novel steroidal antioestrogens. *Cancer Res.* **49**: 4090-4093.
- NICHOLSON *et al.*: PURE ANTIOESTROGENS AND BREAST CANCER 163
49. OSBORNE, C. K., E. B. CORONADO-HENSOHN, B. L. McCUE, A. E. WAKELING & R. I. NICHOLSON. Pure steroidal antioestrogens are superior to tamoxifen in a preclinical model of human breast cancer. Submitted.
50. CHARPIN, C., P. BONNIER, A. KHOUZAMI, H. VAHERET, L. ANDRAC, M. N. LAVAUT, C. ALLASIA & L. PIANA. 1992. Inflammatory breast carcinoma: an immunohistochemical study using anti-pHER-2neu, pS2, cathepsin, ER and PR. *Anticancer Res.* **52**: 591-597.
51. WALKER, K. J., N. BOUZUBAR, J. ROBERTSON, I. O. ELLIS, C. W. ELSTON, R. W. BLAMEY, D. W. WILSON, K. GRIFFITHS & R. I. NICHOLSON. 1988. Immunocytochemical localisation of estrogen receptor in human breast cancer. *Cancer Res.* **48**: 6517-6522.
52. REECE, J. C. & B. S. KATZENELLENBOGEN. 1992. Examination of the DNA-binding ability of ER in whole cells: implications for hormone independent transactivation and the actions of antioestrogens. *Mol. Cell. Biol.* **12**: 4531-4538.
53. NICHOLSON, R. I., A. B. FRANCIS, J. M. W. GEE, D. L. MANNING, A. E. WAKELING & B. S. KATZENELLENBOGEN. Observations arising from the use of pure antioestrogens on oestrogen-Responsive (MCF-7) and oestrogen growth-independent (K3) human breast cancer cells. *Rev. Endocr. Relat. Cancer*. In press.
54. KATZENELLENBOGEN, B. S., K. L. KENDRA, M. J. NORMAN & Y. BERTHOIS. 1987. Proliferation, hormonal responsiveness and estrogen receptor content of MCF-7 human breast cancer cells grown in the short-term and long-term absence of estrogens. *Cancer Res.* **47**: 4355-4360.
55. CHO, H., P. A. NG & B. S. KATZENELLENBOGEN. 1991. Differential regulation of gene expression by estrogen in estrogen growth-independent and -dependent MCF-7 human breast cancer cell sublines. *Mol. Endocrinol.* **5**: 1323-1330.
56. NUNEZ, A., M. BARRY, J. L. IMLER & P. CHAMBER. 1989. The 5'-flanking region of the pS2 gene contains a complex enhancer region responsive to oestrogens, epidermal growth factor, a tumour promoter (TPA), the c-Ha-ras oncoprotein and the c-jun protein. *EMBO J.* **8**: 823-829.
57. HOWELL, A., D. J. DODWELL, I. LAIDLAW, H. ANDERSON & E. ANDERSON. 1990. Tamoxifen as an agonist for metastatic breast cancer. *In Endocrine Therapy of Breast Cancer IV*. A. Goldhirsch, Ed. 49-58. Springer-Verlag. Berlin.
58. HOWELL, A., D. J. DODWELL, H. ANDERSON & J. REDFORD. 1992. Response after withdrawal of tamoxifen and progesterins in advanced breast cancer. *Ann. Oncol.* **3**: 611-617.
59. WILSON, A. J. 1983. Response in breast cancer to a second hormonal therapy. *Rev. Endocr. Relat. Cancer* **14**: 5-11.
60. VOGEL, C. L., I. SHERMAN, J. SCHOENFELDER, P. A. GAMS & M. R. GREEN. 1993. Multicenter Phase II efficacy trial of toremifene in tamoxifen-refractory patients with advanced breast cancer. *J. Clin. Oncol.* **11**: 345-350.
61. NICHOLSON, R. I. 1993. Recent advances in the antihormonal therapy of breast cancer. *Curr. Opin. Invest. Drugs* **2**: 1259-1268.
62. NICHOLSON, R. I., D. L. MANNING & J. M. W. GEE. 1993. New antihormonal approaches to breast cancer therapy. *Drugs Today* **29**: 363-372.

Estrogen Receptors: Bioactivities and Interactions with Cell Signaling Pathways¹

Benita S. Katzenellenbogen²

*Departments of Molecular and Integrative Physiology, Cell and Structural Biology
University of Illinois, Urbana, Illinois 61801–3704*

ABSTRACT

Estrogens regulate the growth, differentiation, and functioning of diverse target tissues, both within and outside of the reproductive system. Most of the actions of estrogens appear to be exerted via the estrogen receptor (ER) of target cells, an intracellular receptor that is a member of a large superfamily of proteins that function as ligand-activated transcription factors, regulating the synthesis of specific RNAs and proteins. To understand how the ER discriminates between estrogen ligands, which activate the ER, and antiestrogen ligands, which fail to effectively activate the ER, we have generated and analyzed human estrogen receptors with mutations in the ER hormone binding domain. These studies provide evidence for the promoter-specific and cell-specific actions of the estrogen-occupied and antiestrogen-occupied ER, highlight a regional dissociation of the hormone binding and transcription activation functions in domain E of the receptor, and indicate that some of the contact sites of estrogens and antiestrogens in the ER are likely different. In addition, multiple interactions among different cellular signaling pathways are involved in the regulation of gene expression and cell proliferation by the ER. In several cell types, protein kinase activators and some growth factors enhance the transcriptional activity of the ER. Cyclic AMP also alters the agonist/antagonist balance of some antiestrogens. Estrogens and, to a lesser extent, antiestrogens, as well as protein kinase activators and growth factors increase phosphorylation of the ER and possibly other proteins involved in the ER-specific response pathway, suggesting that changes in cellular phosphorylation state will be important in determining the biological activity of the ER and the effectiveness of antiestrogens as estrogen antagonists. The ER also has important interrelationships with the progesterone receptor (PR) system in modulation of biological responses. Liganded PR-A and PR-B can each suppress estradiol-stimulated ER activity, with the magnitude of repression dependent on the PR isoform, progestin ligand, promoter, and cell type. These findings underscore the mounting evidence for the importance of interactions between members of the steroid hormone receptor family.

OVERVIEW: THE DIVERSITY OF ESTROGEN TARGET TISSUES

The actions of estrogenic hormones are mediated through the estrogen receptor (ER), a member of a large superfamily of nuclear receptors that function as ligand-activated transcription factors. These receptor proteins share a common structural and functional organization, with distinct domains that are responsible for ligand-binding, DNA-binding, and transcription activation [1–5].

Two highly conserved regions are observed in these receptors, one in approximately the middle of the protein (known as domain C), which is involved in interaction with DNA, and one in the carboxy-terminal region (known as domain E/F) that binds hormones and is structurally and functionally complex. Upon binding estrogen, the ER binds to estrogen-response-element DNA, often located in the 5' flanking region of estrogen responsive genes. These DNA sequences function as enhancers, conferring estrogen inducibility on the genes. The estrogen-occupied receptor is then thought to interact with transcription factors and other components of the transcriptional complex to modulate gene transcription [4–8].

Estrogens, acting via the ER, play important roles in reg-

ulating the growth, differentiation, and functioning of many reproductive tissues including the uterus, vagina, ovary, oviduct, and mammary gland. In the uterus and mammary gland, estrogens increase proliferation and alter cell properties via, at least in part, the induction of growth factors and growth factor receptors, an effect largely antagonized by antiestrogens [9–13]. Estrogens also have important sites of action in the pituitary, hypothalamus, and specific brain regions, while exerting crucial actions as well on other tissues including bone, liver, and the cardiovascular system [14–16]. Thus these hormones exert their effects on many, diverse target tissues. Because of this diversity of estrogen target tissues, much current interest focuses on trying to understand the basis for the cell context- and promoter context-dependent actions of estrogens and antiestrogens [17–20] and on the development of estrogens and antiestrogens with enhanced tissue-selective activities.

The actions of estrogens are antagonized by antiestrogens, which bind to the ER in a manner that is competitive with estrogen; but antiestrogens usually fail to effectively activate gene transcription [21–25]. The structures of some estrogens and antiestrogens are shown in Figure 1 and, as can be seen, they include both steroidal and nonsteroidal compounds. Antiestrogens typically have a basic or polar side chain, and this side chain is essential for their antiestrogenic activity. Antiestrogens are of particular interest and utility because of their effectiveness in suppressing the estrogen-stimulated proliferation and metastatic activity of ER-containing breast cancers [9–11, 13, 21–25].

¹This research was supported by grants CA18119 and CA60514 from the National Institutes of Health and by grant DAMD17-94-J-4205 from the U.S. Army.

²Correspondence: Dr. Benita S. Katzenellenbogen, Department of Molecular and Integrative Physiology, University of Illinois, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, IL 61801–3704. FAX: (217) 244–9906.

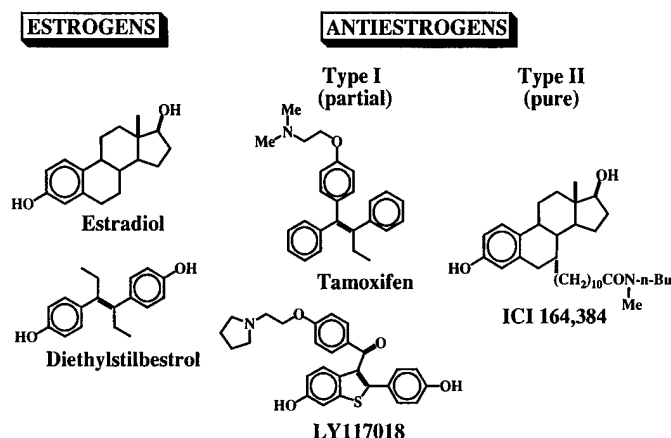


FIG. 1. Structures of several estrogenic and antiestrogenic ligands for the estrogen receptor. The antiestrogens include the nonsteroidal compounds tamoxifen and LY117018 and the steroidal antiestrogen ICI164,384.

ESTROGEN RECEPTOR STRUCTURE-ACTIVITY RELATIONSHIPS

In order to better understand the bioactivities of estrogens and antiestrogens and their differing interactions with the ER, we have focused some of our studies on identifying the regions of the ER that are involved in estrogen and antiestrogen binding and in discriminating between estrogen and antiestrogen [26–31]. Since the hormone-binding domain of the ER is large (more than 250 amino acids), analysis of its structure and its functional complexity is challenging. We have used three approaches for studying estrogen receptor ligand-receptor-response relationships, namely, affinity labeling [32] site-directed mutagenesis, and region-specific chemical mutagenesis of the hormone binding domain.

Many of our studies have analyzed in detail the hormone binding domain of the estrogen receptor, regions E and F, since this domain of the receptor contains both hormone binding and hormone-dependent transactivation functions of the receptor. In our attempts to understand how the receptor discriminates between estrogen and antiestrogen ligands, we have generated and analyzed variant human estrogen receptors with mutations in the ER hormone-binding domain and studied the activity of these receptors on different estrogen-responsive genes in several cell backgrounds when liganded with antiestrogenic or estrogenic ligands. These studies and those of others [17–20] have provided consistent evidence for the promoter-specific and cell-specific actions of the estrogen-occupied and antiestrogen-occupied ER. In addition, although the binding of estrogens and antiestrogens is mutually competitive, studies with ER mutants indicate that some of the contact sites of estrogens and antiestrogens are likely different [29–31, 33]. Our recent studies also reveal that the presence of the carboxy-terminal F domain of the ER is important in the transcription activation and repression activities of antiestrogens and that it affects the magnitude of liganded ER bioactivity in a cell-specific manner [18]. The influence of the

F domain on the agonist/antagonist balance and potency of antiestrogens supports its specific modulatory role in the ligand-dependent interaction of ER with components of the transcription complex. These studies ([18, 26–34], see below) have provided evidence for a regional dissociation of the hormone binding and transcription activation regions in domain E of the receptor and have also shown that mutations in the hormone binding domain and deletions of C-terminal region; result in ligand discrimination mutants, that is, receptors that are differentially altered in their ability to bind and/or mediate the actions of estrogens versus antiestrogens.

A variety of studies [17–20, 26–35] have provided strong documentation that the response of genes to estrogen and antiestrogen depend on several important factors: 1) the nature of the estrogen receptor, i.e., whether it is wild type or variant; 2) the ligand; 3) the promoter; and 4) the cell context. The gene response, in addition, can be modulated by cAMP, growth factors, and agents that affect protein kinases and cell phosphorylation [19, 36–40]. These factors, no doubt, account for differences in the relative agonism/antagonism of antiestrogens like tamoxifen on different genes and in different target cells such as those in breast cancer cells versus uterine or bone cells.

Although both estrogens and antiestrogens bind within the hormone binding domain, the association must differ because estrogen binding activates a transcriptional enhancement function, whereas antiestrogens fully or partially fail in this role. Antiestrogens are believed to act in large measure by competing for binding to the ER and altering the conformation of the ER such that the receptor fails to effectively activate gene transcription. In addition, antiestrogens exert antigrowth factor activities via a mechanism that requires ER but is still not fully understood ([41–43] and refs. therein). Models of antiestrogen action at the molecular level are beginning to emerge, and recent biological studies as well indicate that antiestrogens fall into at least two distinct categories: antiestrogens such as tamoxifen that are mixed or partial agonists/antagonists (type I) and compounds such as ICI164,384 that are complete/pure antagonists (type II). The type I antihormone-ER complexes appear to bind as dimers to estrogen response elements; there, they block hormone-dependent transcription activation mediated by region E of the receptor, but they are believed to have little or no effect on the hormone-independent transcription activation function located in region A/B of the receptor [17]. Thus, they are generally partial or mixed agonist/antagonists, and their action must involve some subtle difference in ligand-receptor interaction, very likely associated with the basic or polar side chain that characterizes the antagonist members of this class. In the case of the more complete antagonists such as ICI164,384, ER conformation must clearly differ from that of the estrogen-occupied ER since some differences in ER binding to DNA and reduction of the ER content of target cells appear to contribute to

[44, 45], but may not fully explain, the pure antagonist character of this antiestrogen [41, 42].

In order to understand how the ER "sees" an antiestrogen as different from an estrogen, we have used site-directed and regional chemical mutagenesis of the ER cDNA to generate estrogen receptors with selected changes in the hormone binding domain. We have been particularly interested in identifying residues in the hormone binding domain important for the binding of estrogen and/or antiestrogen and for the transactivation functions of the receptor, and in elucidating the mechanism by which the ER differently interprets agonistic and antagonistic ligands. Our studies have indicated that selective changes near amino acid 380 and amino acids 520–530 and changes at the C-terminus of the ER result in ER ligand discrimination mutants [18, 26, 29, 30]. These data provide evidence that some contact sites of the receptor with estrogen and antiestrogen differ and that the conformation of the receptor with estrogen and antiestrogen must also be different as a consequence ([29, 33] and refs. therein).

Our observations [26, 31], as well as very important studies by Malcolm Parker and colleagues [34, 46], have shown a separation of the transactivation and hormone-binding functions of the ER with amino acids critical in the transactivation function of the receptor being more C-terminal in domain E (see Fig. 2). Interestingly, some transcriptionally inactive receptors with modifications in this domain E C-terminal activation function 2 (AF-2) region of the ER have potent dominant negative activity, being able to suppress the activity of the wild-type ER in cells [27, 28].

ESTROGEN RECEPTOR CROSS TALK WITH OTHER CELL SIGNALING PATHWAYS

We have observed that protein kinase activators enhance the transcriptional activity of the ER and alter the agonist/antagonist balance of some antiestrogens, suggesting that changes in cellular phosphorylation state should be important in determining the biological effectiveness of the estrogen-occupied ER as well as the effectiveness of antiestrogens as estrogen antagonists. Evidence for cross talk between steroid hormone receptors and signal transduction pathways has been increasing. Expression of activator protein (AP)-1, a transcription factor of the *fos/jun* heterodimer known to mediate the protein kinase (PK)-C pathway [47], was shown to suppress steroid hormone receptor-mediated gene expression [48], most likely through direct protein-protein interaction between steroid receptors and these oncoproteins [49]. In addition, the ovalbumin gene promoter containing a half-palindromic estrogen-responsive element (ERE) was coactivated by ER and *fos/jun* oncoproteins [49–52]. Thus, interaction between these oncoproteins and steroid hormone receptors resulted in cell-specific inhibitory or stimulatory effects on transcriptional activation [50].

Previous studies by us and others [36, 37, 39, 53, 54] doc-

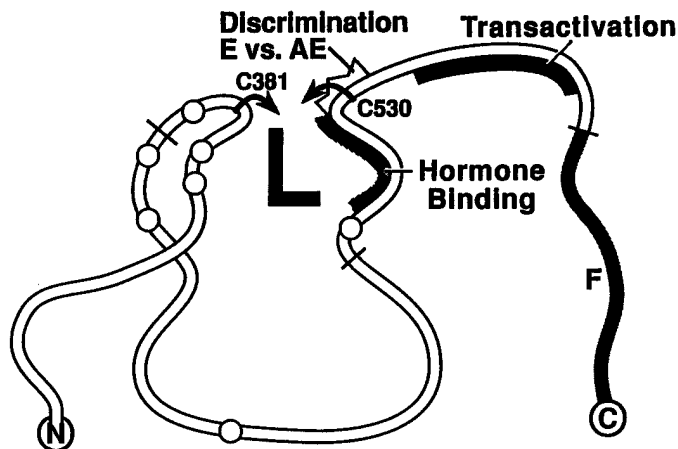


FIG. 2. "Map" of functions in the human estrogen receptor hormone binding domain. Domain E, amino acids 302–553, is shown as is the very C-terminal domain F, amino acids 554–595. Some regions considered to be important in hormone binding, discrimination between estrogen (E) and antiestrogen (AE), and transactivation are highlighted. The ligand (L) is portrayed in a region representing the ligand binding pocket of the receptor. Open circles indicate amino acids in the hormone binding domain where our analyses have shown mutational changes to affect the affinity or stability of hormone binding. See text for description.

umented up-regulation of intracellular progesterone receptor, an estrogen-stimulated protein, by insulin-like growth factor (IGF)-I, epidermal growth factor, phorbol ester, and cAMP in MCF-7 human breast cancer cells and uterine cells. The fact that the stimulation by these diverse agents was blocked by antiestrogen suggested that these agents were presumably acting through the ER pathway [36, 39, 40, 53, 55]. In addition, the fact that protein kinase inhibitors also blocked the effects of estrogen, cAMP, and growth factors suggested the involvement of phosphorylation in these responses. We therefore undertook studies to examine directly whether activators of protein kinases can modulate transcriptional activity of the ER.

In primary cultures of uterine cells, using transient transfection experiments with simple estrogen-responsive reporter genes, we examined the ability of these agents to stimulate ER-mediated gene transcription and also compared the ability of these multiple agents to alter the phosphorylation state of the endogenous uterine ER protein. The results of our study [37] indicate that estrogen, IGF-I, and agents that raise intracellular cAMP are able to stimulate ER-mediated transactivation and ER phosphorylation. The fact that antiestrogen (ICI164,384) evokes a similar increase in ER phosphorylation without a similar increase in transcription activation indicates that an increase in overall ER phosphorylation does not necessarily result in increased transcriptional activity. Also, the observation that transcriptional activation by the ER was nearly completely suppressed by the protein kinase inhibitors H8 and PKI, while the increase in phosphorylation was reduced by 50–75%, indicates that the correlation between transcriptional activation and overall ER phosphorylation is not direct, but it does suggest that some of the effects of estrogen,

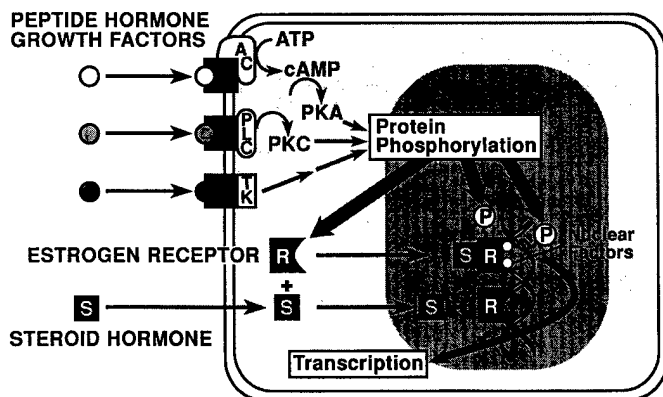


FIG. 3. Model depicting protein kinase-estrogen receptor transcriptional synergism. See text for description.

IGF-I, and cAMP on ER-regulated transactivation are mediated through the activity of protein kinases. Our findings, demonstrating a clear effect of these agents on ER-mediated transactivation, suggest that these agents might also regulate endogenous estrogen target genes, such as that encoding the progesterone receptor, by similar cellular mechanisms.

In order to examine some of the molecular mechanisms controlling transcription of the progesterone receptor gene, we cloned the rat progesterone receptor gene 5'-region and identified two functionally distinct promoters [56]. The two promoters in the rat progesterone receptor gene exhibited differential responsiveness to estradiol and to ER-dependent stimulation by cAMP. The functional differences between these two promoters may lead to altered expression of the A and B progesterone receptor isoforms and, thereby, influence cellular responsiveness to progestins [56].

In MCF-7 human breast cancer cells and other cells, we found that activators of PKA and PKC markedly synergize with estradiol in ER-mediated transcriptional activation and that this transcriptional synergism shows cell- and promoter-specificity [19, 38, 56]. The synergistic stimulation of ER-mediated transcription by estradiol and protein kinase activators did not appear to result from changes in ER content or in the binding affinity of ER for ligand or estrogen response element DNA but, rather, may be a consequence of a stabilization or facilitation of interaction with target components of the transcriptional machinery, possibly either through changes in phosphorylation of ER or other proteins important in ER-mediated transcriptional activation [38]. Of interest also, we have observed that stimulation of the PKA signaling pathway activates the agonist activity of tamoxifen-like but not ICI164,384-like antiestrogens and reduces the effectiveness of tamoxifen as an estrogen antagonist [19]. These findings suggest that agents that enhance intracellular cAMP, such as some growth factors, may contribute to antiestrogen resistance because tamoxifen-like antiestrogens will now be seen by the cell as weak ago-

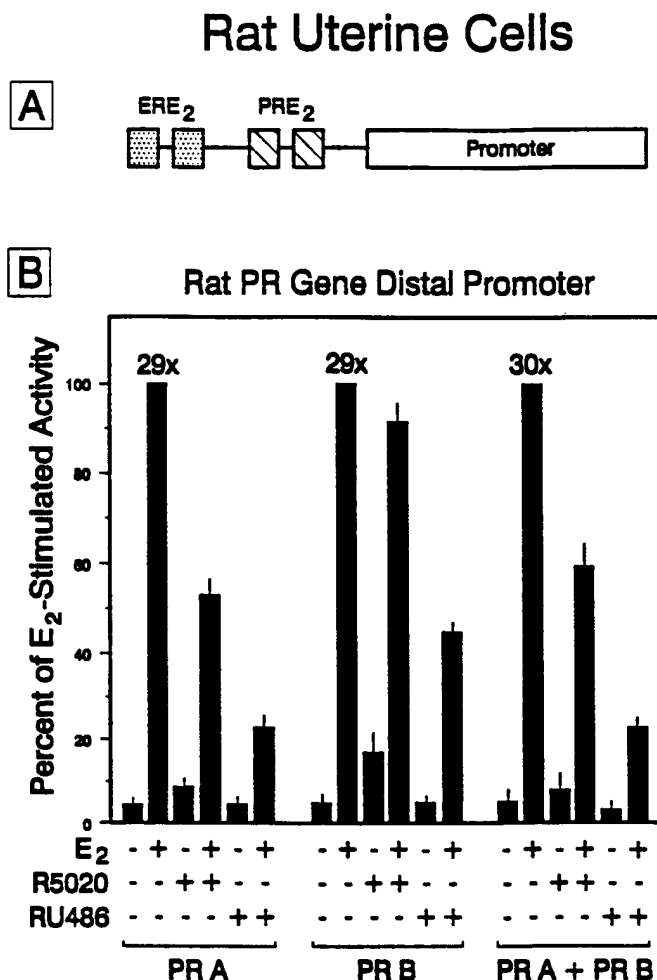


FIG. 4. Repression of ER-mediated transcriptional activity in uterine cells by ligand-occupied progesterone receptors (PRs). A) Schematic diagram of the ERE₂/PRE₂-Promoter-CAT reporter. B) Each 100-mm dish of rat uterine cells was transfected with 500 ng of pRSV-hPRA (labeled PR A), 500 ng of pRSV-hPRB (PR B), or 250 ng each of pRSV-hPRA and pRSV-hPRB (PR A + PR B), in addition to 10 μ g of ERE₂/PRE₂-PR_{Dist}-CAT, 100 ng of pRSV-rER, and 3 μ g of internal control plasmid pCMV β . The cells were treated with one or more of the following as indicated for 24 h: control vehicle, E₂ (10^{-9} M), R5020 (10^{-8} M), and RU486 (10^{-8} M). The CAT activity in each sample was determined. Each bar represents the mean \pm SEM for three or more separate determinations. The fold induction in response to E₂ treatment is indicated above the bars. (From Kraus et al. 1995, ref. [68].)

nists [19, 57]. Related observations have been made with antiestrogens such as RU486 [58–60].

Figure 3 shows a model indicating how we think the protein kinase-estrogen receptor transcriptional synergism might occur. Agents influencing protein kinase pathways may enhance intracellular protein phosphorylation, resulting in either phosphorylation of the ER itself or the phosphorylation of nuclear factors with which the receptor interacts in mediating transcription. Likewise, there is evidence that the steroid hormone itself can alter receptor conformation, increasing the receptor's susceptibility to serve as a substrate for protein kinases [37, 61–64]. There-

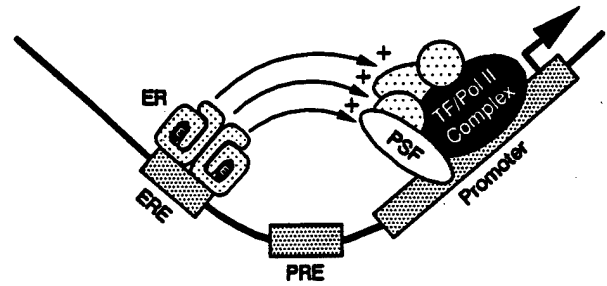
fore, agents that increase phosphorylation may, either through phosphorylation of the ER itself or through phosphorylation of nuclear factors required for ER transcription, result in synergistic activation of ER-mediated transcription.

In direct studies on ER phosphorylation, we have shown that estradiol, the antiestrogens trans-hydroxy-tamoxifen and ICI164,384, as well as PKA and PKC activators enhanced overall ER phosphorylation [63]. Tryptic phosphopeptide patterns of wild-type and domain A/B-deleted receptors and site-directed mutagenesis of several serines involved in known protein kinase consensus sequences allowed us to identify serine 104 and/or serine 106 and serine 118—all three being part of a serine-proline motif—as major ER phosphorylation sites. Mutation of these serines to alanines so as to eliminate the possibility of their phosphorylation resulted in an approximately 50% reduction in transactivation activity in response to estradiol while mutation of only one of these serines showed an approximately 15% decrease in transactivation [63]. Of note, estradiol and antiestrogen-occupied estrogen receptors showed virtually identical two-dimensional tryptic phosphopeptide patterns suggesting similar sites of phosphorylation. In contrast, the cAMP-stimulated phosphorylation likely occurs on different phosphorylation sites as indicated by some of our mutational studies [60]; this aspect remains under investigation in our laboratory. Related studies in COS-1 cells by the Chambon laboratory [61] also identified serine 118 as being a major estrogen-regulated phosphorylation site. In MCF-7 cells, the Notides laboratory has also identified serine 118 as a site of ER phosphorylation but has observed serine 167 to be the most prominent site of phosphorylation in these cells [65]. Aurrichio and coworkers [66] have also provided strong evidence for ER phosphorylation on tyrosine 537. The roles of these phosphorylations in the activities (transcriptional and other) of the ER remains an area of great interest.

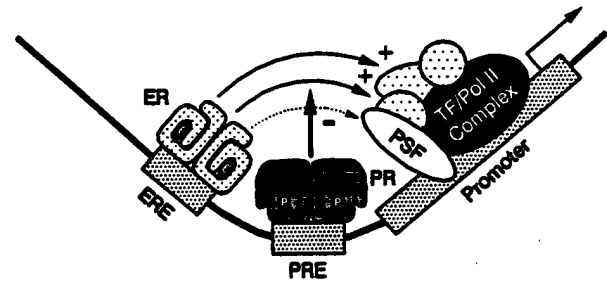
CROSS TALK BETWEEN ESTROGEN RECEPTOR AND PROGESTERONE RECEPTOR SIGNALING SYSTEMS IN MODULATION OF BIOLOGICAL RESPONSES

In addition to interactions with the signaling pathways described above, the ER also has important interrelationships with the progesterone receptor (PR) system in modulation of responses. This has been well documented biologically in many estrogen target tissues. In the uterus, for example, estrogens increase *c-fos* mRNA, cell proliferation, progesterone receptor mRNA and protein levels, gap junction formation, myometrial contractility, and oxytocin receptors, and these effects are largely antagonized by progesterone [12, 56, 67, 68] and references therein). The PR is now known to exist as two isoforms in most species, a smaller A form (PR-A) and a larger B form (PR-B); PR-B contains an N-terminal extension of approximately 164

A) Stimulation of Transcription by Liganded ER



B) Repression by Agonist-Occupied PR



C) Repression by Antagonist-Occupied PR

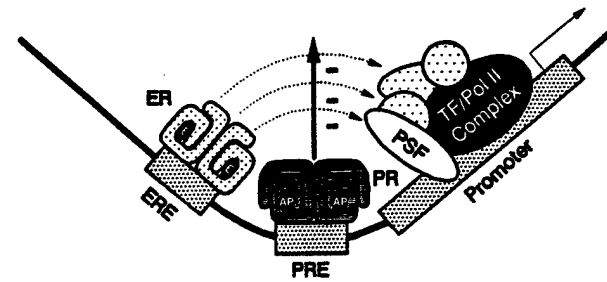


FIG. 5. A model for the repression of ER-mediated transcriptional activity by agonist- and antagonist-occupied PRs. Our findings support a model in which the repression of ER transcriptional activity by liganded PR occurs by quenching. According to this model, liganded PR binds to a site (PRE) distinct from the binding site for ER (ERE) and interferes with the ability of ER to make productive contact with the transcriptional complex. Differences in the magnitude of repression observed for agonist- and antagonist-occupied PRs suggest that agonist-occupied PR only quenches ER-transcription factor interactions that involve the activation function-1 of ER or a promoter-specific component of the ER signaling pathway (PSF), while antagonist-occupied PR quenches a wider range of the ER-transcription factor interactions that occur at the promoter. The individual components of the schematics are labeled. The abbreviations are: AP, antiprogesterin; E, estrogen; ER, estrogen receptor; ERE, estrogen response element; P, progesterin; PR, progesterin receptor; PRE, progesterin response element; PSF, promoter-specific factor; TF/Pol II Complex, general transcriptional machinery. (From Kraus et al., 1995, ref. [68]).

amino acids with exact size varying slightly in different species. PR-A and PR-B have differing biological activities on genes [69–71].

In order to understand better how progestins and anti-progestins are able to antagonize the effects of the estradiol-ER complex, we have developed a simplified model system in which estrogen response elements and progestin response elements have been placed upstream of promoters such as the progesterone receptor gene distal promoter, and the effects of PR-A and PR-B alone or together on ER transcriptional activity can be monitored following transfection into uterine cells or other cells in culture [68]. These studies have shown that liganded PR-A and PR-B can each suppress estradiol-stimulated ER activity (Fig. 4) and that the magnitude of repression depends on several factors: the PR isoform (PR-A more effective than PR-B); the progestin ligand (antiprogestin more effective than progestin agonist); the promoter; and the cell type. The effect of cell background is of particular interest since it has been documented that the inhibitory effect of progestin on estrogen action is not equal in all cell types in the uterus [12]. The repression of ER activity by PR in this model system is not due to a reduction of ER levels or to interference with the binding of ER to its response element since PR is still very suppressive even when the progestin response elements are placed more than 2 kb away from the estrogen response elements [68]. Also the fact that PR is antagonistic of ER action at all concentrations of ER studied argues against squelching due to competition for limiting transcription factors.

Our data is most consistent with quenching [72], wherein PR interferes with the ability of ER to interact effectively with the transcription complex, due perhaps to the recruitment of promoter-specific and cell type-specific inhibitory proteins to the promoter (Fig. 5). Related studies by others have also nicely documented PR-A antagonism of ER action [73] as well as the ability of PR-A to suppress the activity of PR-B [71, 74]. These findings underscore the mounting evidence for the importance of interactions between members of the steroid hormone receptor family and begin to address some of the molecular mechanisms underlying these interactions and cross talk.

ACKNOWLEDGMENTS

I wish to express my gratitude to current and recent past members of this laboratory for their excellent contributions, which have formed the basis of this review. I am also grateful for support of these studies by grants from the NIH and US Army.

REFERENCES

1. Beato M. Gene regulation by steroid hormones. *Cell* 1989; 56:335–344.
2. Evans RM. The steroid and thyroid hormone receptor superfamily. *Science* 1988; 240:889–895.
3. Carson-Jurica MA, Schrader W, O'Malley B. Steroid receptor family: structure and functions. *Endocr Rev* 1990; 11:201–220.
4. Tsai MJ, O'Malley BW. Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 1994; 63:451–486.
5. Green S, Chambon P. The oestrogen receptor: from perception to mechanism. In: Parker M (ed.), *Nuclear Hormone Receptors*. New York: Academic Press; 1991: 15–38.
6. Ryffel GU, Klein-Hitpass L, Druge P, Doebbeling U, Heitlinger E, Cato AC. The estrogen-responsive DNA-element: structure and interaction with the estrogen receptor. *J Steroid Biochem* 1988; 35:219–222.
7. Kumar V, Chambon P. The estrogen receptor binds tightly to its responsive element as a ligand-induced homodimer. *Cell* 1988; 55:145–156.
8. Truss M, Beato M. Steroid hormone receptors: interaction with deoxyribonucleic acid and transcription factors. *Endocr Rev* 1993; 14:459–479.
9. Dickson RB, Lippman ME. Estrogenic regulation of growth and polypeptide growth factor secretion in human breast carcinoma. *Endocr Rev* 1987; 8:29–43.
10. Lippman ME, Dickson RB, Gelmann EP, Knabbe C, Jasid K, Bates S, Swain S. Mechanisms of estrogenic and antiestrogenic regulation of growth of human breast carcinoma. In: Klijn JGM (ed.), *Hormonal Manipulation of Cancer: Peptides, Growth Factors, and New (anti)Steroidal Agents*. New York: Raven Press; 1987: 381–403.
11. Katzenellenbogen BS, Bhakoo HS, Ferguson ER, Lan NC, Tatee T, Tsai TL, Katzenellenbogen JA. Estrogen and antiestrogen action in reproductive tissues and tumors. *Recent Prog Horm Res* 1979; 35:259–300.
12. Katzenellenbogen BS. Dynamics of steroid hormone receptor action. *Annu Review Physiol* 1980; 42:17–35.
13. Read LD, Katzenellenbogen BS. Characterization and regulation of estrogen and progesterone receptors in breast cancer. In: Dickson RB, Lippman ME (eds.), *Genes, Oncogenes and Hormones: Advances in Cellular and Molecular Biology of Breast Cancer*. Nowell, MA: Kluwer-Nijhoff Publishers; 1991: 277–299.
14. Sarrel PM, Lufkin EG, Oursler MJ, Keefe D. Estrogen actions in arteries, bone and brain. *Sci Am Sci Med* 1994; 1:44–53.
15. Kneifel MA, Katzenellenbogen BS. Comparative effects of estrogen and antiestrogen on plasma renin substrate levels and hepatic estrogen receptors in the rat. *Endocrinology* 1981; 108:545–552.
16. Toney TW, Katzenellenbogen BS. Antiestrogen action in the medial basal hypothalamus and pituitary of immature female rats: insights concerning relationships among estrogen, dopamine and prolactin. *Endocrinology* 1986; 119:2661–2669.
17. Berry M, Metzger D, Chambon P. Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the antiestrogen 4-hydroxytamoxifen. *EMBO J* 1990; 9:2811–2818.
18. Montano MM, Müller V, Trobaugh A, Katzenellenbogen BS. The carboxy-terminal F domain of the human estrogen receptor: role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 1995; 9:814–825.
19. Fujimoto N, Katzenellenbogen BS. Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen-selectivity and promoter-dependence. *Mol Endocrinol* 1994; 8:296–304.
20. Tzukerman MT, Esty A, Santiso-Mere D, Danielian P, Parker MG, Stein RB, Pike JW, McDonnell DP. Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol Endocrinol* 1994; 9:21–30.
21. Katzenellenbogen BS, Miller MA, Mullick A, Sheen YY. Antiestrogen action in breast cancer cells: modulation of proliferation and protein synthesis, and interaction with estrogen receptors and additional antiestrogen binding sites. *Breast Cancer Res Treat* 1985; 5:231–243.
22. Jordan VC, Murphy CS. Endocrine pharmacology of antiestrogens as antitumor agents. *Endocr Rev* 1990; 11:578–610.
23. Santen R, Manni A, Harvey H, Redmond C. Endocrine treatment of breast cancer in women. *Endocr Rev* 1990; 11:221–265.
24. Katzenellenbogen BS, Kendra KL, Norman MJ, Berthois Y. Proliferation, hormonal responsiveness, and estrogen receptor content of MCF-7 human breast cancer cells grown in the short-term and long-term absence of estrogens. *Cancer Res* 1987; 47:4355–4360.
25. Katzenellenbogen BS. Antiestrogen resistance: mechanisms by which breast cancer cells undermine the effectiveness of endocrine therapy. *J Natl Cancer Inst* 1991; 83:1434–1435.
26. Wrenn CK, Katzenellenbogen BS. Structure-function analysis of the hormone binding domain of the human estrogen receptor by region-specific mutagenesis and phenotypic screening in yeast. *J Biol Chem* 1993; 268:24089–24098.
27. Ince BA, Zhuang Y, Wrenn CK, Shapiro DJ, Katzenellenbogen BS. Powerful dominant negative mutants of the human estrogen receptor. *J Biol Chem* 1993; 268:14026–14032.
28. Ince BA, Schodin DJ, Shapiro DJ, Katzenellenbogen BS. Repression of endogenous estrogen receptor activity in MCF-7 human breast cancer cells by dominant negative estrogen receptors. *Endocrinology* 1995; 136:3194–3199.
29. Pakdel F, Katzenellenbogen BS. Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J Biol Chem* 1992; 267:3429–3437.
30. Pakdel F, Reese JC, Katzenellenbogen BS. Identification of charged residues in an N-terminal portion of the hormone binding domain of the human estrogen receptor important in transcriptional activity of the receptor. *Mol Endocrinol* 1993; 7:1408–1417.

31. Katzenellenbogen BS, Bhardwaj B, Fang H, Ince BA, Pakdel F, Reese JC, Schodin DJ, Wrenn CK. Hormone binding and transcription activation by estrogen receptors: analyses using mammalian and yeast systems. *J Steroid Biochem Mol Biol* 1993; 47:39-48.
32. Harlow KW, Smith DN, Katzenellenbogen JA, Greene GL, Katzenellenbogen BS. Identification of cysteine-530 as the covalent attachment site of an affinity labeling estrogen (ketononestrol aziridine) and antiestrogen (tamoxifen aziridine) in the human estrogen receptor. *J Biol Chem* 1989; 264:17476-17485.
33. Danielian PS, White R, Hoare SA, Fawell SF, Parker MG. Identification of residues in the estrogen receptor that confer differential sensitivity to estrogen and hydroxytamoxifen. *Mol Endocrinol* 1993; 7:232-240.
34. Danielian PS, White R, Lees JA, Parker MG. Identification of a conserved region required for hormone dependent transcriptional activation by steroid hormone receptors. *EMBO J* 1992; 11:1025-1033.
35. Reese JC, Katzenellenbogen BS. Characterization of a temperature-sensitive mutation in the hormone binding domain of the human estrogen receptor: studies in cell extracts and intact cells and their implications for hormone-dependent transcriptional activation. *J Biol Chem* 1992; 267:9868-9873.
36. Aronica SM, Katzenellenbogen BS. Progesterone receptor regulation in uterine cells: stimulation by estrogen, cyclic adenosine 3',5'-monophosphate and insulin-like growth factor I and suppression by antiestrogens and protein kinase inhibitors. *Endocrinology* 1991; 128:2045-2052.
37. Aronica SM, Katzenellenbogen BS. Stimulation of estrogen receptor-mediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic adenosine monophosphate, and insulin-like growth factor-I. *Mol Endocrinol* 1993; 7:743-752.
38. Cho H, Katzenellenbogen BS. Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Mol Endocrinol* 1993; 7:441-452.
39. Katzenellenbogen BS, Norman MJ. Multihormonal regulation of the progesterone receptor in MCF-7 human breast cancer cells: interrelationships among insulin/insulin-like growth factor-I, serum, and estrogen. *Endocrinology* 1990; 126:891-898.
40. Cho H, Aronica SM, Katzenellenbogen BS. Regulation of progesterone receptor gene expression in MCF-7 breast cancer cells: a comparison of the effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology* 1994; 134:658-664.
41. Reese JR, Katzenellenbogen BS. Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor overexpressed in mammalian cells. *Nucleic Acids Res* 1991; 19:6595-6602.
42. Reese JC, Katzenellenbogen BS. Examination of the DNA binding abilities of estrogen receptor in whole cells: implications for hormone-independent transactivation and the action of the pure antiestrogen ICI164,384. *Mol Cell Biol* 1992; 12:4531-4538.
43. Freiss G, Vignon F. Antiestrogens increase protein tyrosine phosphatase activity in human breast cancer cells. *Mol Endocrinol* 1994; 8:1389-1396.
44. Dauvois S, Danielian PS, White R, Parker MG. Antiestrogen ICI164,384 reduces cellular estrogen receptor content by increasing its turnover. *Proc Natl Acad Sci USA* 1992; 89:4037-4041.
45. Fawell SE, White R, Hoare S, Sydenham M, Page M, Parker MG. Inhibition of estrogen receptor-DNA binding by the "pure" antiestrogen ICI164,384 appears to be mediated by impaired receptor dimerization. *Proc Natl Acad Sci USA* 1990; 87:6883-6887.
46. Fawell SE, Lees JA, White R, Parker MG. Characterization and colocalization of steroid binding and dimerization activities in the mouse estrogen receptor. *Cell* 1990; 60:953-962.
47. Angel P, Imagawa M, Chiu M, Stein B, Imbra RJ, Rahmsdorf HJ, Jonat C, Herrlich P, Karin M. Phorbol ester-inducible genes contain a common *cis*-acting element recognized by a TPA-modulator trans-acting factor. *Cell* 1987; 49:729-739.
48. Schüle R, Rangarajan P, Klierer S, Ransone LJ, Bolado J, Yang N, Verma IM, Evans RM. Functional antagonism between oncoprotein *c-jun* and the glucocorticoid receptor. *Cell* 1990; 62:1217-1226.
49. Yang-Yen H, Chambard J, Sun Y, Smeal T, Schmidt TJ, Drouin J, Karin M. Transcriptional interference between *c-jun* and the glucocorticoid receptor: mutual inhibition of DNA binding due to direct protein-protein interaction. *Cell* 1990; 62:1205-1215.
50. Gaub M, Ballard M, Scheuer I, Chambon P, Sassone-Corsi P. Activation of the ovalbumin gene by the estrogen receptor involves the *fos-jun* complex. *Cell* 1990; 53:1267-1276.
51. Shemshedini L, Knauth R, Sassone-Corsi P, Pornon A, Gronemeyer H. Cell-specific inhibitory and stimulatory effects of *fos* and *jun* on transcription activation by nuclear receptors. *EMBO J* 1991; 10:3839-3849.
52. Strähle U, Schmid W, Schütz G. Synergistic action of the glucocorticoid receptor with transcription factors. *EMBO J* 1988; 7:3389-3395.
53. Sumida C, Lecerf F, Pasqualini JR. Control of progesterone receptors in fetal uterine cells in culture: effects of estradiol, progestins, antiestrogens, and growth factors. *Endocrinology* 1988; 122:3-11.
54. Sumida C, Pasqualini JR. Stimulation of progesterone receptors by phorbol ester and cyclic AMP in fetal uterine cells in culture. *Mol Cell Endocrinol* 1990; 69:207-215.
55. Sumida C, Pasqualini JR. Antiestrogen antagonizes the stimulatory effect of epidermal growth factor on the induction of progesterone receptor in fetal uterine cells in culture. *Endocrinology* 1989; 124:591-597.
56. Kraus WL, Montano MM, Katzenellenbogen BS. Cloning of the rat progesterone receptor gene 5' region and identification of two functionally distinct promoters. *Mol Endocrinol* 1993; 7:1603-1616.
57. Gottardis MM, Jordan VC. Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res* 1988; 48:5183-5187.
58. Groul DJ, Altschmeid J. Synergistic induction of apoptosis with glucocorticoids and 3',5'-cyclic adenosine monophosphate reveals agonist activity by RU486. *Mol Endocrinol* 1993; 7:104-113.
59. Sartorius CA, Tung L, Takimoto GS, Horwitz KB. Antagonist-occupied human progesterone receptors bound to DNA are functionally switched to transcriptional agonists by cAMP. *J Biol Chem* 1993; 268:9262-9266.
60. Beck CA, Weigel NL, Moyer ML, Nordeen SK, Edwards DP. The progesterone antagonist RU486 acquires agonist activity upon stimulation of cAMP signaling pathways. *Proc Natl Acad Sci USA* 1993; 90:4441-4445.
61. Ali S, Metzger D, Bornert J-M, Chambon P. Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J* 1993; 12:1153-1160.
62. Denton RR, Koszewski NJ, Notides AC. Estrogen receptor phosphorylation: hormonal dependence and consequence on specific DNA binding. *J Biol Chem* 1992; 267:7263-7268.
63. Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS. Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem* 1994; 269:4458-4466.
64. Lahooti H, White R, Danielian PS, Parker MG. Characterization of ligand-dependent phosphorylation of the estrogen receptor. *Mol Endocrinol* 1994; 8:182-188.
65. Arnold SF, Obourn JD, Jaffe H, Notides AC. Serine 167 is the major estradiol-induced phosphorylation site on the human estrogen receptor. *Mol Endocrinol* 1994; 8:1208-1214.
66. Castoria G, Migliaccio A, Green S, Di Domenico M, Chambon P, Auricchio F. Properties of a purified estradiol-dependent calf uterus tyrosine kinase. *Biochemistry* 1993; 32:1740-1750.
67. Kraus WL, Katzenellenbogen BS. Regulation of progesterone receptor gene expression and growth in the rat uterus: modulation of estrogen actions by progesterone and sex steroid hormone antagonists. *Endocrinology* 1993; 132:2371-2379.
68. Kraus WL, Weis KE, Katzenellenbogen BS. Inhibitory cross-talk between steroid hormone receptors: differential targeting of estrogen receptor in the repression of its transcriptional activity by agonist- and antagonist-occupied progestin receptors. *Mol Cell Biol* 1995; 15:1847-1857.
69. Kastner P, Krust A, Turcotte B, Stropp U, Tora L, Gronemeyer H, Chambon P. Two distinct estrogen-regulated promoters generate transcripts encoding two functionally different human progesterone receptor forms A and B. *EMBO J* 1990; 9:1603-1614.
70. Tora L, Gronemeyer H, Turcotte B, Gaub MP, Chambon P. The N-terminal region of the chicken progesterone receptor specifies target gene activation. *Nature* 1988; 333:185-188.
71. Vegeto E, Shahbaz MM, Wen D X, Goldman ME, O'Malley BW, McDonnell DP. Human progesterone receptor A form is a cell- and promoter-specific repressor of human progesterone receptor B function. *Mol Endocrinol* 1993; 7:1244-1255.
72. Levine M, Manley JL. Transcriptional repression of eukaryotic promoters. *Cell* 1989; 59:405-408.
73. McDonnell DP, Goldman ME. RU486 exerts antiestrogenic activities through a novel progesterone receptor A form-mediated mechanism. *J Biol Chem* 1994; 269:11945-11949.
74. Tung LT, Mohamed MK, Hoeffler JP, Takimoto GS, Horwitz KB. Antagonist-occupied human progesterone B-receptors activate transcription without binding to progesterone response elements and are dominantly inhibited by A-receptors. *Mol Endocrinol* 1993; 7:1256-1265.

MINIREVIEW

Tripartite Steroid Hormone Receptor Pharmacology: Interaction with Multiple Effector Sites as a Basis for the Cell- and Promoter-Specific Action of These Hormones

John A. Katzenellenbogen, Bert W. O'Malley,
and Benita S. Katzenellenbogen

Department of Chemistry (J.A.K.)
Departments of Molecular and Integrative Physiology,
and Cell and Structural Biology (B.S.K.)
University of Illinois
Urbana, Illinois 61801

Department of Cell Biology (B.W.O.)
Baylor College of Medicine
Houston, Texas 77030

INTRODUCTION

The selective action that steroid hormones and the hormones for the other nuclear receptors have in different tissues and on different responses is well known. In fact, this recognized selectivity forms the basis for major efforts, currently underway in the pharmaceutical industry and at universities, toward the development of new, synthetic hormones whose profile of desired activities is optimized for specific therapeutic and preventative applications. This commentary will examine the pharmacological mechanisms that underlie this selectivity.

The study of steroid hormone pharmacology poses particular challenges. *In vivo*, many steroids have pleiotropic activity, displaying a variety of effects in different tissues. Even in cell-based *in vitro* systems, attempts to investigate the molecular basis for steroid hormone action and the selectivity of this action are confounded by the fact that the genomic responses elicited by these ligands can be both primary and secondary (*i.e.* cascade) responses. In the latter situation, the correlation between molecular interaction and response is complex and indirect; this makes it difficult to clearly determine what interactions define the pharmacological parameters of potency and bio-

character (biological character, *i.e.* agonist vs. antagonist activity) of a specific hormone. Even the genomic actions vary: most involve direct receptor-DNA interaction, but some appear to be mediated via interaction of receptor with other DNA-binding proteins. Steroid hormones may also exert nongenomic effects, some of which may still involve the receptor. In this commentary, we are focusing on the genomic action of steroid hormones that involves the regulation of gene transcription mediated by nuclear receptors.

THREE MECHANISMS FOR STEROID HORMONE SELECTIVITY

The selectivity that steroid and other hormones for nuclear receptors display at three different levels—the tissue, the cell, and the gene—may be mediated by three distinct mechanisms (Table 1): 1) ligand-based selectivity, 2) receptor-based selectivity, and 3) effector site-based selectivity. Since the first two mechanisms are well recognized, they will be described only briefly; the third mechanism merits careful examination and will be discussed in greater detail.

Ligand-Based Selectivity

By this mechanism, selectivity at the tissue or cell level may be achieved by differences in pharmacokinetics

Table 1. Types of Selectivity in the Action of Ligands for Nuclear Hormone Receptors

Type of selectivity	Components			Level of Selectivity			Mechanism
	Ligand	Receptor	Effector	Tissue	Cell	Gene	
Ligand-based	Different	Same	Same	Yes	Yes	No	Ligand(s) undergoes different metabolism in different tissues/cells (selective bioactivation; selective bioinactivation)
Receptor-based	Same	Different	Same	Yes	Yes	No	Composition of receptors (concentration, subtypes, isoforms, variants) is different in different tissues/cells
Effector-based	Same	Same	Different	Yes	Yes	Yes	The same ligand(s) and same receptor(s) experience different interactions at different effector sites regulating gene transcription

or differential ligand metabolism. The same hormone or set of hormones is presented to different target tissues through the circulation, but their relative amounts within the cell are altered by differential uptake or metabolism—at the level of the target tissue cell. The differential metabolism mechanism may involve either a bioactivation, such as the tissue-selective conversion of the naturally circulating androgen testosterone to the more potent 5α -dihydrotestosterone by the action of 5α -reductase (1), or a bioinactivation, such as the selective oxidation of cortisol, but not aldosterone, by an 11β -dehydrogenase found in tissues that respond to mineralocorticoids (2). Thus, this differential metabolism creates a ligand-based selectivity in which the *same receptor* in different target tissues or cells can experience a *different complexation of hormones* and thereby mediate responses in a selective manner (cf. Table 1).

Receptor-Based Selectivity

By the second mechanism, different target tissues experiencing the same hormones may respond in a selective fashion because they have a different composition of receptors. This difference could include variations in the concentrations or ratios of receptor subtypes, isoforms, or splice variants or receptors having different states of covalent modification (e.g. phosphorylation) (Refs. 3–5 and references cited therein). This mechanism is especially well represented in the retinoid, thyroid hormone, and vitamin D₃ receptor systems, where multiple receptor forms are found, and different patterns of receptor dimerization are known to be dependent upon both the structure and composition of the ligands and the response elements (6, 7). It appears to be important in the progesterone receptor system, where progesterone receptor A and B isoforms are known to differ in their ability to activate genes (8). Additionally, progesterone receptor A can act as an inhibitor of progesterone receptor B transcriptional activity (9–11). Receptor-based selectivity may also play a role among androgen receptors and glucocorticoid receptors, where two isoforms have been reported (12, 13), and even in some estro-

gen-responsive cells where full length estrogen receptor and splice variants may coexist (14–18). In these systems, the *same hormone* or set of hormones could effect tissue- or cell-selective action as a result of the *different complexation of receptors* present in different target sites (cf. Table 1).

Effector Site-Based Selectivity

Although the former two mechanisms may explain some of the tissue- and cell-selective actions of steroid and related hormones, the selectivity of these hormones clearly also derives from a deeper level. Even in cases where there seems to be no differential hormone metabolism in target tissues and only a single receptor is involved (*i.e.* mechanisms 1 and 2 are not operating), hormones for nuclear receptors are capable of selective action. Most striking is the different biocharacter that some estrogens and their analogs show in terms of certain responses elicited in different target tissues.

For example, in the rat, we have shown that the antiestrogens tamoxifen, nafoxidine, and CI-628 are partial agonists/antagonists in the modulation of pituitary PRL and dopamine turnover in the medial basal hypothalamus (19) and of various responses in the uterus (uterine weight gain, progesterone receptor induction, and plasminogen activator and peroxidase activity stimulation) (20–23), yet they are full agonists in increasing plasma renin substrate in liver (24). In women, raloxifene (originally called keoxifene) shows tissue-selective differences, with strong agonist activity indicated by maintenance of bone density and estrogenic blood lipid profiles, but little stimulation of the uterus (25–30). Tamoxifen therapy in postmenopausal women with breast cancer has also revealed estrogen-like actions of this agent on bone mineral density (31) and lipoprotein levels (32), as well as estrogen-like stimulation of the uterus (33–35), yet little agonism occurs in the breast, where tamoxifen reduces recurrence of breast cancer (36). In contrast, the estradiol-based antiestrogens ICI164,384 and ICI182,780 have almost complete antagonist character in all estrogen target tissues examined, both in experimental cell and

animal systems and in clinical trials in women (37, 38). Regardless of their varying level of agonist or antagonist character in different tissues, these compounds appear to be acting through a single receptor, the estrogen receptor.

The study of the molecular details of steroid hormone pharmacology has been assisted greatly by the development of transient transfection assays, whereby one can achieve independent control over four critical variables, the ligand, the receptor, the gene context, and the cellular milieu. Transfection of estrogen-responsive promoter-reporter constructs into different cells has enabled the regulation of specific genes to be studied in these different cell backgrounds. However, one should keep in mind that hormonal regulation of transfected gene constructs does not always precisely mimic that observed in the native gene context, as local chromatin architecture may be different (39, 40). Nevertheless, the results of these investigations illustrate clearly that *cell-specific factors* can affect the biocharacter (agonist/antagonist balance) of different estrogens.

In studies in several cell types with either wild type or variant estrogen receptors lacking their C-terminal F domains (ΔF), we have observed that the response of these receptors to estrogen and antiestrogen ligands is markedly influenced by cell context (41). For example, in Chinese hamster ovary (CHO) cells and MDA-MB-231 human breast cancer cells expressing wild type or ΔF estrogen receptors, estradiol stimulated equally transcription of several estrogen-responsive promoter reporter gene constructs. By contrast, in HeLa human cervical cancer cells and 3T3 mouse fibroblast cells, the ΔF estrogen receptor exposed to estradiol was much less effective than wild type estrogen receptor in stimulating transcription, and antiestrogens were less potent in suppressing estrogen-stimulated transcription by the ΔF estrogen receptor. These differences in response of the ΔF and wild type estrogen receptor to estrogen or antiestrogen do not appear to be due to a change in receptor expression level, binding affinity for ligands, or binding to estrogen response element DNA. Rather, our data support the supposition that the conformation of the receptor-ligand complex is different with estrogen vs. antiestrogen and with wild type vs. ΔF estrogen receptor, such that its potential for interaction with protein cofactors or transcription factors is different and is markedly influenced by cell context (41). Likewise, studies by McDonnell and co-workers (42, 43) have provided extensive documentation of the fact that cell background profoundly influences estrogen receptor transcriptional response to ligand. Several groups have shown as well that the transcriptional response of progesterone receptor A and B isoforms to progestin ligands is greatly influenced by the test cell used, as is the ability of progesterone receptor to repress estrogen receptor transcriptional activity (44, 45). This very likely reflects the differing activities of the different activation functions (AF-1, AF-2, and others) in a receptor, a concept

nicely documented by Berry *et al.* in 1990 (46) for the estrogen receptor to explain the differing agonist/antagonist activity of tamoxifen in different cells (see below).

Even within the same cell, it is possible to effect selective stimulation of different endogenous genes with different ligands. For example, in estrogen receptor-containing MCF-7 human breast cancer cells, antiestrogens such as tamoxifen are pure antagonists for plasminogen activator activity (47, 48) but show weak agonism for other responses, such as pS2 (39) and progesterone receptor induction (47, 49). By transfecting estrogen-responsive promoter-reporter constructs into these (MCF-7) cells, it has been shown that antiestrogens exhibit *promoter-specific agonism* (50). This promoter-specific agonistic activity of antiestrogens is also observed when these estrogen-responsive promoters are transfected, along with wild type estrogen receptor, into a variety of estrogen receptor-negative cells (41, 42). Further evidence for gene-specific agonist and antagonist properties of tamoxifen and other antiestrogens is evident from studies in GH4 and GC3 pituitary tumor cells, where these compounds act like a full estrogen on some responses yet as an antagonist of estrogen stimulation of other responses (51, 52).

The phenomenon of promoter-specific agonism is particularly well highlighted by the observations made in bone cells with antiestrogens using two different estrogen receptor-dependent responses. Here, raloxifene, a benzothiophene compound typically considered an antiestrogen, tamoxifen, and ICI 164,384 are, in fact, stronger agonists of transforming growth factor- β 3 (TGF β 3) promoter activity than estradiol. By contrast, in the same MG-63 osteosarcoma cell cultures, all three ligands act as pure antagonists of the dramatic stimulation of the reporter gene construct estrogen response element-vitellogenin-chloramphenicol acetyl transferase by estradiol (53, 54). Interestingly, the nucleotide sequences comprising the estrogen response elements in these two genes (TGF β 3 and vitellogenin) are quite different, vitellogenin containing a palindromic consensus estrogen response element and TGF β 3 quite a different nucleotide sequence; only the former was shown to bind the estrogen receptor in gel shift assays. The DNA-binding domain of the estrogen receptor appears not to be required for raloxifene induction of the TGF β 3 gene. Since the estrogen receptor does not bind directly to this unusual estrogen response element, an additional DNA-binding protein that tethers estrogen receptor to this enhancer is implied (54). Thus, at least some of the proteins interacting with the ligand-receptor complex at these two promoters would be predicted to be different, to account for the reversed pharmacology displayed by these estrogen receptor ligands at these two genes.

As was mentioned earlier, these findings are also mirrored in tissue-specific differences in the estrogen agonist/antagonist character of these compounds in

vivo. Tamoxifen and raloxifene are strong estrogen-like agonists for bone density maintenance in rats and women. They have either some (tamoxifen) or little to no (raloxifene) stimulatory effect on uterine proliferation, yet they are full antagonists of estrogen-stimulated breast cancer cell proliferation and responses such as induction of plasminogen activator activity in breast cancer cells. These observations indicate that these ligands are "selective estrogen receptor modifiers" (27, 30), displaying estrogen agonist or antagonist activity that is dependent on the particular cell and gene endpoint.

Such observations form the basis for efforts currently being directed at the development of tissue-selective estrogen/antiestrogen agents with specific profiles optimal for treatment of women with breast cancer and for postmenopausal bone loss (osteoporosis) prevention: no agonism on breast or uterus; estrogen agonism on bone (for good bone maintenance), the cardiovascular system, and some aspects of liver function (such as blood lipid profile). Such compounds would exploit what is now known about the gene- and cell-selective actions of hormonal ligands and the importance of effector site components in a ligand's pharmacological profile (see below). Thus, in some systems, the same ligand working through a single receptor can elicit a different spectrum of responses from different genes in hormone-responsive cells (cf. Table 1). These gene-selective actions cannot be readily explained by either of the first two mechanisms (see above).

EVOLVING MODELS FOR THE ROLE OF THE RECEPTOR IN STEROID HORMONE ACTION—MOLECULAR INTERACTIONS THAT DEFINE POTENCY AND BIOCHARACTER

The Pharmacology of Classical Bipartite (Ligand-Receptor) Systems

The development of the concept of "receptors" in classical pharmacology arose from the need to postulate a molecular species that served as the interface between a drug or hormone and the behavioral or physiological responses that it evoked. The original receptor concept, conceived by Ehrlich (55) and Langley (56), formalized by Clark (57) and Gaddum (58), and refined by Ariens and Simonis (59) and Stephenson (60) was basically an operational one. Nevertheless, it permitted the different dose-response relationships displayed by various drugs and hormones to be related to a hypothesized molecular interaction that these species had as ligands for the receptor. The activity of these ligands could then be interpreted in terms of the pharmacological parameters "potency" and "biocharacter": potency, measured as the median efficacy (EC_{50} , or median inhibition, IC_{50}), was related to the ligand's affinity for the receptor; biocharacter (*i.e.* agonist vs. antagonist character), estimated by the

degree to which this binding resulted in activation of the receptor to elicit a response, was related to the ligand's efficacy or intrinsic activity.

At an operational level, the receptor was considered to represent the interface where the molecular interactions with the ligand ceased and the biological responses began. In such a bipartite model, involving only the ligand and the receptor, the ligand plays a role much like that of an allosteric effector of an enzyme, altering the conformation of the receptor and thereby *directly* altering its capacity to elicit the response. The conceptual features of such a bipartite scheme are illustrated in Fig. 1. The key issue is that the receptor itself embodies two functions, the capacity to bind a ligand and the capacity to initiate or effect a response as a direct consequence of that binding. The implications of the bipartite model are subtle but important: since the ligand is controlling the shape and the function of the receptor directly, one can assign to each ligand a unique characteristic potency and biocharacter (Table 2).

The Identification of Steroid Receptors and Their Genomic Action

The preparation of high specific activity radiolabeled steroid hormones more than 3 decades ago led to the identification of specific, high affinity binding proteins in target tissues for steroid hormones (61). Since the binding affinity that these proteins showed for various ligands reflected the biological potency of these ligands quite accurately, the binding proteins were

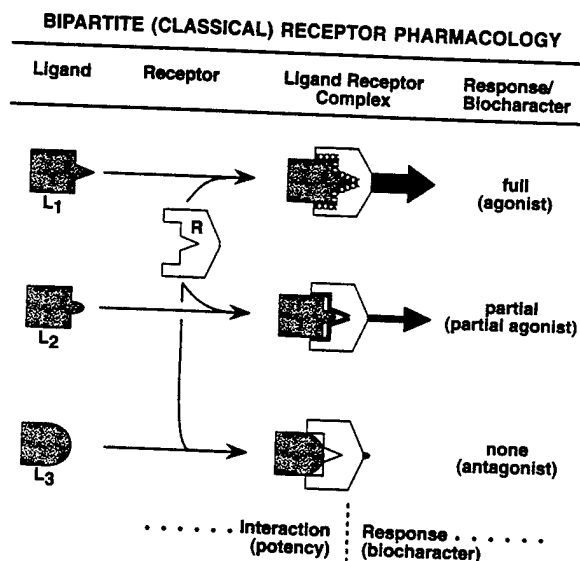


Fig. 1. Classical Bipartite (Ligand-Receptor) Pharmacology
This simple conceptual scheme illustrates how the response to a hormone might be mediated by a bipartite interaction between the hormone, acting as a ligand (L) and a receptor (R). In such a bipartite system, the effect of each hormonal ligand is determined directly by the nature of its interaction with the receptor. Thus, unique potency and biocharacter descriptors can be assigned to each hormone.

Table 2. Pharmacology in Bipartite vs. Tripartite Receptor Systems

Pharmacological characteristic	Bipartite scheme	Tripartite scheme
Potency (EC_{50} , IC_{50})	Determined by the affinity of the L-R interaction	Determined by <i>both</i> L-R binding affinity and L-R coupling with effectors
Biocharacter (efficacy, intrinsic activity)	Determined by effectiveness the conformation of the L-R complex itself	Determined by <i>both</i> shape of the L-R complex and the effectiveness of its coupling with various effector sites
Uniqueness of pharmacological characteristics	Potency and biocharacter can be uniquely assigned to each ligand	Potency and biocharacter are effector dependent; they are not inherent characteristics of a ligand, and cannot be assigned without reference to a particular response

soon referred to as "receptors." Results from other biochemical studies elucidated the principal action of steroids as the activation of gene transcription (for example, Refs. 62–66). The role of these binding proteins as receptors, linking the binding interaction of the steroid with the biochemical response of transcription activation, still appeared to be clear. Nevertheless, it was evident even then that there would be other molecular elements within the cell with which the ligand-receptor complex would need to interact in order for the effect—the transduction of the signal—to continue (67, 68).

In the most recent decade, great strides have been made in determining the structure of these receptors and in elucidating the details of their action. They are multidomain proteins that engage in multiple interactions in the process of eliciting their genetic transcriptional activation or repression responses. In some cases they interact with themselves as homodimers or with other related receptor partners as heterodimers. At each regulated gene, these receptors may interact with DNA via response elements of varying sequence and distribution, with transcription factors and other components of the general transcription apparatus, and with various other activator and adaptor (co-activator and co-repressor) proteins that are associated with the transcriptional regulation of that particular gene (reviewed in Refs. 69–73).

This proliferation of molecular constituents that link ligand to response necessitates a reexamination of the simplistic application of the term "receptor" to this intracellular ligand-binding protein. In fact, *in the nuclear receptor signal transduction cascade, it is no longer so clear where the effect of ligand "interaction" ceases and the biological "response" begins*, and thereby just what molecular entity or entities linking interaction and response merits the appellation "receptor" in the classical pharmacological sense. The "interaction" by which a ligand effects a response in the steroid hormone system is clearly a multipartite phenomenon, one that is much more complex than the bipartite interaction originally envisioned as simply the binding of a hormone to a receptor protein. The proliferation of such terms and phrases as "cell and promoter context," "gene-specific effects," "intracellular

receptor pharmacology," "post-receptor events in ligand discrimination," or the "different biology of various receptor-ligand complexes" to describe steroid hormone pharmacology is a reflection of the inadequacy of the current use of the classical terms "agonist," "antagonist," and "receptor" to describe the selective action of hormones at the level of the cell and gene.

The Tripartite (Ligand-Receptor-Effector) Systems

A tripartite scheme that embodies elements which seem more appropriate to describe steroid hormone molecular pharmacology is shown in Fig. 2 (Table 2). Whereas the bipartite scheme (Fig. 1) embodied the ligand binding and the response initiation functions in one entity, in the tripartite scheme these functions are assigned to separate entities—ligand binding to the receptor, and response initiation to the *effector*. Thus, where there were two partners that defined pharmacology, there are now three: the ligand, the receptor, and the effector.

Tripartite or ligand-receptor-effector schemes were proposed some time ago for certain other signal transduction systems, and more recently even for some glucocorticoid receptor-mediated responses (73a), especially those that showed a discordance between ligand potency in response stimulation (measured as the EC_{50}) and ligand binding to receptor [measured as the dissociation constant (K_d)]. For example in the "spare receptor" hypothesis, the effector was proposed as a response-limiting step beyond the receptor that could account for this potency/binding disjunction (74–77). Many of these systems are now known to be tripartite in reality. For example, the action of extracellular ligands on transmembrane G protein-coupled receptors results in second messenger induction via G protein activation that operates through intracellular sites (78). More recently, the action of immunosuppressants in T cells has been shown to be tripartite; it begins with the binding of the immunosuppressants by immunophilins and then proceeds with the interaction of this complex, as a composite ligand, with the phosphatase calcineurin (79). What is

DIFFERENT MODES OF NUCLEAR RECEPTOR ACTIVATION OF GENES

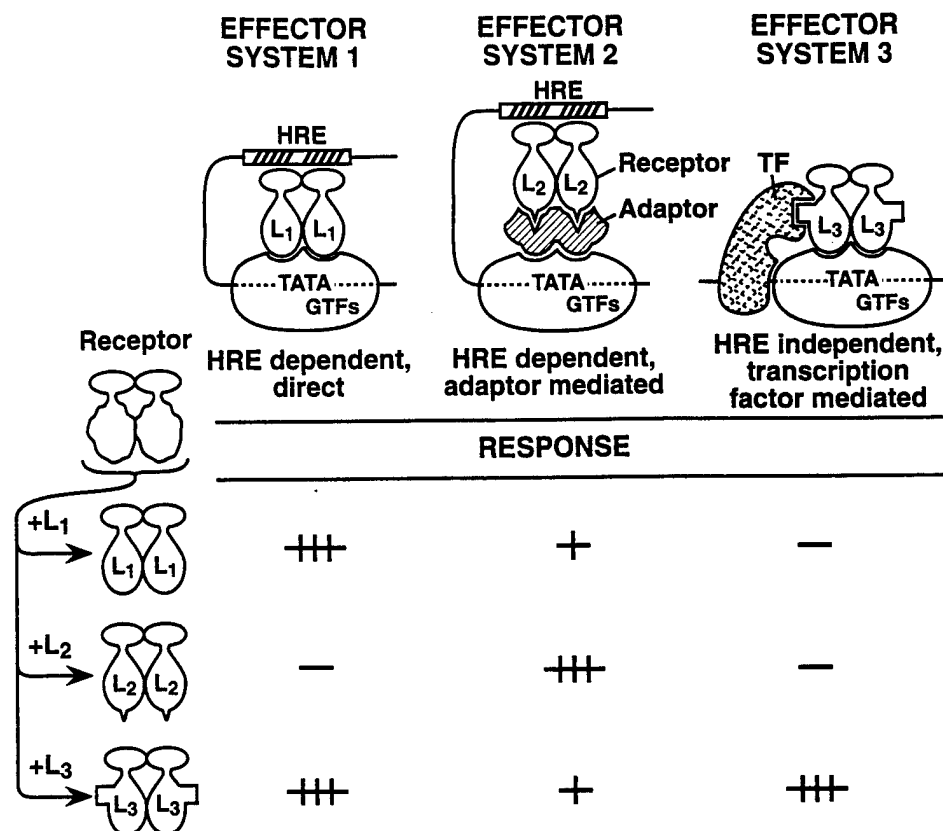


Fig. 2. Tripartite (Ligand-Receptor-Effector) Pharmacology

The response to a hormone is mediated by a tripartite interaction involving the ligand, the receptor, and effector sites through which the ligand-receptor complex regulates the response. The *top* of this scheme illustrates three different modes for nuclear receptor activation of genes; for each mode, an optimal ligand-receptor-effector combination is shown. The *bottom* of the scheme illustrates the activity that each of the three ligand-receptor complexes might have at each of the three effector sites. Note that the receptor adopts a different conformation in its complex with the three ligands and that these different "shapes" affect the nature of the receptor-effector coupling. In a tripartite scheme, the potency of a ligand is determined largely by its affinity of interaction with the receptor, but its biocharacter is determined by the interaction that the ligand-receptor complex has with various effector sites. Therefore, for each receptor, the biocharacter (and to some degree the potency) of a hormone cannot be uniquely assigned without reference to a specific response and effector interaction. Other modes of nuclear receptor gene activation than the three illustrated here, such as the remodeling of nucleosomal and chromatin architecture by hormone receptor complexes, have been identified. However, for simplicity, only three are shown here as examples.

unusual about the tripartite nature of the nuclear hormone receptor system is that there appears to be an unusual number and variety of effectors; this might well be the hallmark of pleiotropic response systems.

The pharmacological implications of the tripartite model are significantly different from the bipartite model. In the bipartite model (Fig. 1 and Table 2), a single interaction, the binding of ligand by receptor, directly regulates receptor function and thereby determines both the potency and the biocharacter of the ligand. By contrast, ligand potency and ligand biocharacter are determined through two different interactions in the tripartite scheme (Fig. 2 and Table 2). In the first interaction, ligand binds to receptor to form a complex, and the affinity of this *binding* is a principal determinant of ligand potency. However, this ligand-

receptor interaction alone does not control the response and therefore is not a direct determinant of ligand biocharacter. The pharmacological nature of the ligand, its biocharacter and its potency, is only fully established through the second interaction. This *coupling*, which occurs between the ligand-receptor complex and the third partner, the effector, is an interaction that has both an affinity and an efficacy dimension.

The Nature of Effectors for Nuclear Receptors

In the nuclear hormone receptor systems, the effector site represents the aggregate of all the other components with which the ligand-receptor complex interacts at each regulated gene. Thus, the effector is obviously complex. It is made up of elements common

to all genes, as well as elements unique to each cell and to each gene, even in systems like the estrogen receptor where only a single receptor exists. The nuclear components that define effector-site selectivity are not well understood at present. Nevertheless, they may be grouped into several classes, three of which are illustrated in Fig. 2.

In most cases, the coupling between the receptor and effector involves direct interaction with DNA through hormone response elements, which at various genes may be consensus, nonconsensus, single, multiple, half-sites, etc.; DNA sequences flanking the response elements, which are known to affect receptor binding affinity, also differ in various responsive genes. For the most part, sequences that bind receptors with high affinity act as tethering sites for these potent gene activators. In certain instances such as the proliferin gene, upstream binding to a specific sequence appears to favor a conformationally inactive form of the glucocorticoid receptor and may be operationally defined as a negative glucocorticoid response element (80).

After binding to their cognate response elements, a number of receptors appear to touch the general transcription factor complex (GTFs) located at the TATA box (*cf.* Fig. 2, effector system 1) (81–83). Although TFIID may be a target for certain receptors, the preferred partner of progesterone, estrogen, thyroid hormone, vitamin D₃ receptors, and COUP-TF often appears to be TFIIB, a rate-limiting component whose presence appears requisite for drawing RNA polymerase (and TFIIF) to the promoter (84). At this level, both positive and negative associations have been predicted for receptors with TFIIB. For example, unoccupied thyroid hormone receptor touches TFIIB at two distinct regions; one of these interactions has been hypothesized to be repressive, to explain the well described silencing activity of ligand-free thyroid hormone receptor at certain genes (82). Thyroid hormone binding to thyroid hormone receptor inhibits this repressive interaction. Nevertheless, effector site interactions appear to be of even greater complexity.

Experimental evidence has predicted the existence of adaptor proteins that may act as either coactivators (85, 86) or corepressors for nuclear receptors (*cf.* Fig. 2, effector system 2). In transfected cells, the ability of activated estrogen receptor to suppress or "squench" the transcriptional capacity of activated progesterone receptor has been interpreted to result from their competitive interactions with limiting concentrations of a putative cellular coactivator (87–89). Recently, this hypothesis has been substantiated by the identification and cloning of a general steroid receptor coactivator (SRC-1), which fulfills many of the criteria that have been preassigned to such a molecule, *i.e.* it enhances ligand-induced gene activity (up to 10-fold) without altering basal transcription levels, and it can reverse interreceptor squenching when transfected into a cell with two active receptors (90). SRC-1 appears to exist in two isoforms and its mRNA is present in all cells. It

specifically interacts with the C-terminal activation domain (AF-2) of receptors in a ligand-dependent manner but functions with all steroid/thyroid/retinoic acid receptors tested to date. This coactivator is inactive with receptors bound to pure antagonists but has been shown recently to enhance mixed agonist/antagonist activation of ER as well as ligand-independent activation of receptor by dopaminergic agonists and growth factors. Other potential adaptor proteins that interact with steroid receptors in a ligand-regulated manner, termed receptor-associated proteins (RAPs) or receptor-interacting proteins (RIPS), have been identified, although none have been proven yet to function as transcriptional coactivators. Cells with an abundance of coactivator should have a more pronounced response to a limiting concentration of receptor. It is clear that receptor-coactivator interactions are an important part of the tripartite response system at the gene level and can play a major role in quantitative aspects of cell response.

Elucidation of the molecular interactions of SRC-1 and other coactivators with receptor should advance our understanding of the mechanism of antagonist action. Previous experimental evidence has indicated that agonist- and antagonist-bound receptors exist in distinct conformations (91, 92). Interestingly, agonist-bound receptor binds efficiently to coactivator *in vitro* and *in vivo*, but the antagonist-bound receptor does not bind coactivator. Such differential interactions are illustrated by the varying activities postulated for the different ligand receptor complexes with effector system 2 (Fig. 2, *bottom*) and suggest that antagonist action has its origin at two levels, that of ligand-induced receptor conformation and that of receptor-effector interaction at the genetic level (see below). In such a scheme, antagonist-bound receptor occupies available hormone response elements in the cell, but its conformation does not allow productive interactions with coactivators or the general transcription factor apparatus at the core promoter (TATA box).

Recent data suggesting the existence of a corepressor(s) for the thyroid hormone receptor (and retinoic acid receptor) add an additional twist (93, 94). Unoccupied nuclear thyroid hormone receptor has been reported to silence target gene activity (95, 96). Presentation of thyroid hormone (T₃) reverses silencing and produces a stimulation of transcription. It has been proposed, using reverse squenching experiments to relieve silencing, that a soluble corepressor in target cells binds to unoccupied but not ligand-bound receptor, thus aiding in the thyroid hormone receptor-induced repression of basal transcription at select target genes (93). Recently, two "corepressor" molecules appear to have been cloned in their entirety and seem to fulfill the expected criteria, *i.e.* selective silencing, which is dependent on unoccupied thyroid hormone receptor or retinoic acid receptor (97, 98). In fact, it is likely that multiple coactivators and corepressors will be shown to be operative in cells. More than one agonist-dependent receptor interactive protein has

been reported already (99–105). Although the full consequences of such interactions are not clear at present, an ever increasing level of complexity is evolving at the effector stage of hormone response.

Perhaps the most influential aspect of promoter context or gene-specific response to a ligand is the array of other transcription factors present at a given gene. Although there is evidence for certain promoter-specific factors, the bulk of interactive regulation appears to occur upstream of the transcription start site at multiple enhancers. It is well known that two receptor dimers bound to the 5'-flanking sequence of a target gene can result in transcriptional synergy (106). This also applies to mixes of receptors and other compatible DNA-bound transcription factors, since a number of synergistic (and antagonistic) interactions have been reported among steroid receptors and unrelated transcription factors (72, 73, 73a, 107). Not surprisingly, the mix of receptors with certain transacting factors located at critical positions upstream of the promoter also may result in transcriptional interference.

A number of laboratories have suggested that interactive regulation between transcription factors can occur in cells even in the absence of DNA binding. For example, transcription factor AP-1 can promote active (or positive) influences on receptors independent of their DNA binding. Interactions in the nucleoplasm may occur or AP-1 (fos/jun) may bind to its regulatory element at a gene and serve as a docking site for a steroid receptor via protein-protein interaction (108) (cf. Fig. 2, effector system 3). Likewise, in some target genes with unusual estrogen-inducible enhancers, such as *c-myc* (109), creatine kinase (110), cathepsin D (111), and the protooncogene *c-jun* (112), receptor association with other known (such as transcription factor Sp1) or as yet unidentified DNA-binding proteins appears to facilitate receptor interaction with the enhancer. Receptor-mediated gene repression also may occur via protein-protein interactions among transcription factors. For example, glucocorticoid receptor down-regulation of certain genes regulated by the transcription factors AP-1 or NF κ B may occur via interactions between such regulators and the glucocorticoid receptor in the absence of DNA binding (113). Finally, nuclear proteins may interact directly and specifically with receptor molecules to antagonize their binding to DNA. Examples of such proteins are calreticulin, which antagonizes steroid receptors (114), and thyroid hormone receptor uncoupling protein (TRUP), which antagonizes thyroid hormone receptor and retinoic acid receptor (115).

Finally, it is worth noting that chromatin structure of genes in their native context provides a significant barrier for receptor to overcome in transcriptional regulation (40, 72, 116, 117). Nucleosomal repression of gene activation must be reversed by receptors, and selected nuclear helper proteins (e.g. SWI, SWE, SNF, Sin, etc.) may play important roles in the chromatin remodeling that appears to coincide with induction of

transcription. In any event, it is certain that a diverse spectrum of interactions can occur at an effector site and that this complexity may represent a mechanism whereby promoter context and cell specificity of response can be generated.

Pharmacology in Tripartite (Ligand-Receptor-Effector) Systems

In Fig. 2, we have laid out three tripartite schemes to illustrate the types of molecular interactions that may be operating in the activation of gene transcription by nuclear hormone receptors. Through this figure, we also have attempted to represent the combinatorial complexity that can arise as a result of the second interaction, between the ligand receptor complex and the effector. The interactions at the *top* of Fig. 2 illustrate the optimal interaction that might occur between three distinct effector systems and three different ligand receptor complexes, each formed from the same receptor with three different types of ligands; shown is the fact that each ligand-receptor complex has a distinct conformation. At the *bottom* of Fig. 2, we attempt to show the consequence—in terms of signal transduction—of the distinct interaction that each of these ligand-receptor complexes might have with all three of the effector systems. While this illustration is obviously limited and simplified (see previous section “The Nature of Effectors for Nuclear Receptors” and see below), it is meant to capture the conceptual basis of pharmacology in a tripartite receptor system, especially the fact that response diversity can be generated at the level of the effector. In addition to the three scenarios shown in Fig. 2, diversity can also be generated further by differences in the nature of the hormone response element, the influence of neighboring DNA-binding sites for other nuclear factors, as well as the nature of the promoter and chromatin state/conformation.

The transcription activation functions ascribed to different regions of nuclear hormone receptors (AF-1 and 2, or τ 1– τ 4) can be thought of as sites through which the receptor has the potential for interaction with these various effectors (70, 72, 73). However, the degree to which a particular ligand may engender the receptor to operate through these different activation function sites depends on the nature of the specific effector system with which the ligand-receptor complex interacts. Again, this is dependent on the cell- and promoter-specific factors and the response elements that constitute the effector. In cotransfection systems, mutant receptors can be used to amplify the varied effects of different ligands in their interaction with specific effector sites (5, 41, 43, 118–123). This approach has assisted in the identification of ligands with specific desired biocharacter, such as ligands for the estrogen receptor that have the proper spectrum of agonist/antagonist activity needed for hormone replacement therapy (43).

In tripartite receptor pharmacology, it is useful to consider that the *potency* of a particular ligand is determined principally through the first interaction (*ligand and receptor binding*), whereas its *biocharacter* (i.e. agonist-antagonist balance) is determined principally through the second interaction (*receptor-effector coupling*). This may prove to be an oversimplification, as there are known exceptions. In model transcription systems in yeast, certain receptor-modulatory proteins (SSN6-TUP1) have been shown to alter ligand potency (EC_{50}) of both estrogens and progestins by several orders of magnitude, not by a perturbation of ligand receptor binding, but by alteration of receptor-effector coupling that is interpreted as a modification of AF-1 activity. In this system, these adaptor proteins also alter the biocharacter of antiestrogens without changing ligand affinity (89). Related studies have defined a glucocorticoid modulatory element in the tyrosine amino transferase gene, and associated transactivating factors, that alter ligand potency and biocharacter (123). Conversely, it is possible that variations in response element sequence that affect receptor-effector coupling might also alter the conformation of the receptor in a manner that would change ligand affinity. Further investigation of ligand-receptor binding and receptor-effector coupling in carefully controlled systems will be required to fully elucidate the relative role that each interaction plays in establishing pharmacological potency and biocharacter. Regardless of these details, however, *in a tripartite receptor system, the pharmacological parameters of potency and biocharacter are not unique characteristics of a ligand; they can be assigned to a ligand only when reference is made to a specific response or its associated effector* (Table 2).

CONSEQUENCES AND EXPECTATIONS

A prerequisite for receptor pharmacology, be it bipartite or tripartite, is that ligand binding effects some conformational change in the receptor that initiates the response (directly—bipartite) or the potential for response (through coupling with effectors—tripartite). It is clear that the binding of a hormone ligand by its nuclear receptor results in significant conformational changes in the receptor. This has been evident for some time through indirect studies that have noted alterations in thermal stability, antibody binding, heat shock protein dissociation, hydrophobicity, DNA binding, and protease sensitivity upon ligand binding. More recently, crystallographic evidence (124–126) has shown that the small nuclear receptor ligands are almost completely surrounded by protein in their complexes with receptor. Moreover, within this complex there appear to be intimate and detailed contacts between protein and ligand over the whole ligand surface so that, of necessity, the conformation of a steroid-nuclear receptor complex must reflect the shape and structure of its ligand. Thus, the affinity and efficacy

with which these conformationally diverse ligand-receptor complexes interact with the various effector sites involved in the transcriptional regulation of different genes reflect the structure of the receptor complex *in its specific ligand-induced conformation*. What are the implications of this ligand-determined conformation of the nuclear hormone receptors?

First, it is not surprising that in the nuclear hormone receptor system, ligands of different structure operating through the same receptor can show distinct cell- and gene-specific effects. One should expect that the same receptor, bound with ligands of different structure, gives rise to complexes of different conformation. Such conformationally different ligand-receptor complexes have the potential for different coupling with the spectrum of effector sites that are present in each target cell and that embody all the cell- and gene-specific factors that enable individual genes to be differentially regulated by different ligands. At the moment, the number of genes whose expression is known to be regulated as a primary response to steroid hormones is rather limited. As more are identified, it is likely that the diversity of response to ligands that is possible with this tripartite receptor system will become even more evident.

Second, in contrast to allosteric effector ligands in enzyme systems and ion channels that bind rapidly to preformed regulatory sites and act like switches controlling the conformation between two states, active and inactive (conformation selection) (127), one should expect the hormonal ligand to affect the conformation of the receptor in more of a progressive or continuum fashion. The rate at which ligands associate with nuclear receptors is slow, far below diffusion control, which characterizes most small molecule-protein interactions. This suggested that the receptor undergoes a substantial conformational reorganization upon binding the ligand. Furthermore, since many unliganded receptors are associated with certain heat shock proteins, the sequences that constitute the ligand-binding pocket were thought to be somewhat disordered in the absence of ligand. Both of these expectations have been confirmed by recent X-ray crystal structures (124–126). Thus, the formation of the ligand-receptor complex in the nuclear hormone receptor system is an excellent example of an induced fit (128), conformation induction (127), or macromolecular perturbation (129), with the receptor conforming to the shape of the ligand (and the ligand, if flexible, having its conformation altered by binding to the receptor as well) (125, 126).

Finally, while structural elucidation methods will soon give us high resolution models for many nuclear receptors binding ligands of varying structure, the impact of this structural information on our understanding of steroid hormone molecular pharmacology, though very useful, will still be limited. The picture will be complete only when the details governing the coupling of these ligand-receptor complexes with the

varying elements of their third partners, the effector sites, also become illuminated.

Acknowledgments

Received October 10, 1995. Revision received and accepted November 28, 1995.

Address requests for reprints to: John A. Katzenellenbogen, 461 Roger Adams Lab 37-5, Department of Chemistry, University of Illinois, 600 South Mathews Ave, Urbana Illinois 61801.

This work received research support from the NIH, the U.S. Army Breast Cancer Program, and the Department of Energy.

REFERENCES

- Russell DW, Wilson JD 1994 Steroid 5 α -reductase: two genes/two enzymes. *Annu Rev Biochem* 63:25-61
- Funder JW 1993 Mineralocorticoids, glucocorticoids, receptors and response elements. *Science* 259:1132-1133
- Orti E, Bodwell JE, Munck A 1992 Phosphorylation of steroid hormone receptors. *Endocr Rev* 13:105-128
- Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS 1994 Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem* 269:4458-4466
- Katzenellenbogen BS, Montano M, Le Goff P, Schodin DJ, Kraus WL, Bhardwaj B, Fujimoto N 1995 Antiestrogens: mechanisms and actions in target cells. *J Steroid Biochem Mol Biol* 53:387-393
- Pfahl M 1994 Vertebrate receptors: molecular biology, dimerization and response elements. *Semin Cell Biol* 5:95-105
- Chambon P 1994 The retinoid signalling pathway: molecular and genetic analyses. *Semin Cell Biol* 5:115-125
- Tora L, Gronemeyer H, Turcotte B, Gaub M-, Chambon P 1988 The N-terminal region of the chicken progesterone receptor specifies target gene activation. *Nature* 333:185-188
- Tung LT, Mohamed MK, Hoeffler JP, Takimoto GS, Horwitz KB 1993 Antagonist-occupied human progesterone B-receptors activate transcription without binding to progesterone response elements and are dominantly inhibited by A-receptors. *Mol Endocrinol* 7:1256-1265
- Vegeto E, Shahbaz MM, Wen DX, Goldman ME, O'Malley BW, McDonnell DP 1993 Human progesterone receptor A form is a cell- and promoter-specific repressor of human progesterone receptor B function. *Mol Endocrinol* 7:1244-1255
- McDonnell DP 1995 Unraveling the human progesterone receptor signal transduction pathway. *Trends Endocrinol Metab* 6:133-138
- Wilson CM, McPhaul MJ 1994 A and B forms of the androgen receptor are present in human genital skin fibroblasts. *Proc Natl Acad Sci USA* 91:1234-1238
- Oakley RH, Madhabananda S, Cidlowski JA, The isoform of the human glucocorticoid receptor: expression, biochemical properties and putative function. Program of the 77th Annual meeting of The Endocrine Society, Washington D.C., 1995 (Abstract 81)
- McGuire WL, Chamness GC, Fuqua SAW 1991 Estrogen receptor variants in clinical breast cancer. *Mol Endocrinol* 5:1571-1577
- Fuqua SAW, Wolf DM 1995 Molecular aspects of estrogen receptor variants in breast cancer. *Breast Cancer Res Treat* 35:233-241
- Miksicek RJ, Lei Y, Wang Y 1993 Exon skipping gives rise to alternatively spliced forms of the estrogen receptor in breast tumor cells. *Breast Cancer Res Treat* 26:163-174
- Friend KE, Ang LW, Shupnik MA 1995 Estrogen regulates the expression of several novel estrogen receptor mRNA isoforms in rat pituitary. *Proc Natl Acad Sci USA* 92:4367-4371
- Skipper JK, Young LJ, Bergeron JM, Tetzlaff MT, Osborn CT, Crews D 1993 Identification of an isoform of the estrogen receptor messenger RNA lacking exon four and present in the brain. *Proc Natl Acad Sci USA* 90:7172-7175
- Toney TW, Katzenellenbogen BS 1986 Antiestrogen action in the medial basal hypothalamus and pituitary of immature female rats: insights concerning relationships among estrogen, dopamine and prolactin. *Endocrinology* 119:2661-2669
- Hayes JR, Rorke EA, Robertson DW, Katzenellenbogen BS, Katzenellenbogen JA 1981 Biological potency and uterine estrogen receptor interactions of the metabolites of the antiestrogens CI628 and U23,469. *Endocrinology* 108:164-172
- Robertson DW, Katzenellenbogen JA, Hayes JR, Katzenellenbogen BS 1982 Antiestrogen basicity-activity relationships: a comparison of the estrogen binding and antiuterotrophic potencies of several analogs of tamoxifen having altered basicity. *J Med Chem* 25:167-171
- Robertson DW, Katzenellenbogen JA, Long DJ, Rorke EA, Katzenellenbogen BS 1982 Tamoxifen antiestrogens. A comparison of the activity, pharmacokinetics and metabolic activation of the cis and trans isomers of tamoxifen. *J Steroid Biochem* 16:1-13
- Kneifel MA, Leytus SP, Fletcher E, Weber T, Mangel WF, Katzenellenbogen BS 1982 Uterine plasminogen activator activity: modulation by steroid hormones. *Endocrinology* 111:493-499
- Kneifel MA, Katzenellenbogen BS 1981 Comparative effects of estrogen and antiestrogen on plasma renin substrate levels and hepatic estrogen receptors in the rat. *Endocrinology* 108:545-552
- Evans G, Bryant HU, Sato M, Turner RT 1993 Raloxifene is a tissue specific estrogen agonist. *J Bone Miner Res* 8 [Suppl 1]:S134
- Black LJ, Sato M, Rowley ER, Magee DE, Bekele A, Williams DC, Cullinan GJ, Bendele R, Kauffman RF, Bensch WR, Frolik CA, Termine JD, Bryant HU 1994 Raloxifene (LY1394881 HCl) prevents bone loss and reduces serum cholesterol without causing uterine hypertrophy in ovariectomized rats. *J Clin Invest* 93:63-69
- Sato M, Glasebrook AL, Bryant HU 1994 Raloxifene: a selective estrogen receptor modulator. *J Bone Miner Met* 12 [Suppl 2]:S9-S20
- Draper MW, Flowers DE, Hustler WJ, Nield JA 1993 Effects of raloxifene (LY139481 HCl) on biochemical markers of bone and lipid metabolism in healthy postmenopausal women. In: Christiansen C, Riis B (eds) Proceedings of the 4th International Symposium on Osteoporosis and Consensus Development Conference. Handelstrykkeriet Aalborg APS, Aalborg, Denmark, pp 119-121
- Termine JD 1994 Therapy for postmenopausal osteoporosis based on fundamental disease etiology and pathophysiology. *J Bone Miner Met* 12 [Suppl 2]:S25-S28
- Fuchs-Young R, Glasebrook AL, Short LL, Draper MW, Rippy MK, Cole HW, Magee DE, Termine JD, Bryant HU 1995 Raloxifene is a tissue-selective agonist/antagonist that functions through the estrogen receptor. *Ann NY Acad Sci* 761:355-360
- Love RR, Mazess RB, Barden HS, Epstein SE, Newcomb PA, Jordan VC, Carbone PP, DeMets DL 1992

- Effects of tamoxifen on bone mineral density in postmenopausal women with breast cancer. *N Engl J Med* 326:852-856
32. Love RR, Newcomb PA, Wiebe DA, Surazicz TS, Jordan VC, Carone PP, DeMets DL 1990 Effects of tamoxifen on bone mineral density in postmenopausal women with breast cancer. *J Natl Cancer Inst* 82:1327-1332
 33. Wolf DM, Jordan VC 1992 Gynecologic complications associated with long-term adjuvant tamoxifen therapy for breast cancer. *Gynecol Oncol* 45:118-128
 34. Jordan VC 1995 Tamoxifen and tumorigenicity: a predictable concern. *J Natl Cancer Inst* 87:623-626
 35. Jordan VC, Assikis VJ 1995 Endometrial carcinoma and tamoxifen: clearing up a controversy. *Clin Cancer Res* 1:467-472
 36. Early Breast Cancer Trialists' Collaborative Group 1992 Systemic treatment of early breast cancer by hormonal, cytotoxic or immune therapy. *Lancet* 339:1-15
 37. Wakeling AE, Bowler J 1988 Biology and mode of action of pure antiestrogens. *J Steroid Biochem* 30:141-147
 38. Wakeling AE, Dukes M, Bowler J 1991 A potent specific pure antiestrogen with clinical potential. *Cancer Res* 51:3867-3873
 39. Weaver CA, Springer PA, Katzenellenbogen BS 1988 Regulation of pS2 gene expression by affinity labeling and reversibly binding estrogens and antiestrogens: comparison of effects on the native gene and on pS2-chloramphenicol acetyltransferase fusion genes transfected into MCF-7 human breast cancer cells. *Mol Endocrinol* 2:936-945
 40. Mymryk JS, Archer TK 1995 Dissection of progesterone receptor-mediated chromatin remodeling and transcriptional activation *in vivo*. *Genes Dev* 9:1366-1376
 41. Montano MM, Müller V, Trobaugh A, Katzenellenbogen BS 1995 The carboxy-terminal F domain of the human estrogen receptor: role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 9:814-825
 42. Tzukerman MT, Esty A, Santiso-Mere D, Danielian P, Parker MG, Stein RB, Pike JW, McDonnell DP 1994 Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol Endocrinol* 8:21-30
 43. McDonnell DP, Clemm DL, Hermann T, Goldman ME, Pike JW 1995 Analysis of estrogen receptor function *in vitro* reveals three distinct classes of antiestrogens. *Mol Endocrinol* 9:659-669
 44. McDonnell DP, Goldman ME 1994 RU486 exerts antiestrogenic activities through a novel progesterone receptor A form-mediated mechanism. *J Biol Chem* 269:11945-11949
 45. Kraus WL, Weis KE, Katzenellenbogen BS 1995 Inhibitory cross-talk between steroid hormone receptors: differential targeting of estrogen receptor in the repression of its transcriptional activity by agonist- and antagonist-occupied progestin receptors. *Mol Cell Biol* 15:1847-1857
 46. Berry M, Metzger D, Chambon P 1990 Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-2818
 47. Katzenellenbogen BS, Norman MJ, Eckert RL, Peltz SW, Mangel SF 1984 Bioactivities, estrogen receptor interactions and plasminogen activator inducing activities of tamoxifen and hydroxytamoxifen isomers in MCF-7 human breast cancer cells. *Cancer Res* 44:112-119
 48. Wei LL, Mangel WF, Katzenellenbogen BS 1985 Biological activities of tamoxifen aziridine, an antiestrogen-based affinity label for the estrogen receptor *in vivo* and *in vitro*. *J Steroid Biochem* 23:875-881
 49. Eckert RL, Katzenellenbogen BS 1982 Effects of estrogens and antiestrogens on estrogen receptor dynamics and the induction of progesterone receptor in MCF-7 human breast cancer cells. *Cancer Res* 42:139-144
 50. Fujimoto N, Katzenellenbogen BS 1994 Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen-selectivity and promoter-dependence. *Mol Endocrinol* 8:296-304
 51. Shull JD, Beams FE, Baldwin TM, Gilchrist CA, Hrbek MJ 1992 The estrogenic and antiestrogenic properties of tamoxifen in GH₄C₁ pituitary tumor cells are gene specific. *Mol Endocrinol* 6:529-535
 52. Ramkumar T, Adler S 1995 Differential positive and negative transcriptional regulation by tamoxifen. *Endocrinology* 136:536-542
 53. Yang NN, Hardikar S, Kim J 1993 Raloxifene, an antiestrogen, simulates the effects of estrogen on inhibiting bone resorption through regulating TGF β -3 expression in bone. *J Bone Miner Res* 8[Suppl 1]:S118
 54. Yang NN, Hardikar S, Venugopalan M, Estrogen receptor regulates the transforming growth factor β 3 (TGF- β 3) gene expression through a novel mechanism. Program of the 77th Annual Meeting of The Endocrine Society, Washington, D.C., 1995, (Abstract 235)
 55. Ehrlich P 1897 Die Wertbemessung des diphtherieheilserums und deren theoretische Grundlagen. *Klin Jahr* 6:299-326
 56. Langley JN 1905 On the reaction of cells and of nerve-endings to certain poisons, chiefly as regards the reaction of striated muscle to nicotine and to curari. *J Physiol (Lond)* 33:374-413
 57. Clark AJ 1926 The reaction between acetyl choline and muscle cells. *J Physiol (Lond)* 61:530-546
 58. Gaddum JH 1926 The action of adrenalin and ergotamine on the uterus of the rabbit. *J Physiol (Lond)* 61:141-150
 59. Ariens EJ, Simonis AM 1964 A molecular basis for drug action. *J Pharm Pharmacol* 16:137-157
 60. Stephenson RP 1956 A modification of receptor theory. *Br J Pharmacol Chemother* 11:379-393
 61. Jensen EV, Jacobson HI 1962 Basic guides to the mechanism of estrogen action. *Recent Prog Horm Res* 18:387-414
 62. Mueller GC, Herranen AM, Jervell KF 1958 Studies on the mechanism of action of estrogens. *Recent Prog Horm Res* 14:95-129
 63. Mueller GC, Gorski J, Aizawa Y 1961 The role of protein synthesis in early estrogen action. *Proc Natl Acad Sci USA* 47:164-169
 64. O'Malley BW, McGuire WL, Kohler PO, Koreman SG 1969 Studies on the mechanism of steroid hormone regulation of synthesis of specific proteins. *Recent Prog Horm Res* 25:105-160
 65. O'Malley BW, Means AR 1974 Female steroid hormones and target cell nuclei. *Science* 183:610-620
 66. Yamamoto KR, Alberts BM 1976 Steroid receptors: elements for modulation of eukaryotic transcription. *Annu Rev Biochem* 45:721-746
 67. Katzenellenbogen BS, Gorski J 1975 Estrogen actions on syntheses of macromolecules in target cells. In: Litwack G (ed) *Biochemical Action of Hormones*. Academic Press, New York, vol 3:187-243
 68. Gorski J, Gannon F 1976 Current models of steroid hormone action: a critique. *Annu Rev Physiol* 38:425-450
 69. Evans RM 1988 The steroid and thyroid hormone receptor superfamily. *Science* 240:889-895
 70. Gronemeyer H 1991 Transcription activation by estrogen and progesterone receptors. *Annu Rev Genet* 25:89-123
 71. Katzenellenbogen JA 1992 Probes for steroid receptors—studies at the molecular, cellular, and whole organism levels. In: *Proceedings of The Robert A. Welch*

- Foundation Conference on Chemical Research: XXXV. Chemistry at the Frontiers of Medicine. Robert A. Welch Foundation, Houston, TX, pp 229-257
72. Truss M, Beato M 1993 Steroid hormone receptors: interaction with deoxyribonucleic acid and transcription factors. *Endocr Rev* 14:459-479
73. Tsai M-J, O'Malley BW 1994 Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 63:451-486
- 73a. Simons Jr SS, Oshima H, Szapary D 1992 Higher levels of control: modulation of steroid hormone-regulated gene transcription. *Mol Endocrinol* 6:995-1002
74. Ariens EJ, Rodrigues De Miranda JF 1979 The receptor concept: recent experimental and theoretical developments. In: Gualtiere F, Giannella M, Melchoirre C (eds) *Recent Advances in Receptor Chemistry*. Elsevier/North-Holland Biomedical Press, Amsterdam, pp 1-37
75. Bourne HR, Roberts JM 1987 Drug receptors and pharmacodynamics. In: Katzung BG (ed) *Basic and Clinical Pharmacology*. Appleton & Lange, Norwalk, CT, pp 9-22
76. Nogrady T 1988 Receptor-effector theories. In: Nogrady T (ed) *Medicinal Chemistry: A Biochemical Approach*, ed. 2. Oxford University Press, New York, pp 58-91
77. Ross EM 1990 Pharmacodynamics: mechanisms of drug action and the relationship between drug concentration and effect. In: Gilman AG, Rall TW, Nies AS, Taylor P (eds) *The Pharmacological Basis of Therapeutics*, ed. 8. Pergamon Press, New York, pp 33-48
78. Hulme EC (ed) 1990 *Receptor-Effector Coupling: A Practical Approach*. Oxford University Press, New York
79. Liu J, Farmer JD, Lane WS, Friedman J, Weissman I, Schreiber SL 1991 Calcineurin is a common target of cyclophilin-cyclosporin A and FKBP-FK506 complexes. *Cell* 66:807-815
80. Diamond MI, Miner JN, Yoshinaga SK, Yamamoto KR 1990 Transcription factor interactions: selectors of positive or negative regulation from a single DNA element. *Science* 249:1266-1272
81. Ing NH, Beekman JM, Tsai SY, Tsai M-J, O'Malley BW 1992 Members of the steroid hormone receptor superfamily interact with TFIIB (S300II). *J Biol Chem* 267:17617-17623
82. Baniahmad A, Ha I, Reinberg D, Tsai SY, Tsai M-J, O'Malley BW 1993 Interaction of human thyroid hormone receptor beta with transcription factor TFIIB may mediate target gene derepression and activation by thyroid hormone. *Proc Natl Acad Sci USA* 90:8832-8836
83. Blanco J, Wang I-M, Tsai SY, Tsai M-J, O'Malley BW, Jurutka PW, Haussler MR, Ozato K 1995 TFIIB and vitamin D receptor cooperatively activate ligand dependent transcription. *Proc Natl Acad Sci USA* 92:1535-1539
84. Zawel L, Reinberg D 1995 Common themes in assembly and function of eukaryotic transcription complexes. *Annu Rev Biochem* 64:533-561
85. Ptashne M, Gann AAF 1990 Activators and targets. *Nature* 346:329-331
86. Flanagan PM, Kelleher RJ, Sayre MH, Tschochner H, Kornberg RD 1991 A mediator required for activation of RNA polymerase II transcription *in vitro*. *Nature* 350:436-438
87. Meyer ME, Gronemeyer H, Turcotte B, Bocquel M, Tasset D, Chambon P 1989 Steroid hormone receptors compete for factors that mediate their enhancer function. *Cell* 57:433-442
88. Conneely OM, Kettelberger DM, Tsai M-J, O'Malley BW 1989 Promoter specific activating domains of the chicken progesterone receptor. In: Roy AK, Clark J (eds) *Gene Regulation by Steroid Hormones IV*. Springer-Verlag, New York, pp 220-223
89. McDonnell DP, Vegeto E, O'Malley BW 1992 Identification of a negative regulatory function for steroid receptors. *Proc Natl Acad Sci USA* 89:10563-10567
90. Onate SA, Tsai SY, Tsai M-J, O'Malley BW 1995 Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270:1354-1357
91. Allan GF, Leng XH, Tsai SY, Weigel NL, Edwards DP, Tsai M-J, O'Malley BW 1992 Hormone and antihormone induce distinct conformational changes which are central to steroid receptor activation. *J Biol Chem* 267:19513-19520
92. Beekman JM, Allan GF, Tsai SY, Tsai M-J, O'Malley BW 1993 Transcriptional activation by the estrogen receptor requires a conformational change in the ligand binding domain. *Mol Endocrinol* 7:1266-1274
93. Baniahmad A, Leng X, Burris TP, Tsai SY, Tsai M-J, O'Malley BW 1995 The α activation domain of the thyroid hormone receptor is required to release a corepressor(s) necessary for transcriptional silencing. *Mol Cell Biol* 15:76-86
94. Nawaz Z, Tsai M-J, O'Malley BW 1995 Specific mutations in the ligand binding domain selectively abolish the silencing function of human thyroid receptor β . *Proc Natl Acad Sci USA* 92:11691-11695
95. Damm K, Evans RM 1993 Identification of a domain required for oncogenic activity and transcriptional suppression by v-erbA and thyroid-hormone receptor alpha. *Proc Natl Acad Sci USA* 90:10668-10672
96. Baniahmad A, Steiner C, Kohne AC, Renkawitz R 1990 Modular structure of a chicken lysozyme silencer: involvement of an unusual thyroid hormone receptor binding site. *Cell* 61:505-514
97. Chen JD, Evans RM 1995 A transcriptional co-repressor that interacts with nuclear hormone receptors. *Nature* 377:454-457
98. Hörlein AJ, Näär AM, Heinzel T, Torchia J, Glass B, Kurokawa R, Ryan A, Kamel Y, Söderström M, Glass CK, Rosenfeld MG 1995 Ligand-independent repression by the thyroid hormone receptor mediated by a nuclear receptor co-repressor. *Nature* 377:397-404
99. Baniahmad C, Nawaz Z, Baniahmad A, Gleeson MAG, Tsai M-J, O'Malley BW 1995 Enhancement of human estrogen receptor activity by SPT6: a potential coactivator. *Mol Endocrinol* 9:34-43
100. Halachmi S, Marden E, Martin G, MacKay H, Abbondanza C, Brown M 1994 Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription. *Science* 264:1455-1458
101. Cavaillès V, Dauvois S, Danielian PS, Parker MG 1994 Interaction of proteins with transcriptionally active estrogen receptors. *Proc Natl Acad Sci USA* 91:10009-10013
102. Cavaillès V, Dauvois S, L'Horset F, Lopez G, Hoare S, Kushner PJ, Parker MG 1995 Nuclear factor RIP140 modulates transcriptional activation by the estrogen receptor. *EMBO J* 14:3741-3751
103. Landel CC, Kushner PJ, Greene GL 1994 The interaction of human estrogen receptor with DNA is modulated by receptor-associated proteins. *Mol Endocrinol* 8:1407-1419
104. Le Douarin B, Zechel C, Garnier J, Luyz Y, Tora L, Pierrat B, Heery D, Gronemeyer H, Chambon P, Losson R 1995 The N-terminal part of TIF1, a putative mediator of the ligand-dependent activation function (AF-2) of nuclear receptors, is fused to B-raf in the oncogenic protein T18. *EMBO J* 14:2020-2033
105. Lee JW, Ryan F, Swaffield JC, Johnston SA, Moore DD 1995 Interaction of thyroid-hormone receptor with a conserved transcriptional mediator. *Nature* 374:91-94
106. Tsai SY, Carlstedt-Duke J, Weigel NL, Dahlman K, Gustafsson J-A, Tsai M-J, O'Malley BW 1988 Molecular interactions of steroid hormone receptor with its en-

- hancer element: evidence for receptor dimer formation. *Cell* 55:361-369
107. Baniahmad C, Muller M, Altschmied J, Renkawitz R 1991 Co-operative binding of the glucocorticoid receptor DNA binding domain is one of at least two mechanisms for synergism. *J Mol Biol* 222:155-165
 108. Webb P, Lopez GN, Uht RM, Kushner PJ 1995 Tamoxifen activation of the estrogen receptor/AP-1 pathway: potential origin for the cell-specific estrogen-like effects of antiestrogens. *Mol Endocrinol* 9:443-456
 109. Dubik D, Shiu RP 1992 Mechanism of estrogen activation of c-myc oncogene expression. *Oncogene* 7:1587-1594
 110. Wu-Peng XS, Pugliese TE, Dickerman HW, Pentecost BT 1992 Delineation of sites mediating estrogen regulation of the rat creatine kinase B gene. *Mol Endocrinol* 6:231-240
 111. Krishnan V, Wang X, Safe S 1994 Estrogen receptor-Sp1 complexes mediate estrogen-induced cathepsin D gene expression in MCF-7 human breast cancer cells. *J Biol Chem* 269:15912-15917
 112. Hyder SM, Chiapetta C, Yokoyama K, Stancel GM 1995 The protooncogene c-jun contains an unusual estrogen-inducible enhancer within the coding sequence. *J Biol Chem* 270:8506-8513
 113. Yoshinaga SK, Peterson CL, Herskowitz I, Yamamoto KR 1992 Roles of SW1, SW2 and SW3 proteins for transcriptional enhancement by steroid receptors. *Science* 258:1598-1604
 114. Burns K, Duggan B, Atkinson EA, Famulski KS, Nemer M, Bleackley RC, Michalak M 1994 Modulation of gene expression by calreticulin binding to the glucocorticoid receptor. *Nature* 367:476-480
 115. Burris TP, Nawaz Z, Tsai M-J, O'Malley BW 1995 A nuclear hormone receptor-associated protein that inhibits transactivation by the thyroid hormone and retinoic acid receptors. *Proc Natl Acad Sci USA* 92:9525-9529
 116. Hager GL, Archer TK 1991 The interaction of steroid receptors with chromatin. In: Parker MG (ed) *Nuclear Hormone Receptors*. Academic Press, London, pp 217-234
 117. Miner JN, Yamamoto KR 1991 Regulatory crosstalk at composite response elements. *Trends Biochem Sci* 16:423-426
 118. Pakdel F, Katzenellenbogen BS 1992 Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J Biol Chem* 267:3429-3437
 119. Wrenn CK, Katzenellenbogen BS 1993 Structure-function analysis of the hormone binding domain of the human estrogen receptor by region-specific mutagenesis and phenotypic screening in yeast. *J Biol Chem* 268:24089-24098
 120. Katzenellenbogen BS, Bhardwaj B, Fang H, Ince BA, Pakdel F, Reese JC, Schodin DJ, Wrenn CK 1993 Hormone binding and transcription activation by estrogen receptors: analyses using mammalian and yeast systems. *J Steroid Biochem Mol Biol* 47:39-48
 121. Danielian PS, White R, Hoare SA, Fawell SF, Parker MG 1993 Identification of residues in the estrogen receptor that confer differential sensitivity to estrogen and hydroxytamoxifen. *Mol Endocrinol* 7:232-240
 122. Jiang S-Y, Langan-Fahey SM, Stella AL, McCague R, Jordan VC 1992 Point mutation of estrogen receptor (ER) in the ligand-binding domain changes the pharmacology of antiestrogens in ER-negative breast cancer cells stably expressing complementary DNA's for ER. *Mol Endocrinol* 6:2167-2174
 123. Lefstin JA, Thomas JR, Yamamoto KR 1994 Influence of a steroid receptor DNA-binding domain on transcriptional regulatory functions. *Genes Dev* 8:2842-2856
 - 123a. Oshima H, Simons Jr SS 1992 Modulation of transcription factor activity by a distant steroid modulatory element. *Mol Endocrinol* 6:416-428
 124. Bourguet W, Ruff M, Chambon P, Gronemeyer H, Moras D 1995 Crystal structure of the ligand-binding domain of the nuclear receptor RXR- α . *Nature* 375:377-382
 125. Renaud J-P, Rochel N, Ruff M, Vivat V, Chambon P, Gronemeyer H, Moras D 1995 Crystal structure of the RAR- γ ligand-binding domain bound to all-trans retinoic acid. *Nature* 378:681-689
 126. Wagner RL, Apriletti JW, McGrath ME, West BL, Baxter JD, Fletterick RJ 1995 A structural role for hormone in the thyroid hormone receptor. *Nature* 378:690-697
 127. Burgen ASV 1970 Receptor mechanisms. *Annu Rev Pharmacol* 10:7-15
 128. Koshland DE, Neet KE 1968 The catalytic and regulatory properties of enzymes. *Annu Rev Biochem* 37:359-410
 129. Belleau B 1964 A molecular theory of drug action based on induced conformational perturbation of receptors. *J Med Chem* 7:776-785



Nuclear hormone receptors: ligand-activated regulators of transcription and diverse cell responses

John A Katzenellenbogen¹ and Benita S Katzenellenbogen²

Signal transduction via nuclear hormone receptors is unusual in that the hormone ligand forms an integral part of the protein complex involved in DNA binding and transcriptional activation. New structural and biochemical results have begun to unravel how these receptors produce different effects in different cells, and the structural changes involved in transcriptional activation.

Addresses: ¹Department of Chemistry, and ²Departments of Molecular and Integrative Physiology and Cell and Structural Biology, University of Illinois, Urbana, IL 61801, USA.

Chemistry & Biology July 1996, 3:529-536

© Current Biology Ltd ISSN 1074-5521

Introduction

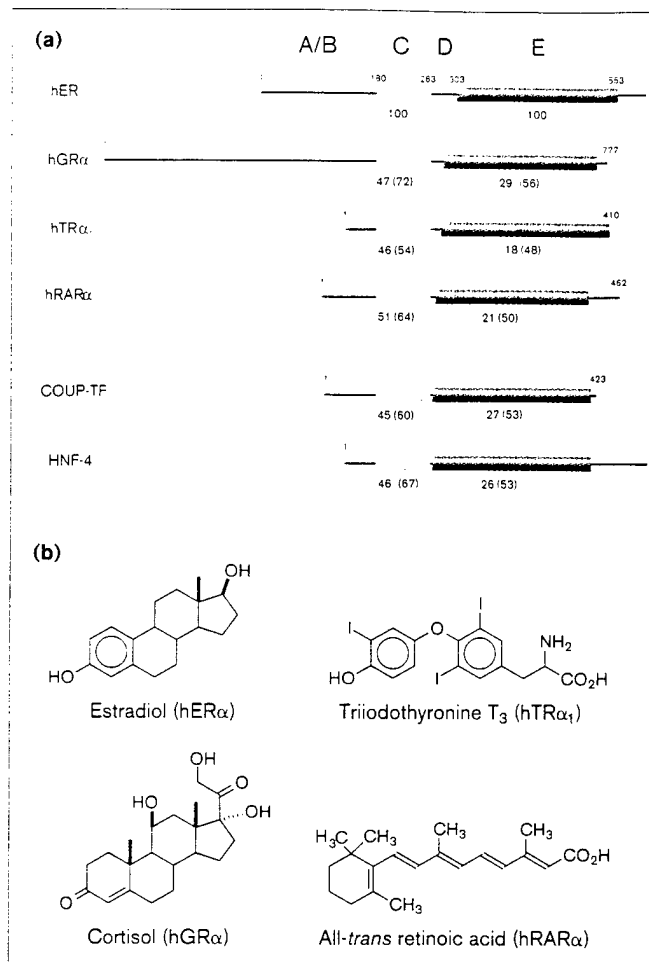
Radiolabeled steroid and thyroid hormones of high specific activity were first prepared in the late 1960's, and were used as probes to identify the sites of hormone action [1]. It has been known for nearly 30 years that these hormones act via intracellular receptor proteins whose principal target for action is in the nucleus. The receptor proteins were quickly surmised to be regulators of transcription [2-6], and are now known to be part of the nuclear receptor superfamily. This large group of transcription factors includes proteins that mediate the action of the steroid hormones (such as estrogens, androgens, glucocorticoids, mineralocorticoids and the insect steroid hormone ecdysone), as well as the non-steroid hormones (for example, thyroid hormone, vitamin D3 and the retinoids) and receptors that mediate the peroxisomal proliferation response to fatty acids and other factors (Fig. 1) [7-11].

Many other members of the superfamily have been identified by low stringency hybridization analysis; some of the genes thus identified encode proteins that are known to be expressed and have the conserved six-domain structure seen in the hormone receptors. Because the hormonal ligands for these proteins are unknown, they are termed 'orphan' receptors [12]. It is however possible that some of these so-called receptors may act as transcription factors alone, without ligands. To add to the complexity of the situation, most classes of receptors within this family contain more than one subtype (i.e., products of closely related genes); sometimes there are also different isoforms (i.e., products from alternate transcription start sites on the same gene) and products of mRNA splice variants. Both the concentration of these receptors and the relative ratio of subtypes and isoforms vary in different target tissues and at different stages of development.

Structure and function of the nuclear receptors

The signature of the nuclear receptor family is a six-domain structure, the most highly conserved portion of which is the small (~70-80 amino acids) domain, C, that is responsible for DNA binding (Fig. 2). This domain has been known for some time to have a helix-loop-helix structure containing two zinc atoms, each chelated by four cysteine thiols at the start of each helix. Three residues at the start of the first helix in this domain 'read' a five to six base pair code in a DNA hormone-response element; the mechanism of this sequence-specific recognition is becoming increasingly clear through structural analysis of domain C-oligonucleotide complexes by X-ray crystallography [13]. The large (~250 amino acid) domain, E, which

Figure 1



Structures of nuclear receptors and their ligands. (a) Common domain structure of representative members of the nuclear receptor superfamily, human estrogen receptor α (hER α), human glucocorticoid receptor α (hGR α), human thyroid hormone receptor (hTR α), human retinoic acid receptor γ (hRAR γ), and two orphan receptors COUP-TF and HNF-4. The DNA-binding domain C and ligand-binding domain E are shown with their percent sequence identity (or similarity, in parentheses) to hER α . (b) The natural ligands for the first four receptors in (a) are shown; there are no known ligands for the orphan receptors COUP-TF and HNF-4.

is moderately conserved across members of the family, is responsible for hormone binding and dimerization, and is critical in the regulation of transcription (see below). The other domains (the amino-terminal A/B domains, the hinge domain D, and sometimes a carboxy-terminal domain, F), which are poorly conserved in length and sequence across the family, are mostly involved in the modulation of receptor function.

Nuclear receptor ligands are directly involved in transcriptional regulation

Recent advances have clarified the various ways in which these nuclear receptors can become activated, as well as

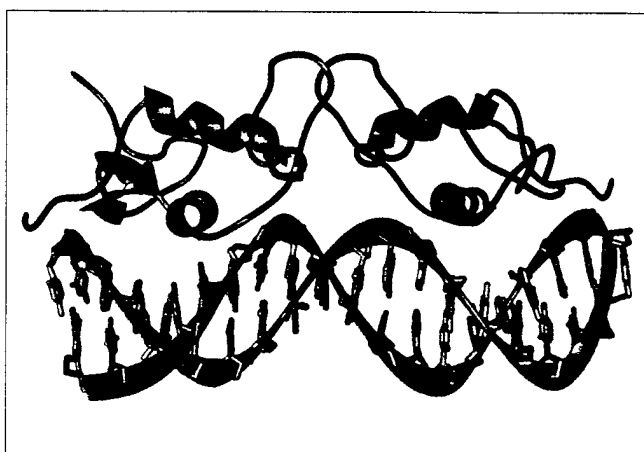
some of the molecular details of the modulation of the transcriptional activity of specific genes. The essential and intricate role of the ligand in controlling the regulation of gene transcription by these receptors is also now becoming clearer (Fig. 3) [14,15]. Although hormones and growth factors that interact with receptors at the cell membrane may ultimately affect gene transcription, they require multiple-step signal transduction pathways to do so (Fig. 3a); the change in transcription factor activity takes place far away from the interaction between the receptor and the provoking hormonal agent. By contrast, a ligand that activates a nuclear receptor forms a part of the multicomponent complex that directly regulates gene transcription. Such direct interactions offer interesting opportunities for selective pharmacology [16].

There is evidence that high affinity binders for steroid hormones exist in cell membranes, especially in some brain, pituitary and cancer cells. These receptors appear to mediate some very rapid effects of steroid hormones, and it is not yet clear whether their modes of action are similar to or different from the nuclear receptors [17,18]. We will focus here exclusively on the nuclear receptors, since the information on this class is most complete.

Variations on a theme

The classical picture of gene activation via nuclear receptors (Fig. 3b) is straightforward. The hormonal ligand binds to the receptor; the receptor-ligand complex thus formed binds (usually as a dimer) to a hormone-response element in the promoter region of a regulated gene, and the transcription of the gene connected to the promoter is thus activated.

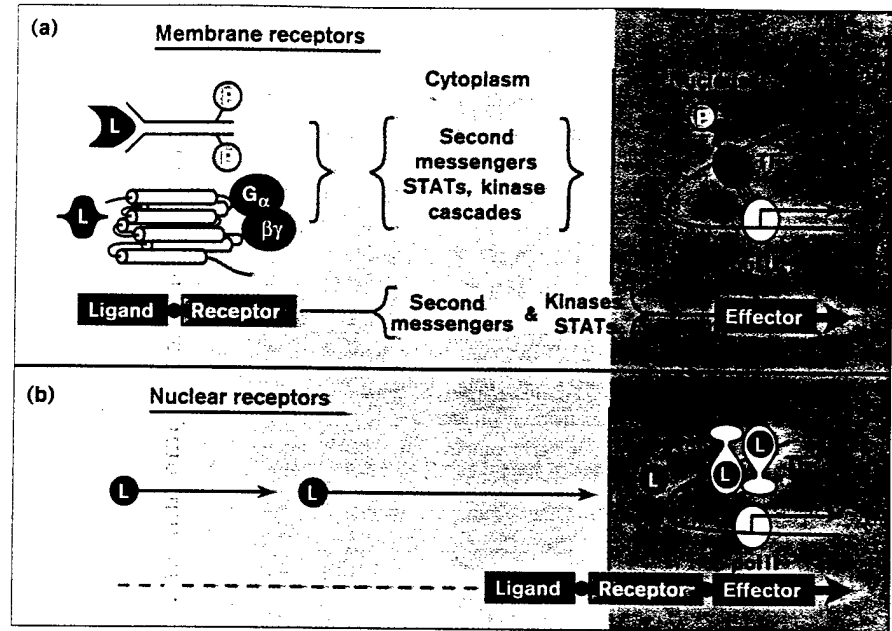
Figure 2



A ribbon structure representation of the human glucocorticoid receptor DNA-binding domain dimer complexed with a glucocorticoid response element (GRE). The DNA contact helices, shown edge on, interact with the palindromic DNA sequences of the GRE in adjacent major grooves.

Figure 3

Both membrane receptors and nuclear receptors modulate gene transcription, but nuclear receptors do so more directly. **(a)** Membrane receptor signaling; **(b)** nuclear receptor signaling. In a membrane receptor signaling system, the signal resulting from the binding of the ligand (L) to the receptor must be transduced to the nucleus via complex signal-transduction cascades, which typically involve second messengers, kinase cascades and/or phosphorylation (P) of intermediary proteins (such as STATs) in the cytoplasm. The end result is a change in the activity of a transcription factor (TF) in the nucleus, affecting the rate of initiation of RNA polymerase II (pol II). The effects of a hormone that acts via a nuclear receptor are much more direct; the ligand and receptor form part of the multicomponent complex that modulates pol II activity.



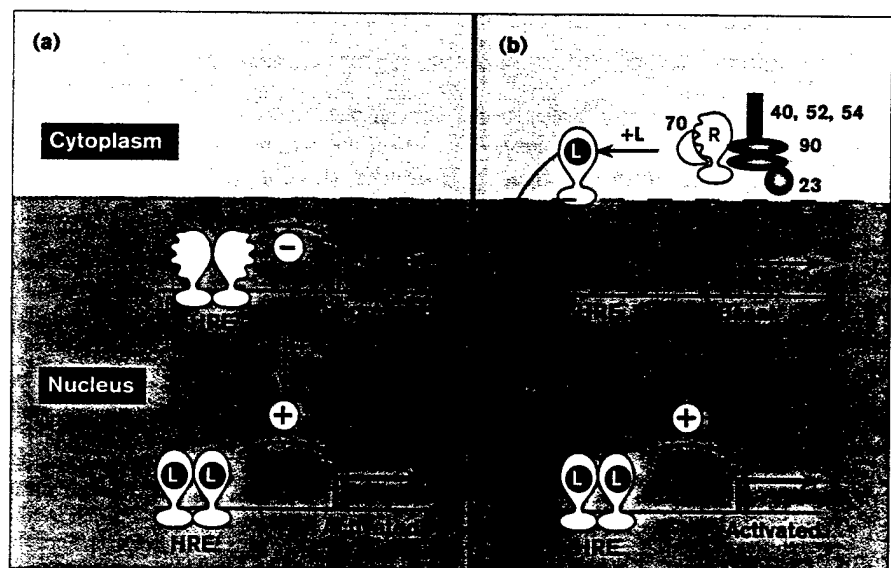
It cannot, however, be this simple. The target of the ligand-receptor complex can clearly vary with cell type, which would be impossible in the rudimentary scheme described above. For example, when estrogen binds to the estrogen receptor in breast cancer and uterine cells the result is the stimulation of transcription from some early response genes, such as *c-myc*, and genes for growth factors (such as TGF- α or pS2) or growth factor receptors (such as the EGF receptor) that are involved in the stimulation of cell proliferation evoked by the hormone [19]. The same

ligand-binding event in pituitary and liver cells results in activation of other genes. In the pituitary, the expression of various secreted proteins such as prolactin is increased, whereas in the liver the level of vitellogenin, among others, is increased.

The variations on the classical picture occur at all levels. One source of variability in the effect of ligand binding is the cellular distribution of the receptor in the absence of ligand. The receptors for certain non-steroid ligands (e.g.,

Figure 4

The subcellular location of unliganded nuclear receptors affects the way that they modulate transcription. **(a)** The unliganded receptors for nonsteroid ligands such as thyroid hormone and retinoic acid are typically bound as dimers to their hormone response elements (HREs), even in the absence of ligand, and can act as transcriptional repressors without ligands or transcriptional activators with ligand. **(b)** The unliganded receptors for some steroid hormones, such as glucocorticoids, are largely held as monomers in the cytoplasm by heat-shock proteins (90, 23), chaperonins (70) and immunophilins (40, 52, 54); in this state they have no effect on transcription. Ligand binding releases the receptors from the cytoplasmic aggregate, and the activated receptors bind as dimers to the HREs and activate transcription.



thyroid hormone and the retinoids) appear to be already bound to their response elements (Fig. 4) [20]. Ligand binding may strengthen DNA binding, and may alter the structure of the receptor so as to enhance transcription (see below). In the absence of ligand, these DNA-bound receptors repress gene transcriptional activity [21,22]. In contrast, many of the steroid nuclear receptors (e.g., the glucocorticoid receptor) are largely cytoplasmic in the absence of ligand. They are held in the cytoplasm in complex with heat-shock proteins, chaperonins, and various other proteins such as immunophilins [23]. Ligand binding helps the receptor to shed these proteins, move into the nucleus, dimerize, and interact with appropriate hormone response elements (Fig. 4). In such a scheme, the unliganded receptor cannot be used as a transcriptional repressor, as it is held in the cytoplasm, away from the DNA. The degree of nuclear versus cytoplasmic localization of unliganded receptors varies with different receptors and in different cells, so the effect of the unliganded receptor on transcription will depend on the cell and response in question.

A second level of variation in our originally simple scheme is the way in which the receptor forms a dimer. The non-steroid nuclear receptors for thyroid hormone, vitamin D and retinoic acid can either form homodimers or heterodimerize with the retinoid X receptor [12,20]. The receptor for the insect steroid hormone ecdysone, on the other hand, is active only as a heterodimer with the protein ultraspiracle, a homolog of the retinoid X receptor (RXR). The preference of the thyroid, vitamin D and retinoic acid receptors for pairing with themselves or with another partner depends on several factors, including the relative concentration of the monomer components (not forgetting the different subtypes and isoforms) and of their cognate ligands. Ligand binding can, in some situations, modulate the formation of specific complexes [24]. A further factor is the structure of the DNA response elements with which the homo- or heterodimers interact [20,24].

The dimerization of steroid receptors at first appeared to be less complicated, since heterodimerization between receptors that bind different ligands (like the thyroid receptor and the RXR) does not seem to occur. Nevertheless, heterodimerization is clearly possible between receptor subtypes (which may have some differences in ligand-binding specificity) and between receptor isoforms (which often have distinctly different transcriptional activities). Examples of subtypes and isoforms that heterodimerize are glucocorticoid receptor α and β , and progesterone receptor A and B forms, respectively. Receptor dimerization and receptor stability are important points for pharmaceutical regulation of transcription via nuclear receptors, and several hormone antagonists (some antiestrogens and antiprogestins, for example) appear to act at this level [25–28].

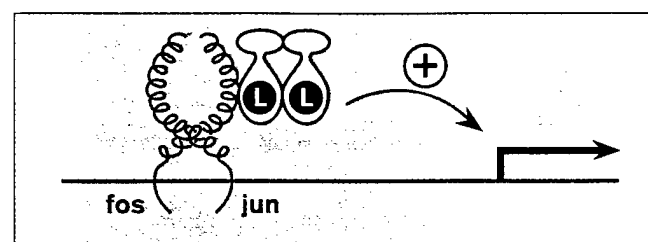
Variations also occur at a third level, the interaction of nuclear receptors with the DNA response elements. Although the response elements are often portrayed as consensus sequences — inverted or direct repeats of a defined five- to six-nucleotide sequence, with various spacers between the repeats — the response elements found in responsive genes are often nonconsensus in sequence; some are half-sites and others have multiple repeats. Often the response elements are found in complex, upstream-enhancer regions, clustered together or even overlapping with response elements for other known transcription factors, which may synergize or compete with the nuclear receptors. Sequences that flank the core response elements can also affect the DNA binding of these receptors (see, for example, [29]). And the structure of the DNA response element, since it affects the recognition between the receptor and the DNA, may also affect the interaction between the receptor and the ligand.

Given all the sources of variation described above, especially the fact that nuclear receptors may interact with or compete with a number of other sequence-specific transcription factors, it is not surprising that the response to a specific hormone depends on both the cell in which it is acting and the gene whose activity it modulates [16].

Nuclear receptor activation without direct DNA binding or without ligand binding

A curious but major deviation from the classical scheme for nuclear receptor action is gene activation in the absence of direct DNA-binding by the receptor. In this situation the promoter for a gene whose activity is clearly regulated by a nuclear receptor and its hormone appears to have no hormone-response element for the receptor, and does not, in fact, require direct DNA binding by the receptor. The hormone–receptor complex seems to function by binding to DNA indirectly via other DNA-tethered transcription factors (see, for example, [30–32]), thus acting as a ligand-modulated co-regulator, rather than a ligand-modulated transcription factor (Fig. 5).

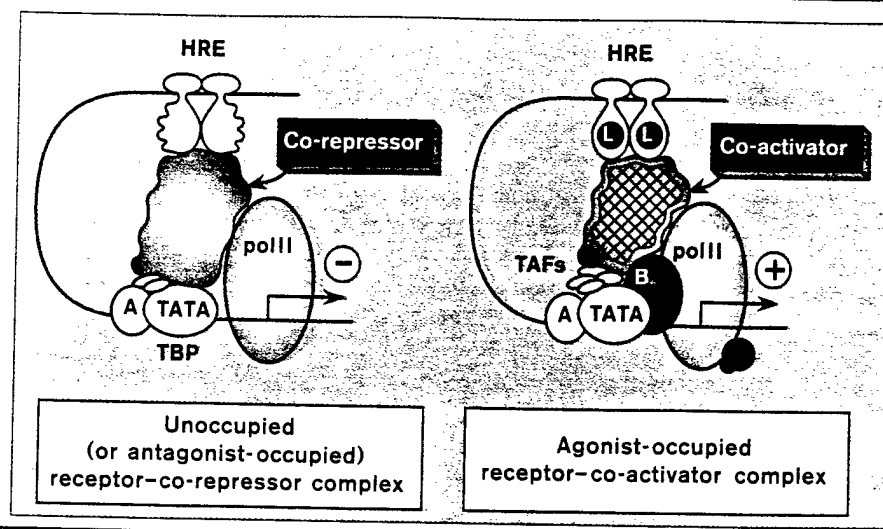
Figure 5



Nuclear receptor gene activation can occur without direct DNA binding. The nuclear receptor is tethered to DNA by a protein–protein interaction with another sequence-specific transcription factor, such as fos/jun (AP1). In such a case, the nuclear receptor has the role of a ligand-modulated co-activator of transcription.

Figure 6

Co-regulators mediate the interaction between the nuclear receptor and components of the transcription complex. Unoccupied or antagonist-occupied receptors can recruit co-repressors (left); when an agonist ligand binds, the ligand-receptor complex can recruit co-activators (right).



Another major deviation from the classical scheme for activation of genes by nuclear receptors is ligand-independent gene activation. In certain systems there appears to be significant crosstalk between signal-transduction pathways that activate transcription. The result is that growth factors or hormones that operate through receptor tyrosine kinases or via cAMP or other second messengers can activate nuclear receptor regulated genes in a manner that requires receptor but not ligand [19]. In some cases, these alternative pathways may synergize with the normal ligand-mediated pathway [33]. The molecular mechanism for such action is not well understood, but it is possible that phosphorylation of specific sites on the nuclear receptors may enhance the transcriptional activity of the unliganded receptor [19,34].

Modulation of gene transcription

Once a nuclear receptor is bound to DNA, what happens next? The final step of the classical pathway, the process by which these receptors modulate the rate of gene transcription (Fig. 6), has its own sources of regulatory complexity. First, it is important to recognize that the rate at which a gene is transcribed depends both on the local chromatin architecture, and on the rate at which an active RNA polymerase preinitiation complex can be assembled. The nuclear receptors appear to affect both of these processes, both directly and indirectly via 'transcription intermediary factors' (TIFs) [9,35,36], although their effect on chromatin architecture is poorly understood. There is evidence that DNA-bound nuclear receptors interact directly with some of the proteins comprising the basal transcription machinery, such as TFIIB or TATA-binding protein associated factors (TAFs) [37–39]. If they suppress or stimulate a rate-limiting step in the assembly of an active RNA polymerase II preinitiation complex, this would result in repression or activation of transcription. In

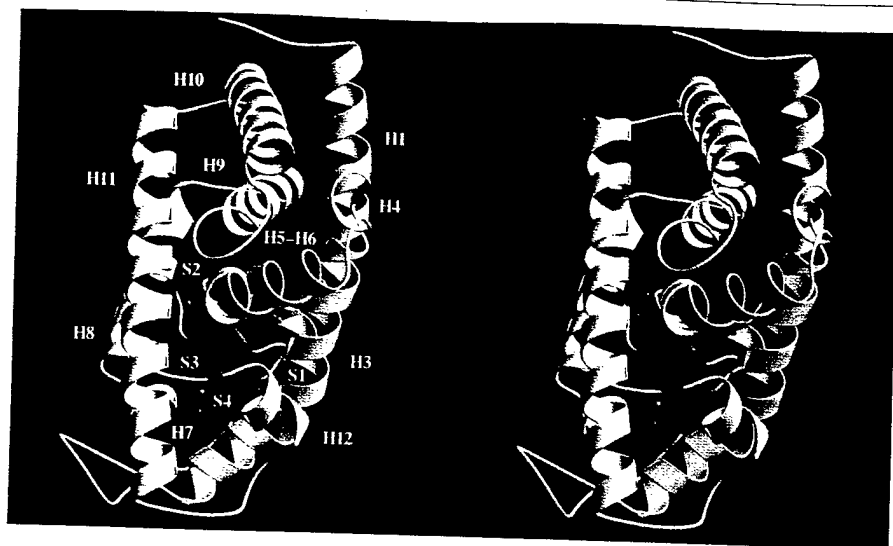
many cases the relevant interactions between nuclear receptors and basal transcription factors appear not to be direct, however, but are mediated by various co-regulators.

The co-regulators involved in nuclear-receptor modulation of gene transcription are diverse, and are being discovered at an increasing rate. They are often large multidomain proteins, with some homology to factors that are known to modulate chromatin structure; some have known protein-interaction domains, or have the ability to interact with various components in the general transcription apparatus [16]. Some also appear to fit nicely into the unliganded-repression/liganded-activation paradigm, in that one set of co-regulators binds to the unliganded thyroid and retinoid receptors to repress transcription [40,41], whereas another set binds to liganded receptor to enhance transcription [42–46]. In the case of the steroid receptors, the co-regulators appear to bind to either the amino-terminal or carboxy-terminal activation domain of the receptors. Some co-regulators interact with and influence the transcriptional activity of many steroid hormone receptors and other related receptors, such as RXR, whereas other co-regulators show a more restricted range of receptor interaction.

Structural and conformational changes on ligand binding

As the interaction between the co-regulators and the nuclear receptor is regulated by ligand binding, it is plausible that ligand binding elicits a conformational change in the receptor that may permit co-activator but not co-repressor binding in the presence of ligand (or co-repressor but not co-activator binding in the absence of ligand). Mutational mapping studies have begun to identify the different regions of the receptor that seem to be responsible for interaction with co-repressors and co-activators [35,43,47]. Most exciting are some of the structural features revealed

Figure 7



Stereoview of a ribbon structure of the ligand-binding domain of the rat thyroid hormone receptor complexed with thyroid hormone (T3), shown as a skeletal structure in the lower half of the protein. The regions of α -helical (H) and β -strand (S) secondary structure are designated.

in the recent X-ray crystal structures of three different receptor ligand-binding domains (domain E) (Fig. 7); these structures provide insight into the conformational reorganization that occurs upon ligand binding [48–51].

The ligand-binding domain of the nuclear receptors is large, larger than most single protein domains, with a unique antiparallel α -helix triple sandwich topology (Fig. 7). Approximately half of the domain consists of a rigid, tightly packed assembly of helices that appear to act as a fundement or fulcrum for the action of the remainder of the domain, which is more flexible and is involved in ligand binding. Although the three structures that have been described so far do not permit a direct comparison between the conformations of a single receptor in the liganded and unliganded state, certain general features have emerged that are likely to hold true for the ligand-induced conformational changes of all of the members of the superfamily.

In the bound state, the ligand is completely engulfed by the flexible portion of the domain, and actually forms the hydrophobic core for this region [49,50]. Six segments of secondary structure, arranged roughly as the six sides of a box, surround the ligand, with more than 20 residues making direct contact with the ligand (Fig. 8). In the liganded state, the carboxy-terminal portion of this domain, an amphipathic helix, termed the activation function 2 activation domain (AF2-AD), interacts with the ligand and is positioned adjacent to two other helical portions of the receptor whose specific orientation is also dependent upon contacts with the ligand (see Fig. 7, helix 12). This composite surface, whose integrity appears to be critically dependent on ligand binding, is one likely site for co-activator binding.

By contrast, in the unbound state, the flexible portion of the ligand-binding domain lacks its hydrophobic core,

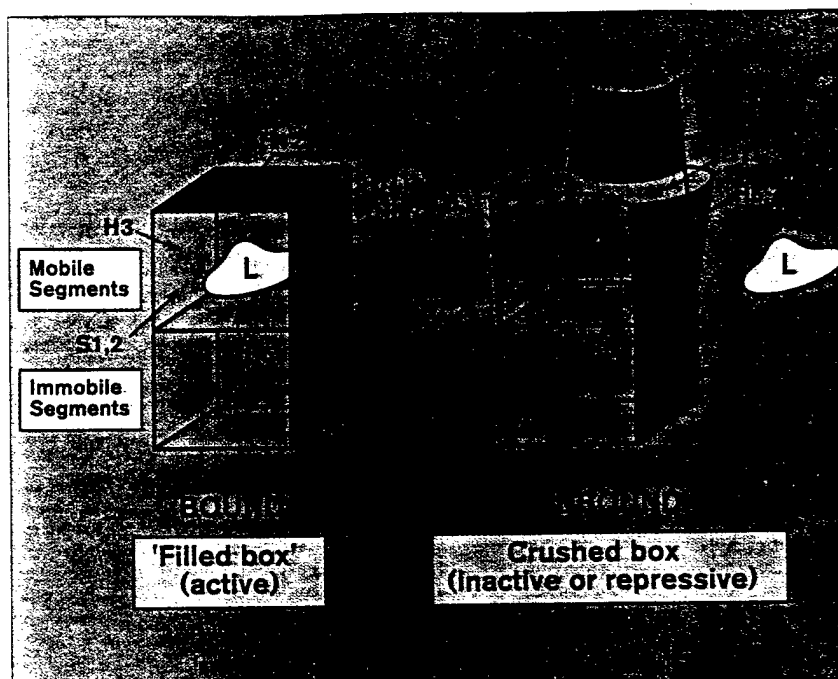
namely, the ligand. In the one published structure for an unliganded receptor [48], the box-like structure of the flexible portion of domain E appears to have collapsed, with two sides tipping inward and two sides tipping outward; the activation helix is dislodged from its position between the other two helices, since their relative position is no longer supported by contacts with the ligand (see Fig. 8). The composite surface for co-activator binding is thus absent or at least substantially modified in the unliganded state. But in the collapsed state, various new topographical features have developed, providing potential sites for co-repressor binding.

X-ray crystallography provides static pictures of protein structure. It is thus possible that the flexible ligand-binding region of domain E in the unliganded state may be rather fluid, perhaps in a molten globule-like state. The binding of heat shock proteins (which normally bind only to unfolded or partially folded proteins) and immunophilins to the unliganded steroid receptors and the sensitivity of the unliganded receptor to proteolysis supports this view [52]. Further studies, especially ones in which a direct comparison can be made between structures of the liganded and unliganded state of the same receptor, will be needed to verify the generality of these conformational transitions.

Ligand binding affects receptor shape — thus, receptor shape reflects ligand shape. As co-repressor/co-activator binding responds to alterations in receptor shape, the ligand is the crucial factor in recruiting or disbanding these important co-regulators. The view that ligand shape determines receptor shape and thus receptor activity can also account for the spectrum of biological activity — from pure agonists to partial agonists/antagonists to pure antagonists — that is known for ligands for some of these nuclear

Figure 8

A 'box model' for the ligand-binding domain of a nuclear receptor. When an agonist ligand is bound, the upper box, made up of mobile segments with the ligand at its core, is 'filled'; in this conformation, it has a structure in which the activation domain (helix 12) is in the active state, where it can interact with co-activators, activating transcription. Without ligand, the upper box is empty and is 'crushed', so that two sides cave inward and two sides bulge outward; the activation helix is displaced from the active state, and the empty receptor is thus either inactive or recruits co-repressors to become repressive. Antagonists and partial agonists fill the top box in a different manner, such that the activation helix is fully or partially misoriented from the activating position. The conformation of the lower box is not affected by ligand binding. (Note that this schematic representation of the ligand binding domain of a nuclear receptor is shown in the orientation opposite to that of the thyroid hormone receptor-T3 complex shown in Figure 7. In Figure 7, the ligand-binding 'box' is at the bottom.)



receptors, such as estrogens and progestins. Given all this, the potential for pharmaceutical modulation of the transcriptional activity of nuclear receptors is obvious [15,16].

Pharmacological issues, however, extend beyond the ligand-receptor interaction. The biological effect that a particular ligand will have, acting via a given receptor, will also depend on the intracellular context (i.e., the levels of the relevant co-regulators and transcription factors with which the receptor cooperates) and the promoter for the specific gene being regulated (i.e., the structure of the hormone-response element and whether any other transcription factors bind to nearby sites). This 'tripartite receptor pharmacology', comprising ligands, receptors, and cell and promoter specific transcriptional effectors, offers rich possibilities for developing tissue- and response-specific pharmaceuticals [16].

The future

There is much more to learn. The details of the ligand-induced conformational changes within one receptor protein are yet to be revealed, and we do not yet know how all the different domains of a nuclear receptor interact with each other. A full appreciation of the molecular interactions involved in the gene-regulating action of the nuclear receptors will require reconstitution of multiprotein complexes involving the intact receptor (as a homo- or heterodimer) interacting with a complete gene regulatory region, together with other associated transcription factors, co-regulator proteins, and elements of the general transcription apparatus. Equally important will be biological

studies detailing regulation of the levels and activity of receptors and their co-regulators as a function of physiological and developmental state in different hormonal target cells and tissues. Clearly, the major and perhaps the most exciting challenges still lie ahead.

Acknowledgements

We are grateful for support of research performed in our laboratories by grants from the National Institutes of Health. We thank Richard L Wagner for providing Figure 7.

References

1. Jensen, E.V. & Jacobson, H.I. (1962). Basic guides to the mechanism of estrogen action. *Recent Prog. Horm. Res.* **18**, 387-414.
2. Mueller, G.C., Herranen, A.M. & Jervell, K.F. (1958). Studies on the mechanism of action of estrogens. *Recent Prog. Horm. Res.* **14**, 95-129.
3. Mueller, G.C., Gorski, J. & Aizawa, Y. (1961). The role of protein synthesis in early estrogen action. *Proc. Natl. Acad. Sci. USA* **47**, 164-169.
4. O'Malley, B.W., McGuire, W.L., Kohler, P.O. & Koreman, S.G. (1969). Studies on the mechanism of steroid hormone regulation of synthesis of specific proteins. *Recent Prog. Horm. Res.* **25**, 105-160.
5. Yamamoto, K.R. & Alberts, B.M. (1976). Steroid receptors: elements for modulation of eukaryotic transcription. *Annu. Rev. Biochem.* **45**, 721-746.
6. Katzenellenbogen, B.S. & Gorski, J. (1975). Estrogen actions on syntheses of macromolecules in target cells. In *Biochemical Action of Hormones* (vol. 3). (Litwack, G. ed.), pp 187-243. Academic Press, New York.
7. Evans, R.M. (1988). The steroid and thyroid hormone receptor superfamily. *Science* **240**, 889-895.
8. Gronemeyer, H. (1991). Transcription activation by estrogen and progesterone receptors. *Annu. Rev. Genet.* **25**, 89-123.
9. Tsai, M.-J. & O'Malley, B.W. (1994). Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu. Rev. Biochem.* **63**, 451-486.
10. Beato, M., Herrlich, P. & Schütz, G. (1995). Steroid hormone receptors: Many actors in search of a plot. *Cell* **83**, 851-857.
11. Mangelsdorf, D.J., et al., & Evans, R.M. (1995). The nuclear receptor

- superfamily: the second decade. *Cell* **83**, 835–839.
12. Mangelisford, D.J. & Evans, R.M. (1995). The RXR heterodimers and orphan receptors. *Cell* **83**, 841–850.
 13. Schwabe, J.W.R., Chapman, L., Finch, J.T., Rhodes, D. & Neuhaus, D. (1993). DNA recognition by the oestrogen receptor: from solution to the crystal. *Structure* **1**, 187–204.
 14. Katzenellenbogen, J.A. (1992). Probes for steroid receptors – studies at the molecular, cellular, and whole organism levels. In *Proceedings of The Robert A. Welch Foundation Conference on Chemical Research, XXXV. Chemistry at the Frontiers of Medicine*, pp. 229–257, Robert A. Welch Foundation, Houston TX.
 15. Rosen, J., Day, A., Jones, T.K., Nadzan, A.M. & Stein, R.B. (1995). Intracellular receptors and signal transducers and activators of transcription superfamilies: novel targets for small-molecule drug discovery. *J. Med. Chem.* **38**, 4855–4874.
 16. Katzenellenbogen, J.A., O'Malley, B.W. & Katzenellenbogen, B.S. (1996). Tripartite steroid hormone receptor pharmacology: interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. *Mol. Endocrinol.* **10**, 119–131.
 17. Pappas, T.C., Gametchu, B. & Watson, C.S. (1995). Membrane estrogen receptors identified by multiple antibody labeling and impeded ligand binding. *FASEB J.* **9**, 404–410.
 18. Migliaccio, A., et al., & Auricchio, F. (1996). Tyrosine kinase/p21^{ras}/MAP-kinase pathway activation by estradiol-receptor complex in MCF-7 cells. *EMBO J.* **15**, 1292–1300.
 19. Katzenellenbogen, B.S. (1996). Estrogen receptors: bioactivities and interactions with cell signaling pathways. *Biol. Reprod.* **54**:287–293.
 20. Glass, C.K. (1994). Differential recognition of target genes by nuclear receptor monomers, dimers, and heterodimers. *Endocr. Rev.* **15**, 391–407.
 21. Qi, J.S., Desai-Yajnik, V., Greene, M.E., Raaka, B.M. & Samuels, H.H. (1995). The ligand binding domains of the thyroid hormone/retinoid receptor gene subfamily function *in vivo* to mediate heterodimerization, gene silencing, and transactivation. *Mol. Cell. Biol.* **15**, 1817–1825.
 22. Nawaz, Z., Tsai, M.-J. & O'Malley, B.W. (1995). Specific mutations in the ligand-binding domain selectively abolish the silencing function of human thyroid hormone receptor β . *Proc. Natl. Acad. Sci. USA* **92**, 11691–11695.
 23. Smith, D.F. & Toft, D.O. (1993). Steroid receptors and their associated proteins. *Mol. Endocrinol.* **7**, 4–11.
 24. Cheskis, B. & Freedman, L.P. (1994). Ligand modulates the conversion of DNA-bound vitamin D3 receptor (VDR) homodimers into VDR-retinoid X receptor homodimers. *Mol. Cell. Biol.* **14**, 3329–3338.
 25. Fawell, S.E., White, R., Hoare, S., Sydenham, M., Page, M. & Parker, M.G. (1990). Inhibition of estrogen receptor-DNA binding by the 'pure' antiestrogen ICI 164,384 appears to be mediated by impaired receptor dimerization. *Proc. Natl. Acad. Sci. USA* **87**, 6883–6887.
 26. Gibson, M.K., Nemmers, L.A., Beckman, Jr, W.C., Davis, V.L., Curtis, S.W. & Korach, K.S. (1991). The mechanism of ICI 164,384 antiestrogenicity involves rapid loss of estrogen receptor in uterine tissue. *Endocrinology* **129**, 2000–2010.
 27. Dauvois, S., Danielian, P.S., White, R. & Parker, M.G. (1992). Antiestrogen ICI 164,384 reduces cellular estrogen receptor content by increasing turnover. *Proc. Natl. Acad. Sci. USA* **89**, 4037–4041.
 28. Reese, J.C. & Katzenellenbogen, B.S. (1992). Examination of the DNA-binding abilities of estrogen receptor in whole cells: implications for hormone-independent transactivation and the action of the pure antiestrogen ICI 164,384. *Mol. Cell. Biol.* **12**, 4531–4538.
 29. Kraus, W.L., Montano, M.M. & Katzenellenbogen, B.S. (1994). Identification of multiple, widely spaced estrogen-responsive regions in the rat progesterone receptor gene. *Mol. Endocrinol.* **8**, 952–969.
 30. Kallio, P.J., Poukka, H., Moilanen, A., Jänne, O.A. & Palvimäki, J.J. (1995). Androgen receptor-mediated transcriptional regulation in the absence of direct interaction with a specific DNA element. *Endocrinology* **9**, 1017–1028.
 31. Webb, P., Lopez, G.N., Uht, R.M. & Kushner, P.J. (1995). Tamoxifen activation of the estrogen receptor/AP-1 pathway: potential origin for the cell-specific estrogen-like effects of antiestrogens. *Mol. Endocrinol.* **9**, 443–456.
 32. Sukovich, D.A., Mukherjee, R. & Benfield, P.A. (1994). A novel cell-specific mechanism for estrogen receptor mediated gene activation in the absence of an estrogen-responsive element. *Mol. Cell. Biol.* **14**, 7134–7143.
 33. Cho, H. & Katzenellenbogen, B.S. (1993). Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Mol. Endocrinol.* **7**, 441–452.
 34. Kato, S., et al., & Chambon, P. (1995). Activation of the estrogen receptor through phosphorylation by mitogen-activated protein kinase. *Science* **270**, 1491–1494.
 35. Le Douarin, B., et al., & Losson, R. (1995). The N-terminal part of TIF1, a putative mediator of the ligand-dependent activation function (AF-2) of nuclear receptors, is fused to B-raf in the oncogenic protein T18. *EMBO J.* **14**, 2020–2033.
 36. vom Baur, E., et al., & Losson, R. (1996). Differential ligand-dependent interactions between the AF-2 activating domain of nuclear receptors and the putative transcriptional intermediary factors mSUG1 and TIF1. *EMBO J.* **15**, 110–124.
 37. Ing, N.H., Beekman, J.M., Tsai, S.Y., Tsai, M.-J. & O'Malley, B.W. (1992). Members of the steroid hormone receptor superfamily interact with TFIIIB (S300II). *J. Biol. Chem.* **267**, 17617–17623.
 38. Blanco, J., et al., & Ozato, K. (1995). TFIIIB and vitamin D receptor cooperatively activate ligand dependent transcription. *Proc. Natl. Acad. Sci. USA* **92**, 1535–1539.
 39. Jacq, X., Brou, C., Lutz, Y., Davidson, I., Chambon, P. & Tora, L. (1994). Human TAFII30 is present in a distinct TFIID complex and is required for transcriptional activation by the estrogen receptor. *Cell* **79**, 107–117.
 40. Chen, J.D. & Evans, R.M. (1995). A transcriptional co-repressor that interacts with nuclear hormone receptors. *Nature* **377**, 454–457.
 41. Hörlein, A.J., et al., & Rosenfeld, M.G. (1995). Ligand-independent repression by the thyroid hormone receptor mediated by a nuclear receptor co-repressor. *Nature* **377**, 397–404.
 42. Oñate, S.A., Tsai, S.Y., Tsai, M.-J. & O'Malley, B.W. (1995). Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* **270**, 1354–1357.
 43. Halachmi, S., Marden, E., Martin, G., MacKay, H., Abbondanza, C. & Brown, M. (1994). Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription. *Science* **264**, 1455–1458.
 44. Cavaillès, V., et al., & Parker, M.G. (1995). Nuclear factor RIP140 modulates transcriptional activation by the estrogen receptor. *EMBO J.* **14**, 3741–3751.
 45. Landel, C.C., Kushner, P.J. & Greene, G.L. (1994). The interaction of human estrogen receptor with DNA is modulated by receptor-associated proteins. *Mol. Endocrinol.* **8**, 1407–1419.
 46. Lee, J.W., Ryan, F., Swaffield, J.C., Johnston, S.A. & Moore, D.D. (1995). Interaction of thyroid-hormone receptor with a conserved transcriptional mediator. *Nature* **374**, 91–94.
 47. Cavaillès, V., Dauvois, S., Danielian, P.S. & Parker, M.G. (1994). Interaction of proteins with transcriptionally active estrogen receptors. *Proc. Natl. Acad. Sci. USA* **91**, 10009–10013.
 48. Renaud, J.-P., et al., & Moras, D. (1995). Crystal structure of the RAR- γ ligand-binding domain bound to all-trans retinoic acid. *Nature* **378**, 681–689.
 49. Wagner, R.L., Apriletti, J.W., McGrath, M.E., West, B.L., Baxter, J.D. & Fletterick, R.J. (1995). A structural role for hormone in the thyroid hormone receptor. *Nature* **378**, 690–697.
 50. Bourguet, W., Ruff, M., Chambon, P., Gronemeyer, H. & Moras, D. (1995). Crystal structure of the ligand-binding domain of the human nuclear receptor RXR- α . *Nature* **375**, 377–382.
 51. Wurtz, J.M., et al., & Gronemeyer, H. (1996). A canonical structure for the ligand-binding domain of nuclear receptors. *Nat. Struct. Biol.* **3**, 87–94.
 52. Allan, G.F., et al., & O'Malley, B.W. (1992). Hormone and antihormone induce distinct conformational changes which are central to steroid receptor activation. *J. Biol. Chem.* **267**, 19513–19520.



Response-specific Antiestrogen Resistance in a Newly Characterized MCF-7 Human Breast Cancer Cell Line Resulting from Long-term Exposure To *Trans*-hydroxytamoxifen

Mary E. Herman and Benita S. Katzenellenbogen*

Departments of Molecular and Integrative Physiology, Cell and Structural Biology, University of Illinois, Urbana, IL 61801-3704, U.S.A.

To understand better the antiestrogen-resistant phenotype that frequently develops in breast cancer patients receiving tamoxifen, we cultured MCF-7 breast cancer cells long-term (>1 yr) in the presence of the antiestrogen *trans*-hydroxytamoxifen (TOT) to generate a subline refractory to the growth-suppressive effects of TOT. This subline (designated MCF/TOT) showed growth stimulation, rather than inhibition, with TOT and diminished growth stimulation with estradiol (E_2), yet remained as sensitive as the parental cells to growth suppression by another antiestrogen, ICI 164,384. Estrogen receptor (ER) levels were maintained at 40% of that in parent MCF-7 cells, but MCF/TOT cells failed to show an increase in progesterone receptor content in response to E_2 or TOT treatment. In contrast, the MCF/TOT subline behaved like parental cells in terms of E_2 and TOT regulation of ER and pS2 expression and transactivation of a transiently transfected estrogen-responsive gene construct. DNA sequencing of the hormone binding domain of the ER from both MCF-7 and MCF/TOT cells confirmed the presence of wild-type ER and exon 5 and exon 7 deletion splice variants, but showed no point mutations. Compared to the parental cells, the MCF/TOT subline showed reduced sensitivity to the growth-suppressive effects of retinoic acid and complete resistance to exogenous TGF- β 1. The altered growth responsiveness of MCF/TOT cells to TOT and TGF- β 1 was partly to fully reversible following TOT withdrawal for 16 weeks. Our findings underscore the fact that antiestrogen resistance is response-specific; that loss of growth suppression by TOT appears to be due to the acquisition of weak growth stimulation; and that resistance to TOT does not mean global resistance to other more pure antiestrogens such as ICI 164,384, implying that these antiestrogens must act by somewhat different mechanisms. The association of reduced retinoic acid responsiveness and insensitivity to exogenous TGF- β with antiestrogen growth resistance in these cells supports the increasing evidence for interrelationships among cell regulatory pathways utilized by these three growth-suppressive agents in breast cancer cells. In addition, our findings indicate that one mechanism of antiestrogen resistance, as seen in MCF/TOT cells, may involve alterations in growth factor and other hormonal pathways that affect the ER response pathway.

Copyright © 1996 Elsevier Science Ltd.

J. Steroid Biochem. Molec. Biol., Vol. 59, No. 2, pp. 121–134, 1996

INTRODUCTION

Tamoxifen is the most common endocrine therapy used in the treatment of estrogen receptor-positive

breast cancer. Unfortunately, the vast majority of tamoxifen-treated breast tumors eventually become refractory to the beneficial effects of this antiestrogen. Characterization of tamoxifen-resistant breast tumors established in nude mice [1, 2] and in culture [3–6] has shown that reductions in estrogen receptor (ER) content or changes in ER binding affinity for ligands are not necessarily causative factors in antiestrogen re-

*Correspondence to Dr Benita S. Katzenellenbogen. Tel: +1 217 333 7838 or +1 217 333 9769; Fax: +1 217 244 9906; e-mail: katzenel@uiuc.edu.

Received 27 Mar. 1996; accepted 6 May 1996.

sistance. Whereas changes in ER-mediated transcriptional activity may confer or promote antiestrogen resistance, it is also possible that this phenotype may be influenced by interactions with other regulatory pathways. There is an emerging body of evidence that shows cross-talk of the ER pathway [7, 8] with peptide growth factors and with other nuclear receptor ligands, such as the retinoids [9–11] suggesting that these may be involved in antiestrogen resistance and in the more aggressive behavior often associated with antiestrogen-resistant tumors.

In normal and neoplastic epithelial cells, the transforming growth factor- β s (TGF- β s) are most frequently associated with growth inhibition, whereas in a number of cell types, such as fibroblasts, the TGF- β s are growth stimulatory (for review, see [12]). The finding that tamoxifen increases TGF- β levels in tumors suggests that the therapeutic effect of tamoxifen in slowing or arresting tumor growth may be partly attributable to the growth-inhibitory action of the TGF- β s [13]. It has been demonstrated that estrogens suppress and antiestrogens augment TGF- β expression in human breast cancer cell lines in culture [14, 15]. Interestingly, a number of advanced stage tumors and cancer cell lines exhibit a TGF- β -resistant phenotype (for example, [16]), suggesting that the development of TGF- β resistance may abrogate the beneficial effects of tamoxifen on breast cancer cells.

We maintained MCF-7 human breast cancer cells in *trans*-hydroxytamoxifen (TOT) for more than 1 year to generate an *in vitro* model for the study of tamoxifen resistance. Herein, we report on the proliferation of the cells, and the activity of the estrogen receptor and its responsiveness to estrogen and to two different classes of antiestrogens, as well as on the effects of TGF- β and retinoic acid on this subline. Our findings suggest interrelationships among the pathways utilized by antiestrogens, TGF- β and retinoic acid in the regulation of these breast cancer cells.

MATERIALS AND METHODS

Materials

Radioinert E_2 and R5020 (promegestone; 17,21-dimethyl-19-nor-pregna-4,9-diene-3,20-dione), nutritional supplements for growth in serum-free conditions, protease inhibitors, TPA (12-O-tetradecanoylphorbol-13-acetate), MTT (thiazolyl blue), all-*trans*-retinoic acid and sera were purchased from Sigma Chemical Co. (St Louis, MO). *Trans*-hydroxytamoxifen (TOT), ICI 182,780 and ICI 164,384 (ICI) were generously provided by Zeneca Pharmaceuticals (Macclesfield, U.K.). Tissue culture media and antibiotics were purchased from GIBCO (Grand Island, NY). Tritiated E_2 (2,4,6,7- 3H -N-estradiol) and 3H -R5020 (17- α -methyl- 3H -promegestone) were purchased from New England

Nuclear Corp. (Boston, MA) and methyl[3H]thymidine from ICN, Costa Mesa, CA.

Cell culture

MCF-7 human breast cancer cells were acquired from the Michigan Cancer Foundation; cells between passage numbers 150 and 300 were used in these studies. Parent MCF-7 cells were routinely cultured in phenol red-containing Eagle's minimal essential medium (MEM) supplemented with 5% heat-inactivated fetal calf serum (FCS), E_2 (10^{-12} M), 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid buffer (10 mM), insulin (6 ng/ml), penicillin (100 units/ml), streptomycin (100 μ g/ml), and gentamicin (50 μ g/ml). To generate TOT-resistant MCF-7 sublines, cells were maintained in the above media without supplemented E_2 , and with 10-fold increases in TOT concentration (10^{-9} M– 10^{-6} M) every 4 weeks. The cells were thereafter routinely maintained with 10^{-6} M TOT. Cells were subcultured weekly at near confluence using 1 mM EDTA prepared in Hank's balanced salt solution and medium was replenished every other day. To generate clonal-derived sublines, 96-well plates were seeded at approximately one cell every three wells. Two weeks after seeding, wells containing only one colony were identified. Clonal-derived sublines were maintained and sequentially transferred to 24-well plates, then six-well plates and T25 flasks.

For all studies involving experimental treatments, cells were grown without E_2 for 1 week or without TOT for 2 weeks and then subsequently in 5% CDFCS IMEM without insulin for an additional 5–10 days prior to the experiment, in order to deplete the cells of E_2 or TOT prior to the onset of experiments.

Cell proliferation studies

To determine cell number, cells were seeded at 150,000 cells/T25 flask in triplicate and after 2 days day 0 flasks were counted and the medium was replaced and treatments added. Media were changed every 2 days and cells in logarithmic phase were harvested on day 6 and counted in a Coulter particle counter (Hialeah, FL).

Anchorage-independent growth was determined by a colony-forming assay. In brief, six-well plates were coated with 0.6% agar in 5% CDFCS IMEM and allowed to cool. Cell suspensions containing 10,000 cells were passed through a 22-gauge needle and then added to a mixture equilibrated to 45°C containing 0.4% agar, 5% CDFCS IMEM and treatments and added to the wells. Plates were grown for 2 weeks with a top layer of media which was replenished every 3 days. Colony size ($>60 \mu$) was determined microscopically with an ocular grid (Wild M40 microscope; Heerbrugg, Switzerland).

In some studies, cell number was determined by the MTT assay. MTT (thiazolyl blue) is converted from a yellow-colored salt to a purple-colored formazan by cleavage of the tetrazolium ring by mitochondrial dehydrogenases, the activity of which is linear with cell number. Cells were seeded at 2000–5000 cells/well in 96-well plates in quadruplicate. After treatment as indicated, 50 μ l of 2 mg/ml MTT was added and plates were incubated at 37°C for 4 h. Wells were drained and formazan crystals were solubilized in 150 μ l buffer (20% w/v sodium dodecyl sulfate dissolved in 50% dimethylformamide/50% dH₂O containing 2.5% acetic acid and 2.5% of 1 N HCl with a final pH of 4.7 [17]. Absorbance at 570 nm was determined on a plate reader.

For [³H]thymidine incorporation studies, cells were seeded at 2000 cells/well in 24-well dishes. Two or 3 days later the wells were washed in serum-free media for 2 h and then treated in serum-supplemented or in serum-free IMEM with 1 μ g/ml fibronectin, 2 μ g/ml transferrin and 1:100 dilution of trace elements. After 3 or 4 days, the cells were incubated with 0.5 μ Ci methyl[³H]thymidine at 37°C for 2 h. Plates were sequentially washed and fixed with ice-cold PBS, 10% TCA (2 \times), MeOH, and then incorporated label was recovered by incubation of the wells in 0.5 N NaOH for 30 min at 37°C. Lysates were transferred to vials containing ScintiVerseTM cocktail (Fisher Scientific, Pittsburgh, PA) and [³H]thymidine was determined in a scintillation counter.

Whole cell binding assays

Whole cell ER and progesterone receptor (PgR) binding assays were done as previously described [18]. Cells were incubated with 10 nM [³H]E₂ or [³H]R5020 in the absence or presence of a 100-fold excess of unlabelled ligand, and for PgR studies, with 3.75 ng/ml hydrocortisone. After incubating at 37°C for 40 min, cells were washed three times with 1% Tween-80 in phosphate-buffered saline and bound radiolabelled ligand was extracted with ethanol and counted in a scintillation counter.

Western blot analysis

Subconfluent cell layers were pelleted and resuspended in 50 mM Tris (pH 7.4), 7.5 mM EDTA, 0.6 M NaCl, 10% glycerol in the presence of proteinase inhibitors (leupeptin, pepstatin A, phenylmethylsulfonylfluoride) and homogenized on ice. Samples were centrifuged for 25 min at 46 K and the protein content in the supernatants determined in a BCA assay (Pierce Chemical Co., Rockford, IL). Samples (150 μ g) were boiled for 5 min in loading buffer, separated on a SDS polyacrylamide stacking gel and transferred to nitrocellulose. Blots were incubated with estrogen receptor-specific antibodies H222 (exon 7 epitope) or with H226 (exon 1,2 epitope) in combination with D547 (exon 4 epitope), then a bridging

rabbit anti-rat IgG, and finally with [¹²⁵I]protein A, and then exposed to film [19].

TGF- β protein determinations

Subconfluent cell layers were washed three times for 1 h in serum-free media and then incubated in serum-free media supplemented with 2 μ g/ml transferrin, 1 μ g/ml fibronectin and 1:100 trace elements. After 48 h, BSA was added to the conditioned media to a final concentration of 0.5 mg/ml and the samples were snap frozen and later tested for the ability to inhibit [³H]thymidine incorporation by MV 1 Lu mink lung epithelial cells. Latent and total TGF- β bioactivity was kindly determined by Anita Roberts and Nan Roche of NCI, Bethesda, MD as described [20].

Transient transfections and assays for reporter activity

To measure responsiveness to E₂, a construct containing the consensus estrogen response element linked to a thymidine kinase promoter and the CAT gene (ERE-tk-CAT) was cotransfected into cells along with the internal control plasmid, CMV- β -gal, exactly as described [20] and cell extracts were assayed for CAT activity. Fold inductions within each assay were normalized against β -galactosidase activity as described [20].

Isolation of RNA

Isolation of total RNA from near confluent cell monolayers was performed using guanidinium thiocyanate-phenol-chloroform extraction with some modifications as described [20].

Northern blot analysis

For studies involving the induction of pS2 mRNA, cells were pretreated in 5% CDFCS IMEM as described in the Materials and methods section and treated with the ligands indicated for 12 h. Twenty micrograms total RNA were separated by electrophoresis, transferred to a nylon support and hybridized with random primer labelled fragments of human pS2 cDNA [21]. Sizes of bands were confirmed by comparison to a 0.24–9.5 kb RNA ladder (GIBCO BRL, Grand Island, NY).

Ribonuclease protection assays

Ten to 30 μ g of RNA was co-precipitated with *in vitro* transcribed, gel purified cRNA labelled with phosphorus-32 and resuspended in 80% formamide/0.1 M Na citrate (pH 6.4)/0.3 M NaOAc (pH 6.4)/1 mM EDTA. Samples were heated to 85°C for 5 min and hybridized overnight at 45°C. Unhybridized total RNA and probe was digested in a final concentration of 5 units/ml RNase A and 1000 units/ml RNase T1 for 30 min at 37°C. The sizes of protected fragments were confirmed by comparison to a lane loaded with a 0.16–1.77 kb RNA ladder (GIBCO). The probes

used were a 240 bp Mbo II segment of TGF- β 1 cDNA, a Hpa I segment of TGF- β 2/sp72 cDNA, and a 125 bp Nde I segment of TGF- β 3 cDNA as described previously [20]. The probes for TGF- β Type I and II receptors were a 300 bp unprotected Hinc II fragment (220 bp protected fragment) and a 360 bp unprotected Xho I fragment (260 bp protected fragment), respectively, kindly provided by Dr M. Brattain. A 125 bp fragment of human β -actin (Ambion Inc., Austin, TX) was used as an internal control. The relative intensity of the bands was quantitated on an UltraScan XL densitometer using GelScan XL evaluation software.

[125 I]TGF- β 1 binding assay

Cells at 75–90% confluency in 24-well plates were washed three times over 1 h with serum-free media supplemented with 0.1% BSA and incubated with 10^{-10} M [125 I]TGF- β 1 with or without a 100-fold excess of cold TGF- β 1 for 45 min. Cells were then washed four times with 0.1% BSA in ice-cold HBSS and solubilized with 1% Triton X-100/20 mM HEPES, pH 7.4/10% glycerol/0.01% BSA for 15 min at 37°C. Solubilized fractions were counted in a gamma counter [20].

RT-PCR amplification, cloning and sequence analysis

Samples of RNA, isolated from parental MCF-7 and MCF/TOT cells as described above, were reverse transcribed by AMV reverse transcriptase (Promega Corp., Madison, WI) and amplified using sense and

antisense primers specific for sequences flanking the hormone binding domain of the estrogen receptor (forward primer corresponding to estrogen receptor cDNA nucleotides 1036–1052, and reverse primer corresponding to nucleotides 1946–1967, respectively) using a PTC-100 programmable thermal controller (MJ Research Inc., Watertown, MA). Products were separated and purified from agarose gel electrophoresis and sequenced directly (Sequenase version 2.0; U.S. Biochemical Corp., Cleveland, OH), according to Newton *et al.* [22]. Sequencing reactions were analysed on 6% denaturing polyacrylamide gels. Sequences were compared to that reported for the human estrogen receptor in the Genetic Sequence Data Bank (EMBL/GenBank).

RESULTS

Growth responsiveness of parent MCF-7 and MCF/TOT cells to estrogen and antiestrogens

To generate TOT-resistant MCF-7 sublines, cells were cultured with 10-fold increases in TOT concentration (10^{-9} M– 10^{-6} M) every 4 weeks, as described in Materials and methods. The cells were thereafter routinely maintained with 10^{-6} M TOT in their culture medium. Under this regimen, dramatically slowed growth rates were observed for approximately 30 weeks from initial TOT exposure, after which time cell growth rates progressively increased. The experiments described herein were conducted between 60 and 140 weeks of maintenance on TOT, during

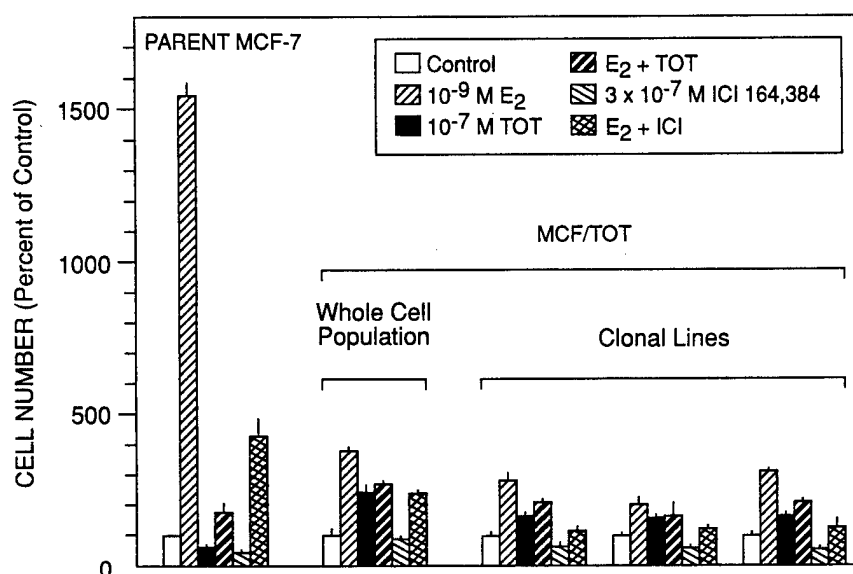


Fig. 1. Anchorage-dependent growth responsiveness of parent MCF-7 and MCF/TOT cells to estrogen and antiestrogens. Cell number in triplicate T25 flasks was determined on day 6 of treatment with the indicated compounds. Treatments were with 10^{-9} M E₂, 10^{-7} M TOT, and 3×10^{-7} M ICI 164,384 alone or together. Values are expressed as percentage of cell number in ethanol-treated control flasks. Cells were depleted of steroids and TOT for 3 weeks prior to the onset of the experiment as described in Materials and methods. The basal growth rates of the MCF-7 and MCF/TOT sublines were 3.95 ± 0.01 and 3.34 ± 0.07 days/population doubling, respectively. Data represent mean \pm SEM ($n = 3$).

which time population doubling rates were comparable in the parent MCF-7 and MCF/TOT cells (1.3 ± 0.1 and 1.6 ± 0.1 days, respectively). To determine the proliferative effects of estrogen and antiestrogens on parent MCF-7 and MCF/TOT cells, growth rates were slowed to approximately 3–4 days/population doubling by transfer from steroid- and/or TOT- and phenol red-containing media to media lacking phenol red [23] and TOT and depleted of steroids by charcoal-dextran treatment of the serum. Parent MCF-7 cells exhibited dramatic increases in cell proliferation rate in response to treatment with 10^{-9} M E_2 ($1535 \pm 374\%$ of control; Fig. 1). Treatment with the pure antiestrogen, ICI 164,384 (ICI), partly reversed estrogen-stimulated growth ($432 \pm 163\%$) and was growth suppressive when administered alone ($44 \pm 10\%$). Similar results were found when a structurally related pure antiestrogen, ICI 182,780, was used (data not shown). Treatment with the antiestrogen TOT reduced the growth of the parent MCF-7 cells ($61 \pm 9\%$ of control) and also very effectively suppressed the proliferation of these cells stimulated by E_2 .

MCF/TOT cells were growth stimulated by 10^{-9} M E_2 ($387 \pm 54\%$; Fig. 1), but this response was modest compared to the dramatic effect of estrogen stimulation on the parent MCF-7 cells. Interestingly, we found that the effect of treatment with TOT shifted from growth suppression, as observed in the parent MCF-7 cells, to growth stimulation in the MCF/TOT subline ($247 \pm 59\%$). These results suggest that MCF/TOT cells were not refractory to TOT, but instead interpreted this ligand as an agonist. Treatment with the pure antiestrogen, ICI 164,384, reduced the growth of MCF/TOT cells slightly ($68 \pm 17\%$), and partly reversed E_2 -stimulated growth ($242 \pm 32\%$), as did ICI 182,780 (data not shown). This indicates that MCF/TOT cells were not cross-resistant to pure antagonists of the estrogen receptor.

We were also interested in determining whether the altered phenotype of the MCF/TOT subline was homogeneous or heterogeneous within the cell population. Clonal lines were found to exhibit a growth phenotype similar to that of the MCF/TOT whole cell population (Fig. 1).

MCF/TOT cells showed responses to estrogen and antiestrogen in anchorage-independent colony formation assays (Fig. 2) similar to those observed in the anchorage-dependent cell proliferation assays of Fig. 1. MCF/TOT cells grown in soft agar were E_2 stimulated in terms of colony formation, although to a lesser extent than the parent MCF-7 cells (Fig. 2), and MCF/TOT cells were also growth stimulated by TOT and growth inhibited by ICI 164,384. In contrast, parental MCF-7 cells were inhibited by both TOT and ICI 164,384. Interestingly, ICI 164,384 reversed the growth stimulation observed in MCF/TOT cells in response to treatment with TOT.

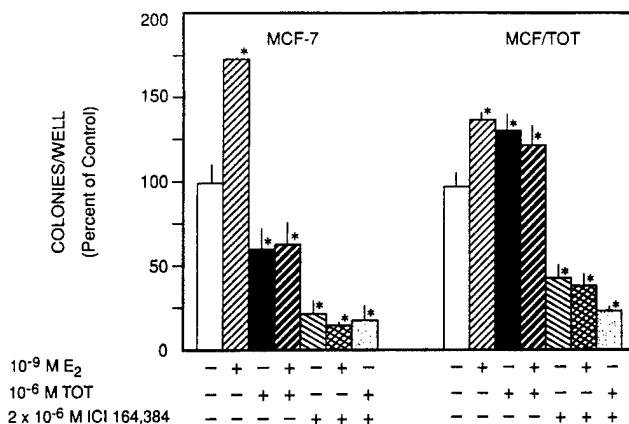


Fig. 2. Anchorage-independent growth responsiveness of parent MCF-7 and MCF/TOT cells to estrogen and antiestrogens. Parent MCF-7 and MCF/TOT cells were seeded at 10,000 cells/well in six-well plates in a top layer of 0.4% agar, 5% CDFCS IMEM and treatments and over a solidified bottom layer of 0.6% agar in 5% CDFCS IMEM. Colonies larger than 60μ were counted microscopically with an ocular grid on day 14 of treatment. Colony number from ethanol-treated control wells was not dramatically different between the MCF-7 and MCF/TOT sublines (838 ± 45 and 951 ± 126 colonies/well, respectively), nor from two separate clonal-derived sublines of MCF/TOT cells (1014 ± 430 colonies/well; data not shown). Values are expressed as percentage of colony number \pm SEM of ethanol-treated control wells from three separate experiments; *value significantly different from the control treatment at $P < 0.05$ by Student's t -test.

Assessment of antiestrogen antagonism of estrogen-stimulated growth and pS2 mRNA expression

Treatment with TOT abolished E_2 -stimulated growth in parent MCF-7 cells in a dose-dependent manner (Fig. 3, panel A). Fifty per cent suppression was achieved with ca. 1×10^{-9} M TOT, and the highest concentration of TOT tested (2×10^{-6} M) gave nearly complete suppression of E_2 -stimulated growth in parental MCF-7 cells. MCF/TOT cells were much less sensitive to suppression of E_2 -stimulated growth by TOT (Fig. 3, panel A). No suppression of E_2 -stimulated growth was seen until concentrations of TOT greater than 2×10^{-9} M were used, and 50% suppression required a concentration of TOT approximately 1000 times greater than that required by the parental MCF-7 cells (i.e., 10^{-6} M). In contrast, the pure antiestrogen, ICI 164,384, showed similar dose-response curves for inhibition of E_2 -stimulated growth in MCF-7 and MCF/TOT cells (Fig. 3, panel B).

Induction of pS2 mRNA, an early primary response to estrogen in MCF-7 cells [24], was used as an additional end-point to compare the ability of TOT to moderate E_2 -stimulated responses in MCF/TOT vs. parental MCF-7 cells. Interestingly, unlike proliferation, the dose response for TOT reversal of E_2 -stimulated pS2 mRNA was similar in parent MCF-7 and MCF/TOT cells (Fig. 4). Also as shown in Fig. 4

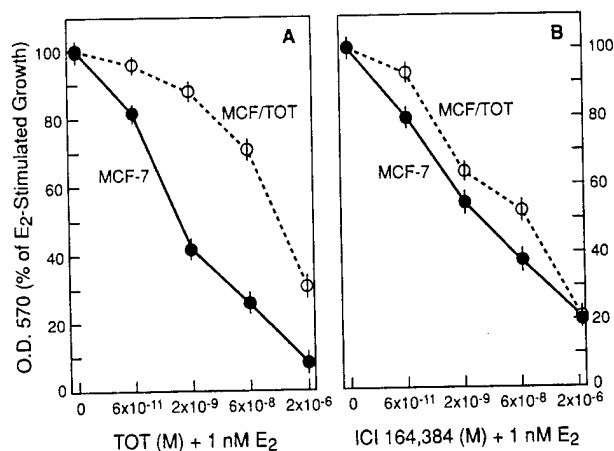


Fig. 3. Antiestrogen antagonism of E₂-stimulated growth. MCF-7 and MCF/TOT cells were seeded in quadruplicate at 2000 cells/well in 96-well plates and cotreated with 10⁻⁹ M E₂ and the indicated concentrations of TOT or ICI 164,384. Treatments were replenished on day 3 and cell number was determined by the MTT assay on day 6. E₂-stimulated growth was 953 ± 50% and 372 ± 14% of untreated, control cells in the parent MCF-7 and MCF/TOT cells, respectively. Values are expressed as percentage of absorbance in E₂-treated wells (*n* = 4; mean ± SEM).

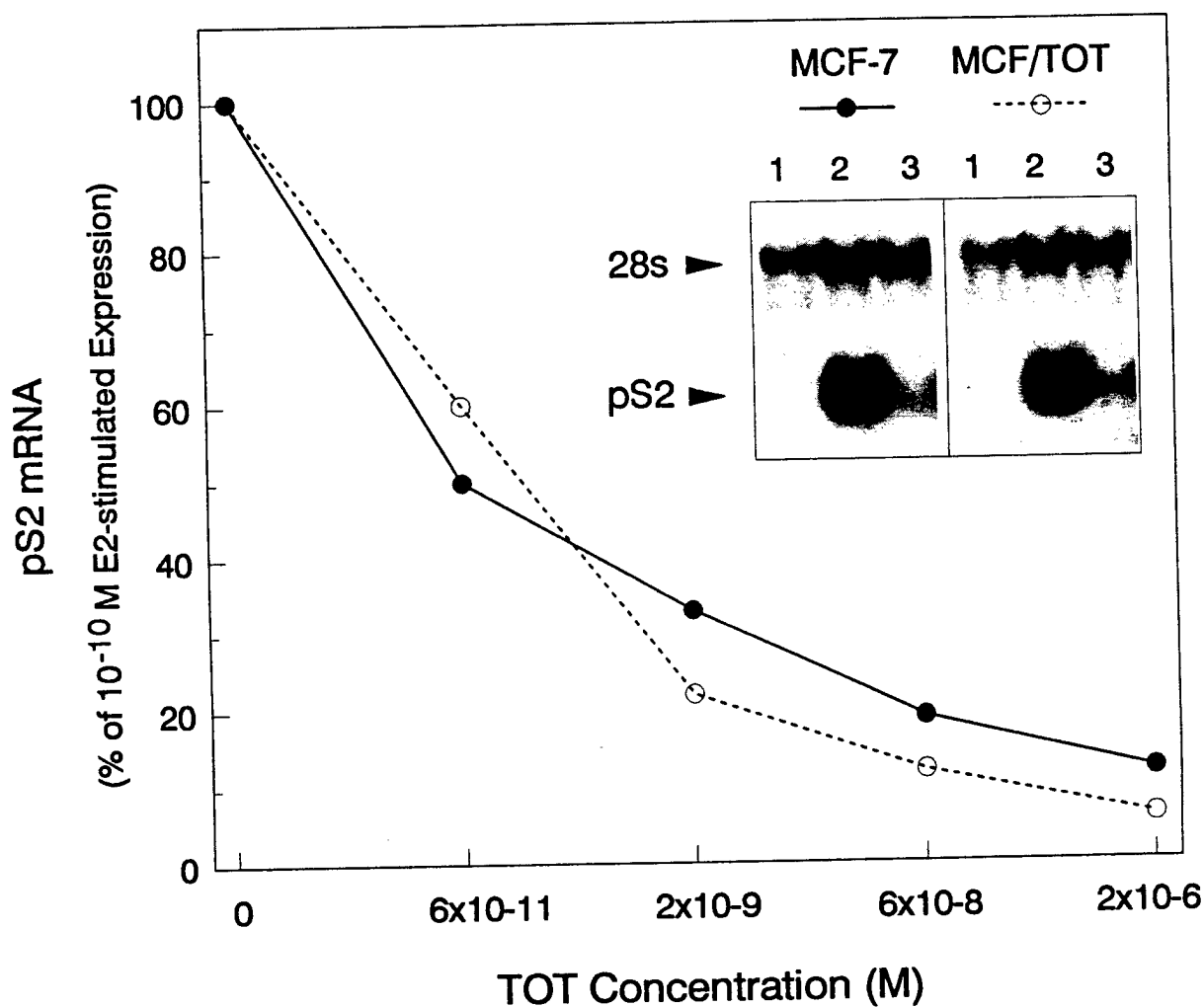


Fig. 4. Antagonism of E₂-stimulated pS2 mRNA expression by TOT. pS2 mRNA expression was analysed by Northern blot analysis of 20 µg of total RNA. Near confluent cell monolayers were treated with the ligands indicated for 12 h. Inset, autoradiogram of pS2 mRNA induction; lane 1, vehicle alone control; lane 2, 10⁻¹⁰ M E₂; lane 3, 10⁻⁶ M TOT.

(inset), pS2 was markedly stimulated by E_2 , but showed no stimulation by TOT in either cell line. Therefore, TOT is not universally seen as an estrogen agonist for all responses in the MCF/TOT cells.

Additional markers of estrogen and antiestrogen responsiveness: regulation of progesterone receptor (PgR) and transactivation of an estrogen-responsive gene construct

Expression of PgR is known to be under tight estrogen regulation. In parent MCF-7 cells, treatment with 10^{-10} M E_2 resulted in a four-fold increase in PgR content (Table 1A). In contrast, treatment of MCF/TOT cells with E_2 had no significant effect on PgR level (26.8 ± 2.2 vs. 15.3 ± 5.2 , respectively, $P > 0.05$). This was despite the presence of significant levels of ER in MCF/TOT cells, about half that present in the parent cell line (Fig. 5). A weak agonist effect of TOT was observed in parent MCF-7 cells in terms of PgR induction, but interestingly, TOT, like

E_2 , had no significant effect on PgR in the MCF/TOT subline (7.8 ± 2.2 vs. 15.3 ± 5.2 fmol 3H -R5020 bound/ 10^6 cells, respectively, $P > 0.05$). Both proliferation and PgR induction thus demonstrated altered regulation by E_2 and antiestrogen in MCF/TOT cells.

We also examined E_2 and antiestrogen responsiveness using another end-point, namely a transiently transfected estrogen-responsive gene construct containing a consensus estrogen response element (ERE) linked to a thymidine kinase (tk) promoter and the chloramphenicol acetyltransferase (CAT) gene (ERE-tk-CAT). In contrast to the loss of estrogen responsiveness of PgR in MCF/TOT cells, the transfected estrogen-responsive gene behaved similarly in parent MCF-7 and in MCF/TOT cells in terms of responsiveness to estrogen and antiestrogens. We observed comparable fold inductions of ERE-tk-CAT activity with 10^{-9} M E_2 in parent MCF-7 and MCF/TOT cells (Table 1B). TOT treatment did not significantly increase CAT activity in either subline, but it did substantially reverse the E_2 -stimulated CAT activity. The response of ERE-tk-CAT was thus similar to that for induction of pS2 mRNA by estrogen and antiestrogen in that responses to these ligands were not altered in the MCF/TOT subline as compared to the parent MCF-7 cells.

Table 1. Markers of estrogen and antiestrogen responsiveness: regulation of progesterone receptor and transactivation of an estrogen-responsive gene construct

A	fmol 3H -R5020 bound/ 10^6 cells	
	Parent MCF-7	MCF/TOT
Control vehicle	9.1 ± 3.3	15.3 ± 5.2
10^{-10} M E_2	$43.4 \pm 3.2^*$	26.8 ± 2.2
10^{-6} M TOT	$26.8 \pm 5.2^*$	7.8 ± 2.2

B	Fold change in ERE-tk-CAT activity	
	Parent MCF-7	MCF/TOT
Control vehicle	1.0 ± 0.2	1.0 ± 0.3
10^{-9} M E_2	$11.5 \pm 2.1^*$	$8.2 \pm 1.0^*$
10^{-6} M TOT	2.1 ± 0.5	0.8 ± 1.0
E_2 + TOT	$3.1 \pm 0.5^*$	2.3 ± 1.0
E_2 + ICI 164,384	0.5 ± 0.7	0.9 ± 0.7

A, Basal and stimulated progesterone receptor content was determined by binding of the progestin, [3H]R5020, by whole cell hormone binding assay after 4 days treatment with ethanol vehicle control, 10^{-10} M E_2 or 10^{-6} M TOT. Values are the mean \pm SEM of triplicate flasks from two experiments (*value significantly different from the control vehicle treated cells at $P < 0.05$ by Student's *t*-test). B, Transactivation of ERE-tk-CAT, a reporter plasmid containing a consensus estrogen response element linked to the Herpes simplex virus thymidine kinase promoter and the CAT reporter gene. ERE-tk-CAT (3 μ g) was transiently cotransfected along with an internal control plasmid containing the lac-Z gene, and cells were treated with the ligands indicated for 24 h. The calculated fold increase in the CAT activity of each group was normalized for the β -galactosidase activity. Values are expressed as the mean \pm SEM of at least three experiments (*value significantly different from the control vehicle treated cells at $P < 0.05$ by Student's *t*-test).

Estrogen receptor content and regulation in MCF-7 and MCF/TOT cells

Estrogen receptor (ER) content was determined by whole cell binding assay and Western blot analysis. Parent MCF-7 cells contained 59.2 ± 4.6 fmol ER/ 10^6 cells (Fig. 5) and this level was stable throughout the time period of these experiments (data not shown). The MCF/TOT subline contained reduced levels of ER (28.7 ± 2.4 fmol ER/ 10^6 cells) at 50 weeks of maintenance in TOT (Fig. 5). This level of ER was maintained at 75 and 125 weeks of culture in TOT (34.1 ± 1.1 and 30.5 ± 1.3 fmol ER/ 10^6 cells, respectively). A comparable decrease in ER protein level in MCF/TOT cells was also observed when analysed by Western blot analysis ($37 \pm 6\%$ of parental level; Fig. 5).

We also used Western blot analyses to assess the ability of several agents to modulate the level of the ER protein. In both the parent MCF-7 cells and MCF/TOT cells, treatment with E_2 resulted in a marked (ca. 60%) decrease in ER protein level, whereas TOT treatment did not affect ER protein level or increased it slightly, and cotreatment of either subline with TOT prevented the decrease in ER protein content induced by treatment with E_2 alone (Fig. 5). Similar to E_2 , treatment with retinoic acid (10^{-6} M) markedly decreased the ER level in both MCF-7 and MCF/TOT cells, and cotreatment with TOT prevented the reduction in ER seen in response to E_2 or retinoic acid treatment. ER level thus showed

	Parent MCF-7								MCF/TOT							
Fmol ³ HE ₂ Bound/10 ⁶ Cells	59.7 ± 4.6								28.7 ± 2.4							
Relative ER Immunoreactivity: Basal ER Content	100 ± 4%								37 ± 6%							
66 kD →																
10 ⁻⁹ M E ₂	-	+	-	-	+	+	-	-	+	-	-	+	+	-		
10 ⁻⁶ M TOT	-	-	+	-	+	-	+	-	-	+	-	+	-	+		
10 ⁻⁶ M Retinoic acid	-	-	-	+	-	+	+	-	-	-	+	-	+	+		

Fig. 5. Estrogen receptor (ER) content and effects of E_2 , TOT and retinoic acid on ER levels in parent MCF-7 and MCF/TOT cells. Estrogen receptor content was determined by whole-cell hormone binding assay and Western blot analysis. For the whole-cell binding assay, cells in T25 flasks were incubated with 10 nM [^3H] E_2 in the absence or presence of a 100-fold excess of unlabelled ligand at 37°C for 40 min ($n = 3$; mean \pm SEM). To measure immunoreactive ER, fractionated cellular protein was isolated from subconfluent T75 flasks treated with the indicated ligands for 24 h, as described in Materials and methods. ER protein was detected by binding of the ER-specific monoclonal antibodies H226 and D547. Detection of ER with the ER-specific antibody, H222 (with an exon 7 epitope), gave the same relative levels for the 66 kDa ER protein.

the same regulation by E_2 , TOT and retinoic acid in parental MCF-7 and MCF/TOT cells.

Reversibility of the TOT growth-stimulated phenotype of MCF/TOT cells

To test whether the altered growth phenotype of the MCF/TOT cells was reversible, we removed TOT from the growth medium for a period of 16 weeks (Fig. 6, panel C) and compared growth response with that of the parent MCF-7 (Fig. 6, panel A) and MCF/TOT cells (Fig. 6, panel B). As a modification, we also generated another TOT-withdrawn subline which received high levels of E_2 (10^{-8} M) simultaneously with the TOT withdrawal for 16 weeks (Fig. 6, panel D). Interestingly, whereas the TOT-withdrawn subline was no longer growth stimulated by TOT, it did not revert to the TOT growth-inhibited phenotype of the parent MCF-7 cells (Fig. 6, panel A). Rather, this subline was refractory to the effects of 10^{-6} M TOT (Fig. 6, panel C; $104 \pm 3\%$ of control values). Similar results were obtained with the TOT-withdrawn, E_2 -supplemented subline ($111 \pm 9\%$ of control values). The TOT-withdrawn subline also exhibited a partial return to the relatively high ER levels of the parent MCF-7 cells (46.4 ± 0.3 vs. 59.2 ± 4.6 fmol ER/ 10^6 cells, respectively) at 16 weeks of TOT deprivation. At 24 weeks of TOT

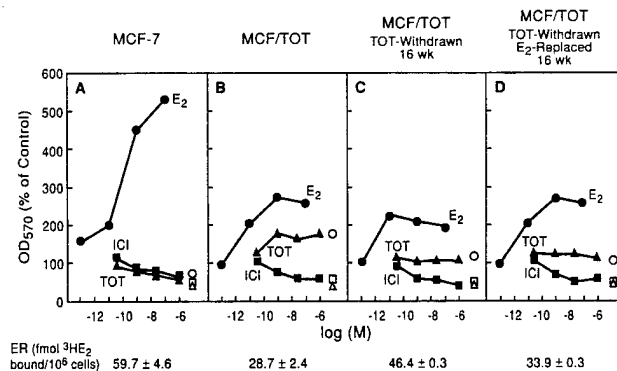


Fig. 6. Reversibility of the TOT growth-stimulated phenotype of the MCF/TOT cells. MCF/TOT cells were cultured in the absence of TOT with or without supplementation with 10^{-8} M E_2 for 16 weeks (panels C and D) and growth responses were compared with those of the parental MCF-7 (panel A) and MCF/TOT cells (panel B). Growth responsiveness to E_2 , TOT and ICI 164,384, alone or in combination, was determined by MTT assay from quadruplicate wells. Closed circle, E_2 ; closed triangle, TOT; closed square, ICI 164,384; open circle, 10^{-9} M E_2 + 10^{-6} M TOT; open triangle, 10^{-9} M E_2 + 2×10^{-6} M ICI 164,384; open square, 10^{-6} M TOT + 2×10^{-6} M ICI 164,384. Values are expressed as percentages of vehicle-treated control wells. Standard errors were less than 10% and are not shown. Estrogen receptor content was determined by whole-cell hormone binding assay ($n = 3$; mean \pm SEM).

deprivation, there was no change in the proliferative profile of the sublines compared to the 16 week TOT-withdrawn cells; both were moderately growth stimulated by E_2 , growth inhibited by ICI 164,384 and refractory to TOT (data not shown).

Estrogen receptor sequence analysis

To assess if alterations in ligand response in the MCF/TOT cells might be due to mutation of the ER, we amplified and sequenced a 1 kb region of the ER encompassing the hormone binding domain. Polymerase chain reaction yielded three cDNA products, which by direct sequence analysis were determined to be the wild type, exon 5 deletion variant ($\Delta E5$) and the exon 7 deletion variant ($\Delta E7$). The presence of these variants in breast cancers has previously been described [25, 26]. Dideoxy sequence analysis failed to reveal point mutations in the ERs from parental MCF-7 or MCF/TOT cells.

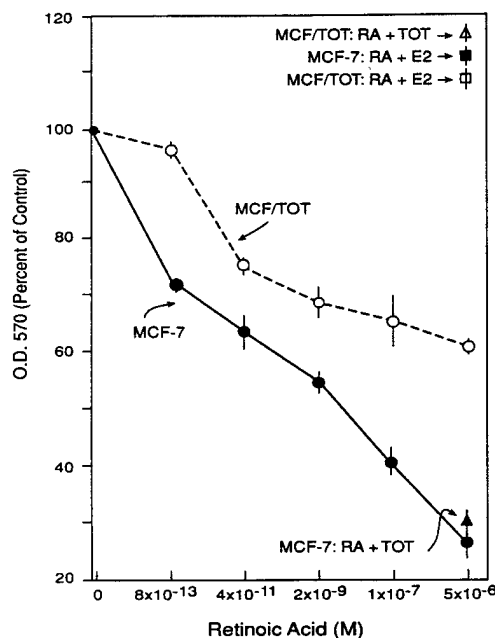


Fig. 7. Decreased responsiveness of MCF/TOT cells to the growth-inhibitory effects of retinoic acid. Cells were seeded at 2000 cells/well in 96-well plates in quadruplicate and treated with the indicated concentrations of retinoic acid for 6 days, with a media change after 3 days. Growth inhibition by retinoic acid was determined by MTT assay. The solid and open markers represent the parent MCF-7 and MCF/TOT cells, respectively; circles, retinoic acid treatment alone; squares, 5×10^{-6} M retinoic acid + 10^{-9} M E_2 ; triangles, 5×10^{-6} M retinoic acid + 10^{-6} M TOT. Values are expressed as the percentage of vehicle-treated control wells. Values for the retinoic acid dose-response curve represent the mean \pm SEM of three separate experiments. Values for cotreatment with retinoic acid plus E_2 or TOT represent the mean \pm range of two separate experiments.

Decreased responsiveness of MCF/TOT cells to the growth-inhibitory effects of retinoic acid

Retinoic acid analogues have been shown to inhibit the growth of a number of cancer cell lines, including MCF-7 cells [27]. To determine whether TOT-maintained MCF-7 cells differed in sensitivity to retinoic acid, we performed the dose-response growth study shown in Fig. 7. Parent MCF-7 cells were strongly growth inhibited by retinoic acid. Some growth suppression was observed even at very low concentrations of retinoic acid (8×10^{-13} M), and a growth suppression of approximately 75% was observed in MCF-7 cells at the highest concentration tested, 5×10^{-6} M retinoic acid. MCF/TOT cells were also sensitive to the growth suppressive effects of retinoic acid, albeit to a much lesser extent. MCF/TOT cells exhibited only $43 \pm 2\%$ growth suppression at 5×10^{-6} M retinoic acid. Cotreatment with retinoic acid and E_2 reversed the growth-suppressive effects of treatment with retinoic acid alone in both sublines (Fig. 7). Interestingly, whereas cotreatment with retinoic acid and TOT had no additional suppressive effect in parent MCF-7 cells (Fig. 7, filled triangle), TOT fully reversed the growth suppression by retinoic acid (Fig. 7, open triangle), indicating that TOT was acting as an agonist (stimulator) like E_2 , in the MCF/TOT cells.

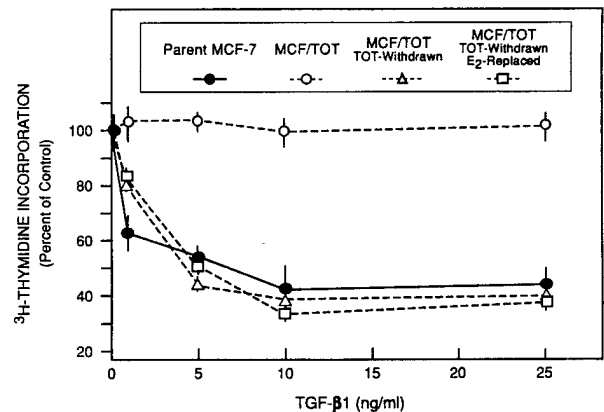


Fig. 8. Loss of growth inhibition by MCF/TOT cells in response to exogenous TGF- β 1. Deprivation of TOT from MCF/TOT cells for 16 weeks, where indicated, was performed as described in the Materials and methods section. Cells were seeded at 2000 cells/well in triplicate in 24-well dishes. Two days later the wells were washed in serum-free media and then treated with TGF- β 1 with or without 10^{-6} M TOT. After 4 days, the cells were incubated with 0.5μ Ci [3 H]thymidine at 37°C for 2 h. Incorporated [3 H]thymidine was determined as described in Materials and methods. Basal [3 H]thymidine incorporation rates were comparable between the two sublines. Treatment with TGF- β 1 in serum-supplemented or in serum-free IMEM yielded comparable results, as did measurement of cell number by MTT assay. Values are expressed as a percentage of vehicle-treated control wells ($n = 3$; SEM).

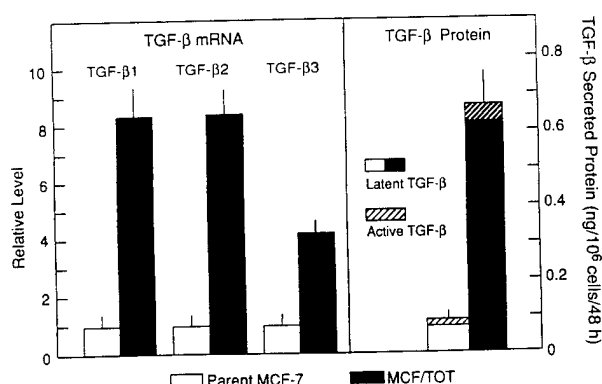


Fig. 9. Elevated TGF- β expression in MCF/TOT cells. TGF- β 1, β 2 and β 3 mRNA expression in near-confluent cell cultures was determined by ribonuclease protection assay of 10 μ g of total RNA, and normalized against human acidic phosphoprotein PO (36B4) as an internal control. RNase protection assays were quantitated by densitometric analyses of autoradiograms, as described in Materials and methods. Values represent the average and range of two experiments. Total and percentage active secreted TGF β protein were determined from duplicate conditioned media collections by inhibition of [3 H]thymidine incorporation in Mv 1 Lu cells. Values represent the mean and range from the two separate experiments.

Loss of growth suppression by exogenous TGF- β 1 in MCF/TOT cells

TGF- β 1 is of interest due to its ability to inhibit the growth of human breast cancer cells [9]. Treatment with exogenous TGF- β 1 resulted in dose-dependent decreases in [3 H]thymidine incorporation in parent MCF-7 cells (Fig. 8). An inhibition of 40% was observed at 1 ng/ml TGF- β 1, and a maximal in-

hibition of approximately 60% was observed at 5 or 10 ng/ml TGF- β 1. Further suppression of growth was accomplished by cotreatment with TOT which resulted in an additional suppression of $20 \pm 3\%$ (data not shown). In contrast, [3 H]thymidine incorporation of MCF/TOT cells was unaffected by treatment with exogenous TGF- β 1, even at 25 ng/ml. Sensitivity to TGF- β 1 was re-established upon withdrawal of TOT from MCF/TOT cells. Removal of TOT from MCF/TOT cells for 16 weeks, either with or without supplementation with E_2 , returned TGF- β 1 sensitivity to that observed in the parent MCF-7 cells (Fig. 8).

Production of TGF- β mRNA and protein in MCF-7 and MCF/TOT cells

TGF- β mRNA level was monitored in parent MCF-7 and MCF/TOT cells by ribonuclease protection assay. As shown in Fig. 9, MCF/TOT cells expressed approximately eight-fold elevated levels of TGF- β 1 and TGF- β 2 mRNA, and approximately four-fold elevated levels of TGF- β 3 mRNA, as compared to parent MCF-7 cells. The levels of bioactive TGF- β protein increased proportionally, as determined by a mink lung cell bioassay. No substantial changes in the proportion of latent and active secreted TGF- β were observed (Fig. 9). Therefore, MCF/TOT cells which no longer responded to the growth-regulating effects of exogenous TGF- β 1 (Fig. 8) secreted elevated levels of TGF- β protein. We next sought to examine if the MCF/TOT cells showed alterations in TGF- β receptor expression.

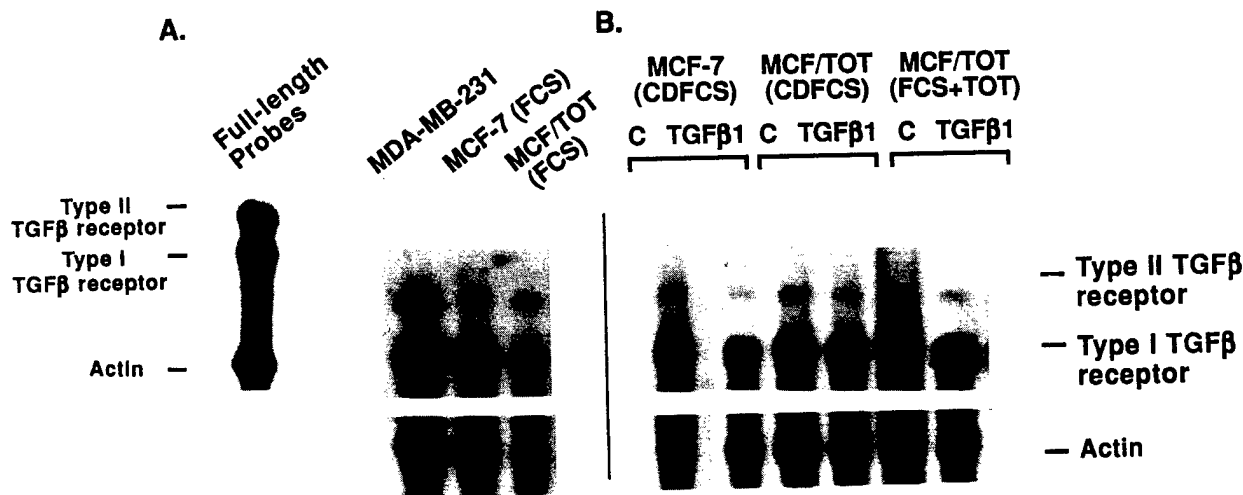


Fig. 10. Expression and ligand-induced regulation of type I and II TGF- β receptors. Total RNA from MCF-7 and MCF/TOT cells was isolated from subconfluent monolayers grown in 5% FCS MEM, with 10^{-6} M TOT where indicated, or in 5% CDFCS IMEM. Cells were treated without (C, control) or with 10 ng/ml TGF- β 1 for 8 h. Thirty micrograms total RNA was hybridized with a 300 bp riboprobe for TGF- β type I receptor (260 bp protected fragment) and a 360 bp riboprobe for TGF- β type II receptor (260 bp protected fragment) and a 300 bp riboprobe for human β -actin (125 bp protected fragment), used as an internal control. RNase protection assays were performed and quantitated as described in Fig. 9 and the Materials and methods section. For comparison, the levels of type I and II TGF- β receptors in MDA-MB-231 breast cancer cells are shown.

Expression and ligand-induced regulation of type I and II TGF- β receptor mRNAs and assessment of TGF- β 1 binding

Since TGF- β signals through a heteromeric complex of the type I and II TGF- β /activin receptors which possess serine-threonine kinase activity [28], we measured expression of type I and II TGF- β receptor mRNAs by ribonuclease protection assay (Fig. 10). There were no significant changes in the levels of these receptor mRNAs between the parent MCF-7 and MCF/TOT cells when lanes were normalized for the amount of RNA loaded. Furthermore, neither treatment with TGF- β 1 for 8 h, nor transfer from full serum to steroid-depleted serum, influenced expression of these mRNAs. These results show that the loss of sensitivity of the MCF/TOT cells to the growth-inhibitory effects of exogenous TGF- β can not be attributed to loss of expression of type I or II TGF- β receptors. We also performed [125 I]TGF- β 1 binding assays to confirm that the receptor moieties present were functionally able to bind exogenous TGF- β 1. We found 282 ± 30 ($n = 3$) [125 I]TGF- β 1 binding sites/cell in the parent MCF-7 cells. The MCF/TOT cells showed an approximate three-fold increase in the number of TGF- β 1 binding sites per cell (949 ± 102 , $P < 0.05$). Therefore, the loss of growth-inhibitory response to exogenous TGF- β 1 by MCF/TOT cells is not due to a decrease in TGF- β 1 binding sites.

DISCUSSION

This report describes a new subline of MCF-7 cells which, in response to long-term exposure to TOT, developed resistance to the growth-inhibitory effects of this antiestrogen and also altered sensitivity to the growth-suppressive effects of exogenous TGF- β 1 and retinoic acid. Furthermore, the weak stimulation of MCF/TOT cell proliferation by TOT implies that growth resistance in these cells really corresponds to a weak growth stimulation by this agent. Interestingly, these MCF/TOT cells were still responsive to suppression by the pure antiestrogens ICI 164,384 and ICI 182,740, implying that these two categories of antiestrogens must act, at least in part, by somewhat different mechanisms. Although one proposed mechanism of antiestrogen resistance is loss or mutation of estrogen receptor [29–32], our observation that the phenotype of the MCF/TOT cells is at least partly reversible following withdrawal from TOT implies a non-mutational change in these cells, consistent with our observation that ER in the parental and MCF/TOT cells had identical hormone-binding domains, as determined by DNA sequencing analysis.

Response-specific antiestrogen resistance

Whereas tamoxifen is associated with growth inhibition of breast tumors, it is also a cell- and promoter-dependent agonist. Tamoxifen shows tissue- and gene-specific estrogen-like effects, being a good estrogen agonist in bone and uterine cells and a good stimulator of some, but not all, estrogen-regulated genes [7, 33]. The ER is now known to interact with multiple proteins, termed coactivators and corepressors (reviewed in [34]), that contact different regions of the ER and can influence ER transcriptional activity greatly. Differences in the interaction of antiestrogen-ER complexes with coactivators and corepressors in different cells and at different gene sites could account for the cell- and gene-selective actions of antiestrogens in parental ER-positive breast cancer cells and in our breast cancer cells selected for resistance to growth suppression by TOT. It is perhaps to be expected, as we have observed in the present studies, that the alteration in TOT-response profile of MCF/TOT cells varied with the end-point monitored. Whereas TOT behaved agonistically in terms of proliferation in the MCF/TOT subline, there was a complete loss of its partial agonistic effects on the induction of progesterone receptor expression (Table 1). Interestingly, estrogen also failed to increase progesterone receptor in this subline, as reported in other tamoxifen-resistant breast cancer cells [32, 35]. We found, however, that the usual stimulatory and inhibitory effects of E_2 and TOT, respectively, were maintained in terms of regulation of pS2 mRNA induction and ERE-tk-CAT transactivation. These results demonstrate that loss of TOT growth inhibition is not synonymous with a global loss of responsiveness to TOT. Other MCF-7 cell variants which were tamoxifen-stimulated in terms of growth also did not exhibit corresponding tamoxifen stimulation of the estrogen-regulated mRNAs pNR-1, -2, -25, and cathepsin-D [36].

In the present work, the growth of MCF/TOT cells was dramatically suppressed by treatment with the pure antiestrogen, ICI 164,384, and this antiestrogen antagonized the effects of either E_2 or TOT on growth and gene regulation in MCF/TOT cells. ICI 164,384 has been shown to block ER action by accelerating ER degradation [37, 38] as well as inefficiently promoting transcription activation [38]. Unlike ICI 164,384, TOT treatment does not decrease ER protein content (Fig. 5; and [38]). These results, as well as the observed beneficial response to the ICI 164,384-related pure antiestrogen ICI 182,780 in tamoxifen-resistant breast cancers in women [39] and nude mouse tumor models [40, 41], support the potential clinical use of ICI 164,384-type antiestrogens in the advent of tamoxifen resistance.

Structure of the estrogen receptor

Whereas it seems plausible that mutations in the ER gene could affect ligand interpretation by the ER, our finding that TOT-stimulated growth in MCF/TOT cells is partly reversible upon withdrawal of TOT for a period of 16 weeks suggests that a readily modifiable process, rather than a mutational event, is responsible for the antiestrogen insensitivity. Alternate splicing of the ER mRNA into receptor species with different functions would allow for modulation of the receptor protein, without gene mutation. A number of ER variant mRNAs are expressed in breast neoplasms and some of these variants have been found to possess either constitutively active or inhibitory receptor activity [25, 42].

Our analysis of the nucleotide sequence of the hormone-binding domain of the ER revealed the presence of wild-type and exon 5 and exon 7 deletion variants, but failed to detect any mutations or other splicing variants in the parent MCF-7 and MCF/TOT sublines. Analysis of the ERs of other hormone-resistant sublines of MCF-7 or T47D human breast cancer cells by RNase protection mapping [43] or PCR amplification [44] also failed to detect variants or mutants of the ER. Recently, Karnik *et al.* [45] screened 20 tamoxifen-sensitive and 20 tamoxifen-resistant human breast tumors by single-strand conformation polymorphism, and found ER mutations were neither frequent nor correlated with an antiestrogen-resistant phenotype. The altered hormonal responsiveness seen in MCF/TOT cells is thus unlikely to be due to mutational change in the ER.

Cross-talk with retinoids and transforming growth factor- β in the antiestrogen resistance of MCF/TOT cells

The antiestrogenic character of the retinoids has implicated them as candidates for combination palliative therapy in ER-containing breast cancers. We found that our MCF/TOT cells exhibited decreased sensitivity to retinoic acid. This may be explained by the fact that retinoids, which have been shown to modulate estrogenic regulation of a number of mRNAs, including those for pS2 and the growth-stimulator TGF α [9], are thought to exert their growth-inhibitory effects through the ER as well as their own receptors [10, 27, 46]. Therefore, the reduced retinoic acid-induced growth suppression we observed could be, at least in part, due to the reduced levels of ER present in the MCF/TOT subline as compared to parent MCF-7 cells. This would be consistent with recent observations that the introduction of ER into ER-negative breast cancer cells re-establishes retinoic acid growth inhibition [10].

We examined TGF- β production and TGF- β receptors in our parental and MCF/TOT cells because expression of TGF- β is known to be significantly influenced by sex steroid hormones [47–50].

Because TGF- β 1 was a good growth inhibitor in our parental MCF-7 cells (Fig. 8), TGF- β resistance might thwart the suppressive, beneficial actions of tamoxifen. We observed that the MCF/TOT subline was resistant to the growth-inhibitory effects of exogenous TGF- β 1 and that this insensitivity to added TGF- β 1 was reversible following withdrawal of TOT. We also failed to observe a decrease in the expression of type I or II TGF- β receptor mRNAs or a decrease in the binding of [125 I]TGF- β 1 in MCF/TOT cells. The TGF- β receptor system is highly complex, however, and includes at least one other characterized protein, the type III TGF- β receptor, and numerous receptors with TGF- β cross-reactivity [28] which were not evaluated in the present work.

Of note, MCF/TOT cells showed elevated production of TGF- β s. The cells contained eight times more TGF- β 1 and TGF- β 2 mRNAs and four times more TGF- β 3 mRNA. They secreted three times more TGF- β bioactive protein and eight times more total (latent plus active) TGF- β protein than parental MCF-7 cells. Therefore, we do not know if their insensitivity to added TGF- β 1 was due to the high level of TGF- β production possibly resulting in the generation of maximum autocrine TGF- β activity. We think this is unlikely, however, because it is worth noting that MCF/TOT cells grow very quickly (ca. 1.6 day doubling time) in the presence of TOT and therefore are not being growth suppressed by the TGF- β s either being made and secreted by the cells, or by the TGF- β 1 we added exogenously. In addition, we previously reported that short-term estrogen-deprived MCF-7 cells contained 10 times more TGF- β 1 mRNA, eight times more TGF- β 2 mRNA and five times more TGF- β 3 mRNA, and secreted four times more bioactive TGF- β and three times more total (active plus latent) TGF- β than parental MCF-7 cells, yet these cells still showed normal, i.e. full, sensitivity to growth suppression by added TGF- β 1 [20]. More detailed analyses of the TGF- β pathway in the MCF/TOT cells will be needed to understand the changes induced by antiestrogen exposure fully.

Our findings highlight the response-specific nature of antiestrogen resistance in breast cancer cells. To our knowledge, this is the first study to compare responses to antiestrogens and to the growth-inhibitory factors retinoic acid and TGF- β in breast cancer cells selected for resistance to tamoxifen. The reduced sensitivity to these agents in the MCF/TOT cells, and the restoration of responsiveness to these agents after TOT withdrawal suggests a possible commonality of components or pathways in their regulation of proliferation of these human breast cancer cells. Our findings also indicate that one mechanism of antiestrogen resistance, as seen in MCF/TOT cells, may involve alterations in growth factor and other hormonal pathways that affect the ER response pathway.

Acknowledgements—We gratefully acknowledge the contributions of the following individuals: Rik Derynck of the University of California at San Francisco for the TGF- β 1, - β 2 and - β 3 cDNAs; Michael Brattain of the Medical College of Ohio for the TGF- β type I and II receptor cDNAs; Pierre Chambon of INSERM, Strasbourg, France for the 36B4 cDNA; Alan Wakeling of Zeneca Pharmaceuticals, Macclesfield, England for the antiestrogens, ICI 164,384 and ICI 182,780; and Abbott Laboratories, North Chicago, IL and Geoffrey L. Greene of the University of Chicago for the ER-specific antibodies. Special thanks to Anita Roberts and Nan Roche of the National Cancer Institute for performing the TGF- β bioassay and Karen Weis, David Schodin, and Heather Herbolzheimer for assistance with the estrogen receptor sequence analysis. This work was supported by NIH grant CA18119 and US Army Grant DAMD17-94-J-4205 (to B.S.K.), and in part by Carle Foundation Cancer Research Funds (to M.E.H.).

REFERENCES

- Osborne C. K., Coronado E. B. and Robinson J. P.: Human breast cancer in the athymic mouse: cytostatic effects of long-term antiestrogen therapy. *Eur. Cancer Clin. Oncol.* **23** (1987) 1189–1196.
- Gottardis M. M. and Jordan V. C.: Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res.* **48** (1988) 5183–5187.
- Nawata H., Bronzert D. and Lippman M. E.: Isolation and characterization of a tamoxifen-resistant cell line derived from MCF-7 human breast cancer cells. *Biol. Chem.* **256** (1981) 5016–5021.
- Bronzert D. A., Greene G. L. and Lippman M. E.: Selection and characterization of a breast cancer cell line resistant to the antiestrogen LY 117018. *Endocrinology* **117** (1985) 1409–1417.
- Lykkesfeldt A. E. and Sorensen E. K.: Effect of estrogen and antiestrogens, on cell proliferation and synthesis of secreted proteins in the human breast cancer cell line MCF-7 and a tamoxifen resistant variant subline, AL-1. *Acta Oncologica* **31** (1992) 131–138.
- Brunner N., Frandsen T. L., Holst-Hansen C., Bei M., Thompson E. W., Wakeling A. E., Lippman M. E. and Clarke R.: MCF-7/LCC2: a 4-hydroxytamoxifen resistant human breast cancer variant that retains sensitivity to the steroidal antiestrogen ICI 182,780. *Cancer Res.* **53** (1993) 3229–3232.
- Katzenellenbogen B. S., Montano M., Le Goff P., Schodin D. J., Kraus W. L., Bhardwaj B. and Fujimoto N.: Antiestrogens: mechanisms and actions in target cells. *J. Steroid Biochem. Molec. Biol.* **53** (1995) 387–393.
- Arteaga C. L. and Osborne C. K.: Growth factors as mediators of estrogen/antiestrogen action in human breast cancer cells. In *Regulatory Mechanisms in Breast Cancer*, ed. M. E. Lippman and R. B. Dickson. Kluwer Academic Press, Boston, 1991, pp. 289–305.
- Fontana J. A., Nervi C., Shao Z. M. and Jetten A. M.: Retinoic acid antagonism of estrogen-responsive transforming growth factor alpha and pS2 gene expression in breast carcinoma cells. *Cancer Res.* **52** (1992) 3938–3945.
- Sheikh M. S., Shao Z. M., Chen J.-C., Hussain A., Jetten A. M. and Fontana J. A.: Estrogen receptor-negative breast cancer cells transfected with the estrogen receptor exhibit increased RAR α gene expression and sensitivity to growth inhibition by retinoic acid. *J. Cell Biochem.* **53** (1993) 394–404.
- Boettger-Tong H. L. and Stancel M.: Retinoic acid inhibits estrogen-induced uterine stromal and myometrial cell proliferation. *Endocrinology* **136** (1995) 2975–2983.
- Roberts A. B. and Sporn M. B.: Physiological actions and clinical applications of transforming growth factor β (TGF- β). *Growth Factors* **8** (1993) 1–9.
- Thompson A. M., Kerr D. J. and Steel C. M.: Transforming growth factor β 1 is implicated in the failure of tamoxifen therapy in human breast cancer. *Br. J. Cancer* **63** (1991) 609–614.
- Knabbe C., Lippman M. E., Wakefield L. M., Flanders K. C., Kasid A., Derynck R. and Dickson R. B.: Evidence that transforming growth factor- β is a hormonally regulated negative growth factor in human breast cancer cells. *Cell* **48** (1987) 417–428.
- Knabbe C., Zugmaier C., Schmahl M., Dietel M., Lippman M. E. and Dickson R. B.: Induction of transforming growth factor P by the antiestrogens droloxifene, tamoxifen, and toremifene in MCF-7 cells. *Am. J. Clin. Oncol. (CCT)* **14** (Suppl. 2) (1991) S15–S20.
- Manning A. M., Williams A. C., Game S. M. and Paraskeva C.: Differential sensitivity of human colonic adenoma and carcinoma cells to transforming growth factor- β : conversion of an adenoma cell line to a tumorigenic phenotype is accompanied by a reduced response to the inhibitory effects of TGF- β . *Oncogene* **6** (1991) 1471–1476.
- Hirte H. W.: A quantitative colony-forming bioassay for transforming growth factor- β using a calorimetric endpoint. *Proc. Am. Ass. Cancer Res.* **34** (1993) 1429.
- Katzenellenbogen B. S. and Norman M. J.: Multihormonal regulation of the progesterone receptor in MCF-7 human breast cancer cells: interrelationships among insulin/insulin-like growth factor-I, serum, and estrogen. *Endocrinology* **126** (1990) 891–898.
- Read L. D., Greene G. L. and Katzenellenbogen B. S.: Regulation of estrogen receptor messenger ribonucleic acid and protein levels in human breast cancer cell lines by sex steroid hormones, their antagonists and growth factors. *Molec. Endocr.* **3** (1989) 295–304.
- Herman M. E. and Katzenellenbogen B. S.: Alterations in transforming growth factor α and β production and cell responsiveness during the progression of MCF-7 human breast cancer cells to estrogen-autonomous growth. *Cancer Res.* **54** (1994) 5867–5874.
- Cho H., Ng P. A. and Katzenellenbogen B. S.: Differential regulation of gene expression by estrogen in estrogen-growth-independent and -dependent MCF-7 human breast cancer cell sublines. *Molec. Endocr.* **5** (1991) 1323–1330.
- Newton C. R., Kalsheker N., Graham A., Powell S., Gammack A., Riley J. and Markham A. F.: Diagnosis of alpha1-antitrypsin deficiency by enzymatic amplification of human genomic DNA and direct sequencing of polymerase chain reaction products. *Nucl. Acids Res.* **16** (1988) 8223–8243.
- Berthois Y., Katzenellenbogen J. A. and Katzenellenbogen B. S.: Phenol red in tissue culture media is a weak estrogen: implications concerning the study of estrogen-responsive cells in culture. *Proc. Natn. Acad. Sci. U.S.A.* **83** (1986) 2496–2500.
- Brown A. M. C., Jeltsch J. M., Roberts M. and Chambon P.: Activation of pS2 gene transcription is a primary response to estrogen in the human breast cancer cell line MCF-7. *Proc. Natn. Acad. Sci. U.S.A.* **81** (1984) 6344–6348.
- McGuire W. L., Chamness G. C. and Fuqua S. A. W.: Estrogen receptor variants in clinical breast cancer. *Molec. Endocr.* **5** (1991) 1571–1577.
- Miksicsek R. J., Lei Y. and Wang Y.: Exon skipping gives rise to alternatively spliced forms of the estrogen receptor in breast tumor cells. *Breast Cancer Res. Treat.* **26** (1993) 163–174.
- Ueda H., Takenawa T., Millan J. C., Gesell M. S. and Brandes D.: The effects of retinoids on proliferative capacities and macromolecular synthesis in human breast cancer MCF-7 cells. *Cancer (Phil.)* **46** (1980) 2203–2709.
- Massague J.: Receptors for the TGF- β family. *Cell* **69** (1992) 1067–1070.
- Jordan V. C.: The control of hormone-dependent breast cancer growth — are we talking about estrogen alone?. *Eur. J. Clin. Cancer Oncol.* **24** (1988) 1245–1248.
- Katzenellenbogen B. S.: Antiestrogen resistance: mechanisms by which breast cancer cells undermine the effectiveness of endocrine therapy. *J. Natn. Cancer Inst.* **83** (1991) 1434–1435.
- Touchette N.: Tamoxifen resistance in breast cancer. *J. NIH Res.* **4** (1992) 67–72.
- Wolf D. M. and Jordan V. C.: The estrogen receptor from a tamoxifen stimulated MCF-7 tumor variant contains a point mutation in the ligand binding domain. *Breast Cancer Res. Treat.* **31** (1994) 129–138.
- McDonnell D. P., Clemm D. L., Herman T., Goldman M. E., Pike J. W.: Analysis of estrogen receptor function *in vitro* reveals three distinct classes of antiestrogens. *Molec. Endocr.* **9** (1995) 659–669.

34. Katzenellenbogen J. A., O'Malley B. W. and Katzenellenbogen B. S.: Tripartite steroid hormone receptor pharmacology: interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. *Molec. Endocr.* **10** (1996) 119-131.
35. Lykkesfeldt A. E., Madsen M. W. and Briand P.: Altered expression of estrogen-regulated genes in a tamoxifen-resistant and ICI 164,384 and ICI 182,780 sensitive human breast cancer cell line, MCF-7/TAM^R-1. *Cancer Res.* **54** (1994) 1587-1595.
36. Wiseman L. R., Johnson M. D., Wakeling A. E., Lykkesfeldt A. E., May F. E. B. and Westley B. R.: Type-I IGF receptor and acquired tamoxifen resistance in oestrogen-responsive human breast cancer cells. *Eur. J. Cancer* **29A** (1993) 2256-2264.
37. Dauvois S., Danielian P. S., White R. and Parker M. G.: Antiestrogen ICI 164,384 reduces cellular estrogen receptor content by increasing its turnover. *Proc. Natn. Acad. Sci. U.S.A.* **89** (1992) 4037-4041.
38. Reese J. C. and Katzenellenbogen B. S.: Examination of the DNA-binding ability of estrogen receptor in whole cells: implications for hormone-independent transactivation and the actions of antiestrogens. *Molec. Cell. Biol.* **12** (1992) 4531-4538.
39. Howell A., DeFriend D., Robertson J., Blarney R. and Walton P.: Response to a specific antiestrogen (ICI 182780) in tamoxifen-resistant breast cancer. *Lancet* **345** (1995) 29-30.
40. Hu X. F., Veroni M., De Luise M., Wakeling A., Sutherland R., Watts C. K. W. and Zalcberg J. R.: Circumvention of tamoxifen resistance by the pure antiestrogen ICI 182,780. *Int. J. Cancer* **55** (1993) 873-876.
41. Osborne C. K., Coronado-Heinsohn E. B., Hilsenbeck S. G., McCue B. L., Wakeling A. E., McClelland R. A., Manning D. L. and Nicholson R. I.: Comparison of the effects of a pure steroidal antiestrogen with those of tamoxifen in a model of human breast cancer. *J. Natn. Cancer Inst.* **87** (1995) 746-750.
42. Fuqua S. A. W., Chamness G. C. and McGuire W. L.: Estrogen receptor mutations in breast cancer. *J. Cell. Biochem.* **51** (1993) 135-139.
43. Mullick A. and Chambon P.: Characterization of the estrogen receptor in two antiestrogen-resistant cell lines, LY2 and T47D. *Cancer Res.* **50** (1990) 333-338.
44. Jiang S. Y., Wolf D. M., Yingling J. M., Chang C. and Jordan V. C.: An estrogen receptor positive MCF-7 clone that is resistant to antiestrogens and estradiol. *Molec. Cell. Endocr.* **90** (1992) 77-86.
45. Karnik P. S., Kulkarni S., Liu X., Budd G. T. and Bukowski R. M.: Estrogen receptor mutations in tamoxifen-resistant breast cancer. *Cancer Res.* **54** (1994) 349-353.
46. Sheikh M. S., Shao Z. M., Li X. S., Dawson M., Jettan A. M., Wu S. L., Conley B. A., Garcia M., Rochefort H. and Fontana J. A.: Retinoic-resistant estrogen receptor-negative human breast carcinoma cells transfected with retinoic acid receptor-alpha acquire sensitivity to growth inhibition by retinoids. *J. Biol. Chem.* **269** (1994) 21440-21447.
47. Butta A., MacLennan K., Flanders K. C., Sacks N. P. M., Smith I., McKinna A., Dowsett M., Wakefield L. M., Sporn M. B., Baum M. and Colletta A. A.: Induction of transforming growth factor β 1 in human breast cancer *in vivo* following tamoxifen treatment. *Cancer Res.* **52** (1992) 4261-4264.
48. Colletta A. A., Wakefield L. M., Howell F. V., Van Roozendaal K. E. P., Danielpour D., Ebbs S. R., Sporn M. B. and Baum M.: Antioestrogens induce the secretion of active TGF- β from human fetal fibroblasts. *Br. J. Cancer* **62** (1990) 405-409.
49. Arteaga C. L., Coffey R. J. Jr., Dugger T. C., McCutchen C. M., Moses H. L. and Lyons R. M.: Growth stimulation of human breast cancer cells with anti-transforming growth factor beta antibodies: evidence for negative autocrine regulation by transforming growth factor beta. *Cell Growth Diff.* **1** (1990) 367-374.
50. Arrick B. A., Korc M. and Derynck R.: Differential regulation of expression of three transforming growth factor beta species in human breast cancer cell lines by estradiol. *Cancer Res.* **50** (1990) 299-303.

Katzenellenbogen, B.S., Montano, M. M., Kraus, W. L., Aronica, S. M., Fujimoto, N., and LeGoff, P. Estrogen-receptor and antiestrogen-receptor complexes: cell and promoter specific effects and interactions with second messenger signaling pathways. In: Baird et al. (Eds.), Ernst Schering Foundation Workshop 16, Organ-Selective Actions of Steroid Hormones, Springer-Verlag Publishers, Heidelberg, Germany, pp. 29-50, 1995.

2 Estrogen-Receptor and Antiestrogen-Receptor Complexes: Cell- and Promoter-Specific Effects and Interactions with Second Messenger Signaling Pathways

B. S. Katzenellenbogen, M. M. Montano, W. L. Kraus,
S. M. Aronica, N. Fujimoto, and P. LeGoff

2.1	Introduction and Overview	29
2.2	Estrogen and Antiestrogen Binding and Discrimination by the ER	31
2.3	The Carboxy-Terminal F Domain of the ER: Role in the Transcriptional Activity of the Receptor and the Effectiveness of Antiestrogens as Estrogen Antagonists in Different Target Cells	35
2.4	Cross-Talk Between the ER and Second Messenger Signaling Pathways in Cells	36
2.5	Phosphorylation of the ER	41
2.6	Antiestrogen Selectivity and Promoter Dependence in the cAMP-Dependent Signaling Pathway Involvement in Activation of the Transcriptional Activity of ERs Occupied by Antiestrogens	42
2.7	Bidirectional Cross-Talk Between Estrogen and cAMP Signaling Pathways	43
	References	45

2.1 Introduction and Overview

Estrogens regulate the differentiation, growth, and functioning of many reproductive tissues. They also exert important actions on other tissues, including bone, liver, and the cardiovascular system. Most of the actions of estrogens appear to be exerted via the estrogen receptor (ER) of target

cells, an intracellular receptor that is a member of a large superfamily of proteins that function as ligand-activated transcription factors, regulating the synthesis of specific RNAs and proteins. Because of this diversity of estrogen target tissues, much current interest focuses on trying to understand the basis for the cell and promoter context-dependent actions of estrogens and antiestrogens. Antiestrogens, which antagonize the actions of estrogens, have much potential as important therapeutic agents. Although antiestrogens bind to the ER in a manner that is competitive with estrogen, they fail to effectively activate gene transcription (Jordan and Murphy 1990; Katzenellenbogen et al. 1985; Santen et al. 1990). Two of the major challenges in studies on antiestrogens are to understand what accounts for their antagonistic effectiveness, as well as the partial agonistic effects of some antiestrogens, and to understand how one can attain tissue-selective agonist/antagonist effects of these compounds.

In order to address these issues, many of our analyses have focused in detail on the hormone-binding domain of the ER, regions E and F, since this domain of the receptor contains both hormone-binding and hormone-dependent transactivation functions of the receptor. In our attempts to understand how the ER discriminates between estrogen and antiestrogen ligands, and between different categories of antiestrogens, we have generated and analyzed variant human ERs with mutations in the ER hormone-binding domain and studied the activity of these receptors on different estrogen-responsive genes in several cell backgrounds when liganded with antiestrogenic or estrogenic ligands. These studies, and those of others, have provided consistent evidence for the promoter-specific and cell-specific actions of the estrogen-occupied and antiestrogen-occupied ER. In addition, although the binding of estrogens and antiestrogens is mutually competitive, studies with ER mutants indicate that some of the contact sites of estrogens and antiestrogens are likely different. Our recent studies reveal that the presence of the C-terminal F domain of the ER is important in the transcription activation and repression activities of antiestrogens and that it affects the magnitude of liganded ER bioactivity in a cell-specific manner (Montano et al. 1994, 1995). The influence of the F domain on the agonist/antagonist balance and potency of antiestrogens supports its specific modulatory role in the ligand-dependent interaction of ER with components of the transcription complex. These studies (Ince et al. 1993, 1995; Katzenellenbogen

et al. 1993; Pakdel and Katzenellenbogen 1992; Wrenn and Katzenellenbogen 1993, see below) have provided evidence for a regional dissociation of the hormone-binding and transcription-activation regions in the domain E of the receptor and have also shown that mutations in the hormone-binding domain and deletions of C-terminal regions result in ligand discrimination mutants, that is, receptors that are differentially altered in their ability to bind and/or mediate the actions of estrogens versus antiestrogens.

In addition, in studies described below, we have observed that protein kinase activators enhance the transcriptional activity of the ER and alter the agonist/antagonist balance of some antiestrogens, suggesting that changes in the cellular phosphorylation state should be important in determining the effectiveness of antiestrogens as estrogen antagonists. The ability of estrogens and antiestrogens to also increase cAMP levels in target cells suggests that the interaction of estrogens with second messenger signaling pathways may be bidirectional.

2.2 Estrogen and Antiestrogen Binding and Discrimination by the ER

A variety of studies (Berry et al. 1990; Fujimoto and Katzenellenbogen 1994; Pakdel et al. 1993a, b; Pakdel and Katzenellenbogen 1992; Reese and Katzenellenbogen 1991, 1992a, b; Tzukerman et al. 1994; Wrenn and Katzenellenbogen 1993) have provided strong documentation that the response of genes to estrogen and antiestrogen depend on several important factors: (a) the nature of the ER, i.e., whether it is wild type or variant; (b) the ligand; (c) the promoter; and (d) the cell context. The gene response, in addition, can be modulated by cAMP, growth factors, and agents that affect protein kinases and cell phosphorylation (Aronica and Katzenellenbogen 1991, 1993; Cho et al. 1994; Fujimoto and Katzenellenbogen 1994; Kraus et al. 1993). These factors, no doubt, account for differences in the relative agonism/antagonism of antiestrogens, for instance, tamoxifen, on different genes and in different target cells such as those in breast cancer cells versus uterine or bone cells.

Although both estrogens and antiestrogens bind within the hormone-binding domain, the association must differ because estrogen binding

activates a transcriptional enhancement function, whereas antiestrogens fully or partially fail in this role. Antiestrogens are believed to act in large measure by competing for binding to the ER and altering the conformation of the ER such that the receptor fails to effectively activate gene transcription. In addition, antiestrogens exert anti-growth factor activities via a mechanism that requires ER but is still not fully understood (Freiss and Vignon 1994). Models of antiestrogen action at the molecular level are beginning to emerge, and recent biological studies also indicate that antiestrogens fall into at least two distinct categories: antiestrogens such as tamoxifen that are mixed or partial agonists/antagonists (type I), and compounds such as ICI164,384 that are complete/pure antagonists (type II). The type I antihormone-ER complexes appear to bind as dimers to EREs; there, they block hormone-dependent transcription activation mediated by region E of the receptor, but are believed to have little or no effect on the hormone-independent transcription activation function located in region A/B of the receptor (Berry et al. 1990). Thus, they are generally partial or mixed agonist/antagonists, and their action must involve some subtle difference in ligand-receptor interaction very likely associated with the basic or polar side chain that characterizes the antagonist members of this class. In the case of the more complete antagonists such as ICI164,384, ER conformation must clearly differ from that of the estrogen-occupied ER since alteration in ER binding to DNA and reduction of the ER content of target cells appear to contribute to (Dauvois et al. 1992; Fawell et al. 1990b), but may not fully explain, the pure antagonist character of this antiestrogen (Reese and Katzenellenbogen 1992b). Of note is the fact that antiestrogens, whether steroidal or nonsteroidal, typically have a bulky side chain which is basic or polar. This side chain is important for antiestrogenic activity; removal of this side chain results in a compound which is no longer an antiestrogen and, instead, has only estrogenic activity. Therefore, interaction of this side chain with the ER must play an important role in the interpretation of the ligand as an antiestrogen.

In order to understand how the ER "sees" an antiestrogen as different from an estrogen, we have used site-directed and random chemical mutagenesis of the ER cDNA to generate ERs with selected changes in the hormone-binding domain. We have been particularly interested in identifying residues in the hormone-binding domain important for the

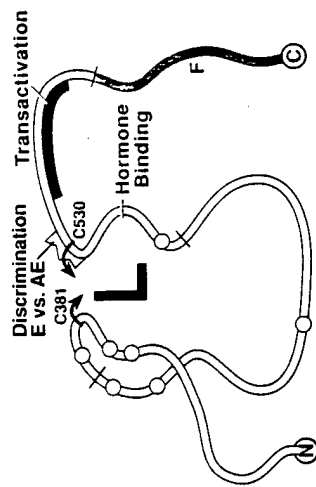


Fig. 1. "Map" of functions in the human estrogen receptor hormone-binding domain (HBD). Domain E, amino acids 302-553, is shown as is the very C-terminal domain F, amino acids 554-595. Some regions considered to be important in hormone binding, discrimination between estrogen (E) and antiestrogen (AE), and transactivation are highlighted. The ligand (L) is portrayed in a region representing the ligand binding pocket of the receptor. *Open circles* indicate amino acids in the HBD where our mutational analyses have shown mutational changes to affect the affinity or stability of hormone binding. See text for description. N, N-terminus; C, C-terminus of receptor

ligand binding of estrogen and antiestrogen and transactivation functions of the receptor and in elucidating the mechanism by which the ER differently interprets agonistic and antagonistic ligands. Our studies have indicated that selective changes near amino acid 380 and amino acids 520-530 and changes at the C-terminus of the ER result in ER ligand discrimination mutants (Montano et al. 1994, 1995; Pakdel et al. 1993b; Pakdel and Katzenellenbogen 1992). These data provide evidence that some contact sites of the receptor with estrogen and antiestrogen differ and that the conformation of the receptor with estrogen and antiestrogen must also be different as a consequence (Danielian et al. 1993; Pakdel and Katzenellenbogen 1992, references therein).

Our structure-function analysis of the hormone-binding domain of the human ER has utilized region-specific mutagenesis of the ER cDNA and phenotypic screening in yeast, followed by the analysis of interesting receptor mutants in mammalian cells (Katzenellenbogen et al. 1993; Wrenn and Katzenellenbogen 1993). A great advantage of the yeast system is that it allows the rapid screening of a library of many mutants, a situation that is not possible in mammalian cells. Our obser-

variations, as well as very important studies by Malcolm Parker and colleagues (Danielian et al. 1992, Fawell et al. 1990a), have shown a separation of the transactivation and hormone-binding functions of the ER, with amino acids critical in the transactivation function of the receptor being more C-terminal in domain E (see Fig. 1). Interestingly, some transcriptionally inactive receptors with modifications in this domain E C-terminal activation function 2 (AF-2) region of the ER have potent dominant negative activity, being able to suppress the activity of the wild-type ER in cells (Ince et al. 1993, 1995).

Since the basic or polar side chain is essential for antiestrogenic activity, and our previous studies identified cysteine 530 as the amino acid covalently labeled by affinity-labeling ligands (Harlow et al. 1989), we introduced by site-directed mutagenesis of the ER cDNA changes of specific charged residues close to C530 (Pakdel and Katzenellenbogen 1992). Interestingly, two mutants in which lysines at position 529 and 531 were changed to glutamines, so that the local charge was changed, resulted in ligand discrimination mutants. These receptors showed an approximately thirtyfold increased potency of antiestrogen in suppressing estradiol (E₂)-stimulated reporter gene activity. Interestingly, these mutant receptors had a reduced binding affinity for estrogens, but retained unaltered binding affinity for antiestrogen. These observations suggest that we are able to differentially alter estrogen and antiestrogen effectiveness by rather modest changes in the ER, and that the region near C530 is a critical one for sensing the fit of the side chain of the estrogen antagonist. Studies from the Parker Laboratory (Danielian et al. 1993) have shown that nearby residues (i.e., G525 and M521 and/or S522 in the mouse ER) are also importantly involved in conferring differential sensitivity to these two categories of ligands.

2.3 The Carboxy-Terminal F Domain of the ER: Role in the Transcriptional Activity of the Receptor and the Effectiveness of Antiestrogens as Estrogen Antagonists in Different Target Cells

Among the nuclear hormone receptors, ER is unusual in having a large C-terminal F region (42 amino acids) and its role in ER bioactivity and in the activities of antiestrogens has not been well defined. Previous studies by us and others have shown that domain F is not required for transcriptional response to E₂ (Kumar et al. 1986, 1987; Pakdel et al. 1993a), and, in addition, our studies have shown that this region does not affect the turnover rate of ER in target cells (Pakdel et al. 1993a). However, a more complete examination (Montano et al. 1994, 1995) has revealed that the presence of the F domain is important in the transcription activation and repression activities of antiestrogens, and that it affects the magnitude of liganded ER bioactivity in a cell-specific manner. Thus, in ER-negative breast cancer cells and Chinese hamster ovary (CHO) cells, E₂ stimulated equally the transcription of several estrogen-responsive promoter-reporter gene constructs with wild-type ER, and with ER lacking the carboxy-terminal F domain or ER altered in the F domain by point mutations. Of interest, the antiestrogens trans-hydroxytamoxifen and ICI164,384, which showed considerable agonistic activity on some of the reporter constructs with the wild-type ER, showed no agonistic activity with the ER lacking the F domain. In addition, the antiestrogens were more potent antagonists of E₂-stimulated transcription by the F domain-deleted ER than by wild-type ER. Interestingly, the effect of the F domain was very dependent on the cell type. In HeLa human cervical cancer cells, the F domain-deleted ER exposed to E₂ was much less effective than wild-type ER in stimulating transcription, and antiestrogens were less potent in suppressing E₂-stimulated transcription by the F domain-deleted ER. Since we find that the F domain does not appear to affect estrogen or antiestrogen ligand-binding affinity or DNA binding of the receptor, the fact that this region makes the liganded ER either more or less transcriptionally effective in different cells suggests that it plays an important role in maintaining ER conformation optimal for protein-protein interactions needed for transcriptional effectiveness. Its influence on the agonist/antagonist balance

and potency of antiestrogens further supports its specific modulatory role in the ligand-dependent interaction of ER with components of the transcription complex. Therefore, these studies (Montano et al. 1994, 1995) as well as several others by us (Cho and Katzenellenbogen 1993; Fujimoto and Katzenellenbogen 1994; Katzenellenbogen et al. 1995) and those of others have highlighted that beyond differences in estrogen and antiestrogen binding to the ER, the cell context and promoter context of the ER influences the estrogenic/antiestrogenic activity of antiestrogens.

2.4 Cross-Talk Between the ER and Second Messenger Signaling Pathways in Cells

Evidence for cross-talk between steroid hormone receptors and signal transduction pathways has been increasing. Expression of AP-1, a transcription factor of the Fos/Jun heterodimer known to mediate the protein kinase-C (PKC) pathway (Angel et al. 1987), was shown to suppress steroid hormone receptor-mediated gene expression (Schüle et al. 1990), most likely through direct protein-protein interaction between steroid receptors and these oncoproteins (Yang-Yen et al. 1990). In addition, the ovalbumin gene promoter containing a half-palindromic estrogen-responsive element (ERE) was coactivated by ER and Fos/Jun oncoproteins (Gaub et al. 1990). Thus, interaction between these oncoproteins and steroid hormone receptors resulted in cell-specific inhibitory or stimulatory effects on transcriptional activation (Gaub et al. 1990; Shemshedini et al. 1991; Strähle et al. 1988; Yang-Yen et al. 1990).

Previous studies by us and others (Aronica and Katzenellenbogen 1991; Katzenellenbogen and Norman 1990; Sumida et al. 1988; Sumida and Pasqualini 1989, 1990) documented upregulation of intracellular progesterone receptor, an estrogen-stimulated protein, by insulin-like growth factor-I (IGF-I), epidermal growth factor, phorbol ester, and cAMP in MCF-7 human breast cancer cells and uterine cells. The fact that the stimulation by these diverse agents was blocked by antiestrogen suggested that these agents were presumably acting through the ER pathway (Aronica and Katzenellenbogen 1991; Katzenellenbogen and Norman 1990; Sumida et al. 1988; Sumida and Pasqualini 1989). In

addition, the fact that protein kinase inhibitors also blocked the effects of estrogen, cAMP, and growth factors suggested the involvement of phosphorylation in these responses. We therefore undertook studies to examine directly whether activators of protein kinases can modulate transcriptional activity of the ER.

In primary cultures of uterine cells using transient transfection experiments with simple estrogen-responsive reporter genes, we examined the ability of these agents to stimulate ER-mediated gene transcription and also compared the ability of these multiple agents to alter the phosphorylation state of the endogenous uterine ER protein. The results of our study (Aronica and Katzenellenbogen 1993) indicate that E₂, IGF-I, and agents which raise intracellular cAMP are able to stimulate ER-mediated transactivation and ER phosphorylation. The fact that antiestrogen (ICI164,384) evokes a similar increase in ER phosphorylation without a similar increase in transcription activation indicates that an increased overall ER phosphorylation does not necessarily result in increased transcriptional activity. Also, the observation that transcriptional activation by the ER was nearly completely suppressed by the protein kinase inhibitors H8 and PKI, while the increase in phosphorylation was reduced by 50%–75%, indicates that the correlation between transcriptional activation and overall ER phosphorylation is not direct but does suggest that some of the effects of E₂, IGF-I and cAMP on ER-regulated transactivation are mediated through the activity of protein kinases. Our findings, demonstrating a clear effect of these agents on ER-mediated transactivation, suggest that these agents might also regulate endogenous estrogen target genes, such as that encoding the progesterone receptor, by similar cellular mechanisms.

In order to examine some of the molecular mechanisms controlling transcription of the progesterone receptor gene, we cloned the rat progesterone receptor gene 5'-region and identified two functionally distinct promoters (Kraus et al. 1993). The two distinct promoters in the rat progesterone receptor gene exhibited differential responsiveness to E₂ and to ER-dependent stimulation by cAMP. The functional differences between these two promoters may lead to altered expression of the A and B progesterone receptor isoforms and, thereby, influence cellular responsiveness to progestins (Kraus et al. 1993).

In MCF-7 human breast cancer cells and other cells, we found that activators of PKA and PKC markedly synergize with E₂ in ER-mediated

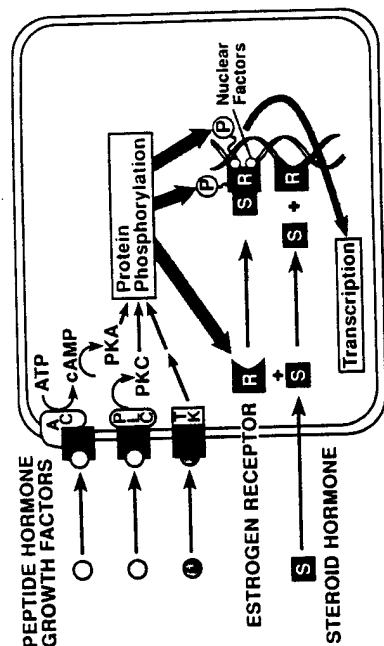


Fig. 2. Protein kinase-estrogen receptor transcriptional synergism. See text for description. AC, adenylate cyclase; PLC, phospholipase C; TK, tyrosine kinase; PKC, protein kinase C; PKA, protein kinase A; R, estrogen receptor; S, steroid hormone; P, phosphate groups on proteins

transcriptional activation and that this transcriptional synergism shows cell and promoter specificity (Cho and Katzenellenbogen 1993; Fujimoto and Katzenellenbogen 1994; Kraus et al. 1993). The synergistic stimulation of ER-mediated transcription by E₂ and protein kinase activators did not appear to result from changes in ER content or in the binding affinity of ER for ligand or the ERE DNA, but rather may be a consequence of a stabilization or facilitation of interaction of target components of the transcriptional machinery, possibly either through changes in phosphorylation of ER or other proteins important in ER-mediated transcriptional activation (Cho and Katzenellenbogen 1993).

Figure 2 shows a model indicating how we think the protein kinase-ER transcriptional synergism might occur. Agents influencing protein kinase pathways may enhance intracellular protein phosphorylation resulting in either phosphorylation of the ER itself or the phosphorylation of nuclear factors with which the receptor interacts in mediating transcription. Likewise, there is evidence that the steroid hormone itself can alter receptor conformation increasing its susceptibility to serve as a substrate for protein kinases (Ali et al. 1993; Aronica and Katzenellenbogen 1993; Denton et al. 1992; Lahooti et al. 1994; Le Goff et al. 1994). Therefore, agents which increase phosphorylation may, either

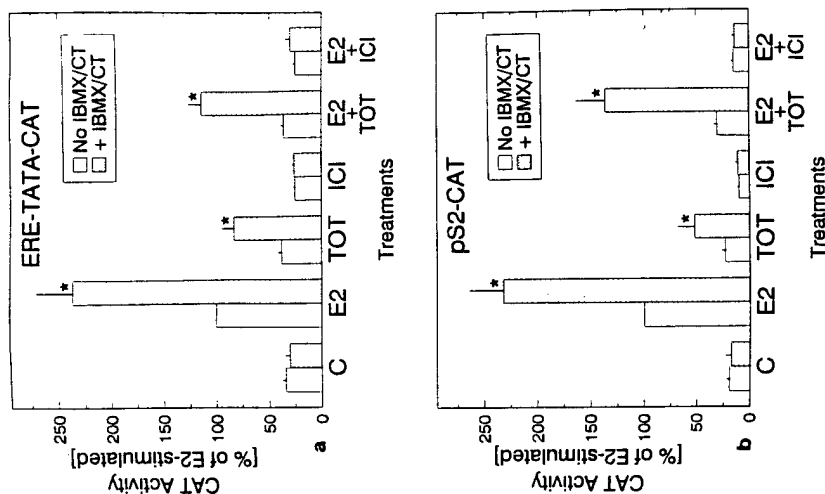


Fig. 3a,b. Effect of IBMX/CT on the ability of E₂ and antiestrogens to stimulate transactivation of ERE-TATA-CAT (a) and pS2-CAT (b), and on the ability of antiestrogens to suppress E₂-stimulated transactivation. MCF-7 cells were transfected with the indicated reporter plasmid and an internal control plasmid that expresses β -galactosidase and were treated with the agents indicated for 24 h. Each bar represents the mean \pm SEM ($n=3$ experiments). * indicates significant difference from the no IBMX/CT cells ($p < 0.05$). C, control ethanol vehicle; E₂, 10^{-9} M; TOT, *trans*-hydroxytamoxifen (10^{-6} M); ICI, ICI 164384 (10^{-6} M); IBMX, 3-isobutyl-1-methyl-xanthine (10^{-4} M); CT, cholera toxin (1 μ g/ml). (From Fujimoto and Katzenellenbogen 1994)

through phosphorylation of the ER itself or through phosphorylation of nuclear factors required for ER transcription, result in synergistic activation of ER-mediated transcription.

As shown in Fig. 3, we have compared the effects of cAMP on the transcriptional activity of E₂ liganded and antiestrogen-liganded ER complexes. We find that increasing the intracellular concentration of cAMP, or of protein kinase A catalytic subunits by transfection (Fujimoto and Katzenellenbogen 1994), activates and/or enhances the transcriptional activity of type I but not type II antiestrogen-occupied ER complexes and reduces the estrogen antagonist activity of the type I *trans*-hydroxy-tamoxifen (TOT) antiestrogen. In Fig. 3a,b, we have determined in MCF-7 human breast cancer cells the effect of cAMP on the activity of TOT, ICI164,384, and E₂ on a simple TATA promoter with one consensus ERE upstream of the CAT gene and on the more complex pS2 gene promoter and 5' flanking region (-3000 to +10) containing an imperfect ERE. The endogenous pS2 gene is regulated by E₂ in MCF-7 breast cancer cells. E₂ increased the transcription of both of these gene constructs, and treatment with isobutyl-methyl-xanthine and cholera toxin (IBMX/CT) and E₂ evoked a synergistic increase in transcription, with activity being ca. 2.5 times that of E₂ alone. Both antiestrogens (TOT and ICI) failed to stimulate transactivation of these reporter gene constructs, but in the presence of IBMX/CT, TOT gave significant stimulation of transcription (85% or 60% that of E₂ alone). ICI failed to stimulate transactivation even in the presence of IBMX/CT, and ICI fully blocked E₂ stimulation in the presence or absence of cAMP. By contrast, treatment with IBMX/CT reduced the ability of TOT to inhibit E₂ transactivation. While TOT returned E₂ stimulation down to that of the control in the absence of IBMX/CT (compare open bars E₂ vs. E₂+TOT in Fig. 3), TOT only partially suppressed the E₂ stimulation in the presence of IBMX/CT (compare stippled bars E₂ vs. E₂+TOT in Fig. 3).

Although alteration in the agonist and antagonist activity of TOT was observed with promoter-reporter constructs containing a simple TATA promoter and a more complex pS2 promoter, elevation of cAMP did not enhance the transcription by either TOT or E₂ of the reporter plasmid ERE-thymidine kinase-CAT (Fujimoto and Katzenellenbogen 1994). Thus, this phenomenon is promoter specific. Of note, cAMP and PKA catalytic subunit transfection failed to evoke transcription by the more

pure antiestrogen ICI164,384 with any of the promoter-reporter constructs tested. These findings, which document that stimulation of the PKA signaling pathway activates the agonist activity of tamoxifen-like antiestrogens, may in part explain the development of tamoxifen resistance by some ER-containing breast cancers. They also suggest that the use of antiestrogens such as ICI164,384 that fail to activate ER transcription in the presence of cAMP may prove more effective for long-term antiestrogen therapy in breast cancer.

2.5 Phosphorylation of the ER

Since our data suggested that estrogens, and agents that activate protein kinases, might influence ER transcription by altering the state of phosphorylation of the ER and/or other factors required for ER regulation of transcription, we undertook studies to examine directly the effects of these agents on ER phosphorylation. In addition, we compared the effects of the type I and type II antiestrogens on phosphorylation of the ER. E₂, the antiestrogens, *trans*-hydroxy-tamoxifen and ICI164,384, as well as PKA and PKC activators enhanced overall ER phosphorylation, and in all cases, this phosphorylation appeared to be on serine residues (Le Goff et al. 1994).

Tryptic phosphopeptide patterns of wild-type and domain A/B-deleted receptors and site-directed mutagenesis of several serines involved in known protein kinase consensus sequences allowed us to identify serine 104 and/or serine 106 and serine 118, all three being part of a serine-proline motif, as major ER phosphorylation sites. Mutation of these serines to alanines so as to eliminate the possibility of their phosphorylation resulted in an approximately 40% reduction in transactivation activity in response to E₂ while mutation of only one of these serines showed an approximately 15% decrease in activation (Le Goff et al. 1994). Of note, E₂ and antiestrogen-occupied ERs showed virtually identical two-dimensional phosphopeptide patterns, suggesting similar sites of phosphorylation. In contrast, the cAMP-stimulated phosphorylation likely occurs on different phosphorylation sites as indicated by some of our mutational studies (Le Goff et al. 1994) and this aspect remains under investigation in our laboratory.

Related studies in COS-1 cells by the Chambon laboratory (Ali et al. 1993) also identified serine 118 as being a major estrogen-regulated phosphorylation site. In MCF-7 cells, the Notides laboratory has also identified S118 as a site of ER phosphorylation but has observed S167 to be the most prominent site of phosphorylation in these cells (Arnold et al. 1994). Aurichio and coworkers (Castoria et al. 1993) have also provided strong evidence for ER phosphorylation on tyrosine 537. The roles of these phosphorylations in the activities (transcriptional and other) of the ER remains an area of great interest.

2.6 Antiestrogen Selectivity and Promoter Dependence in the cAMP-Dependent Signaling Pathway Involvement in Activation of the Transcriptional Activity of ERs Occupied by Antiestrogens

Our data provide strong evidence for the involvement of cAMP-dependent signaling pathways in the agonist actions of tamoxifen-like estrogen antagonists. The promoter specificity of the transcriptional enhancement phenomenon suggests that factors in addition to ER are probably being modulated by PKA pathway stimulation. The findings imply that changes in the cAMP content of cells, which can result in activation of the agonist activity of tamoxifen-like antiestrogens, might account, at least in part, for the resistance to antiestrogen therapy that is observed in some breast cancer patients. Of interest, MCF-7 cells transplanted into nude mice fail to grow with tamoxifen treatment initially, but some hormone-resistant cells grow out into tumors after several months of tamoxifen exposure (Gottardis and Jordan 1988; Jordan and Murphy 1990; Osborne et al. 1991). Studies have shown that this resistance to tamoxifen is, more correctly, a reflection of tamoxifen stimulation of proliferation, representing a change in the interpretation of the tamoxifen-ER complex and its agonist/antagonist balance. It is of interest that we found the pS2 gene, which is under estrogen and antiestrogen regulation in breast cancer (Brown et al. 1984), to be activated by tamoxifen in the presence of elevated cAMP. By contrast, however, antiestrogens such as ICI164,384, shown in many systems to be more complete estrogen antagonists, are not changed in their agonist/antagonist balance by increasing intracellular concentrations of cAMP. Therefore,

ICI164,384-like compounds may prove to be more efficacious and less likely to result in antiestrogen-stimulated growth.

The transcriptional enhancement we have observed between PKA activators and ER occupied by tamoxifen-like antiestrogens and E₂ provides further evidence for cross-talk between the ER and signal transduction pathways regulated by cAMP that are important in ER-dependent responses.

2.7 Bidirectional Cross-Talk Between Estrogen and cAMP Signaling Pathways

For many years, steroid hormones and peptide hormones have been considered to act via distinctly different mechanisms, the former via intracellular receptors acting through the genome and the latter via membrane-localized receptors that initially affect extranuclear activities, including the generation of second messengers such as cAMP. However, there has been increasing evidence for interactions between cAMP and estrogen in enhancing the growth of the mammary gland and breast cancer cells (Sheffield and Welsch 1985; Silberstein et al. 1984) and for cAMP induction of estrogen-like uterine growth (see Aronica et al. 1994, references therein). As early as 1967, Szego and Davis (1967) demonstrated a very rapid, acute elevation of uterine cAMP by estrogen treatment of rats *in vivo* that was confirmed in other reports, but several subsequent studies either failed to confirm this observation or reported only minimal effects that were considered to represent indirect effects of estrogen on cAMP mediated by estrogen-induced release of uterine epinephrine (see Aronica et al. 1994, references therein). Recently, cAMP and other protein kinase activators have been documented to synergize with steroid hormone-occupied receptors, leading to enhanced receptor-mediated transcription (Aronica and Katzenellenbogen 1993; Beck et al. 1993; Cho and Katzenellenbogen 1993; Fujimoto and Katzenellenbogen 1994; Groul and Altschmeid 1993; Sartorius et al. 1993), possibly by a mechanism involving phosphorylation of the receptor or associated transcription factors (Ali et al. 1993; Aronica and Katzenellenbogen 1993; Le Goff et al. 1994; Montminy et al. 1986).

Our recent studies have shown that estrogen activates adenylate cyclase, markedly increasing the concentration of cAMP in estrogen-re-

sponsive breast cancer and uterine cells in culture and in the intact uterus of rats treated with estrogen *in vivo*, in a manner that does not require new RNA or protein synthesis (Aronica et al. 1994). The intracellular concentrations of cAMP achieved by low, physiological levels of estrogen are substantial and sufficient to stimulate cAMP response element (CRE)-mediated gene transcription. Therefore, this nongenomic action of the steroid hormone estrogen involves the production of an important second messenger and the resultant activation of second messenger-stimulated genes. These findings document a two-way directionality in the cross-talk between steroid hormone- and cAMP-signaling pathways.

Although the hormonal specificity in the stimulation of cAMP is consistent with it being mediated by a high-affinity, ER-like binder, the nature of the potential membrane binder remains to be determined. Likewise, the mechanism by which estrogen enhances intracellular cAMP levels remains to be further examined. Several other publications have indicated an important role for sex hormone-binding globulin in the actions of sex steroids in enhancing intracellular cAMP (Fissore et al. 1994, Fortunati et al. 1993). The role of serum factors, including sex hormone-binding globulin, remains an important aspect for future analyses. These current observations imply a possibly broad involvement of steroid hormone action on cyclic nucleotide and second-messenger systems. The increasing evidence for rapid, membrane effects of estrogens and progestins and for vitamin D in a variety of target cells suggests that this aspect of steroid hormone action deserves further investigation (Ke and Ramirez 1987, 1990; Kim and Ramirez 1986; Lieberherr et al. 1993; Pappas et al. 1995).

Thus, data from this laboratory and others provide increasing evidence for extensive two-way cross-talk between estrogen and cAMP signaling pathways: in one way, cAMP is able to enhance the transcription of estrogen-regulated genes containing EREs; in the second way, estrogens appear able to act via the cAMP system to potentially regulate cAMP-mediated gene transcription. Further analyses of the underlying mechanisms in a variety of target cells should provide further insights in understanding the biology and regulation of estrogen-responsive cells.

References

- Ali S, Metzger D, Bornert J-M, Chambon P (1993) Modulation of transcriptional activation by ligand-dependent phosphorylation of the human estrogen receptor A/B region. *EMBO J* 12:1153-1160
- Angel P, Imagawa M, Chiu M, Stein B, Imbra RJ, Rahmsdorf HJ, Jonat C, Herrlich P, Karin M (1987) Phorbol ester-inducible genes contain a common cis-acting element recognized by a TPA-modulator trans-acting factor. *Cell* 49:729-739
- Arnold SF, Obourn JD, Jaffe H, Noides AC (1994) Serine 167 is the major estradiol-induced phosphorylation site on the human estrogen receptor. *Mol Endocrinol* 8:1208-1214
- Aronica SM, Katzenellenbogen BS (1991) Progesterone receptor regulation in uterine cells: stimulation by estrogen, cyclic adenosine 3',5'-monophosphate and insulin-like growth factor I and suppression by antiestrogens and protein kinase inhibitors. *Endocrinology* 128:2045-2052
- Aronica SM, Katzenellenbogen BS (1993) Stimulation of estrogen receptor-mediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic adenosine monophosphate, and insulin-like growth factor-I. *Mol Endocrinol* 7:743-752
- Aronica SM, Kraus WL, Katzenellenbogen BS (1994) Estrogen action via the cAMP signaling pathway: stimulation of adenylate cyclase and cAMP-regulated gene transcription. *Proc Natl Acad Sci USA* 91:8517-8521
- Beck CA, Weigel NL, Moyer ML, Nordeen SK, Edwards DP (1993) The progesterone antagonist RU486 acquires agonist activity upon stimulation of cAMP signaling pathways. *Proc Natl Acad Sci USA* 90:4441-4445
- Berry M, Metzger D, Chambon P (1990) Role of the two activating domains of the estrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-estrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-2818
- Brown AMC, Jeltsch JM, Roberts M, Chambon P (1984) Activation of pS2 gene transcription is a primary response to estrogen in the human breast cancer cell line MCF-7. *Proc Natl Acad Sci USA* 81:6344-6348
- Castoria G, Migliaccio A, Green S, Di Domenico M, Chambon P, Auricchio F (1993) Properties of a purified estradiol-dependent calf uterus tyrosine kinase. *Biochemistry* 32:1740-1750
- Cho H, Katzenellenbogen BS (1993) Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Mol Endocrinol* 7:441-452
- Cho H, Aronica SM, Katzenellenbogen BS (1994) Regulation of progesterone receptor gene expression in MCF-7 breast cancer cells: a comparison of the

B. S. Katzenellenbogen et al.

- effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology* 134:658-664
- Danielian PS, White R, Lees JA, Parker MG (1992) Identification of a conserved region required for hormone dependent transcriptional activation by steroid hormone receptors. *EMBO J* 11:1025-1033
- Danielian PS, White R, Hoare SA, Fawell SF, Parker MG (1993) Identification of residues in the estrogen receptor that confer differential sensitivity to estrogen and hydroxytamoxifen. *Mol Endocrinol* 7:232-240
- Dauvois S, Danielian PS, White R, Parker MG (1992) Antiestrogen ICI164,384 reduces cellular estrogen receptor content by increasing its turnover. *Proc Natl Acad Sci USA* 89:4037-4041
- Denton RR, Koszewski NJ, Notides AC (1992) Estrogen receptor phosphorylation: hormonal dependence and consequence on specific DNA binding. *J Biol Chem* 267:7263-7268
- Fawell SE, Lees JA, White R, Parker MG (1990a) Characterization and colocalization of steroid binding and dimerization activities in the mouse estrogen receptor. *Cell* 60:953-962
- Fawell SE, White R, Hoare S, Sydenham M, Page M, Parker MG (1990b) Inhibition of estrogen receptor-DNA binding by the "pure" antiestrogen ICI164,384 appears to be mediated by impaired receptor dimerization. *Proc Natl Acad Sci USA* 87:6883-6887
- Fissore F, Fortunati N, Comba A, Fazzari A, Gaidano G, Berta L, Frairia R (1994) The receptor-mediated action of sex steroid binding protein (SBP, SHBG): accumulation of cAMP in MCF-7 cells under SBP and estradiol treatment. *Steroids* 59:661-667
- Fortunati N, Fissore F, Fazzari A, Berta L, Benedusi-Pagliano E, Frairia R (1993) Biological relevance of the interaction between sex steroid binding protein and its receptor of MCF-7 cells: effect on the estradiol-induced cell proliferation. *J Steroid Biochem Mol Biol* 45:435-444
- Freiss G, Vignon F (1994) Antiestrogens increase protein tyrosine phosphatase activity in human breast cancer cells. *Mol Endocrinol* 8:1389-1396
- Fujimoto N, Katzenellenbogen BS (1994) Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen-selectivity and promoter-dependence. *Mol Endocrinol* 8:296-304
- Gaub M, Bellard M, Scheuer I, Chambon P, Sassone-Corsi P (1990) Activation of the ovalbumin gene by the estrogen receptor involves the fos-jun complex. *Cell* 53:1267-1276
- Gottardis MM, Jordan VC (1988) Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res* 48:5183-5187
- Groul DJ, Altschmeid J (1993) Synergistic induction of apoptosis with glucocorticoids and 3',5'-cyclic adenosine monophosphate reveals agonist activity by RU486. *Mol Endocrinol* 7:104-113
- Harlow KW, Smith DN, Katzenellenbogen JA, Greene GL, Katzenellenbogen BS (1989) Identification of cysteine-530 as the covalent attachment site of an affinity labeling estrogen (ketononestrol aziridine) and antiestrogen (tamoxifen aziridine) in the human estrogen receptor. *J Biol Chem* 264:17476-17485
- Ince BA, Zhuang Y, Wrenn CK, Shapiro DJ, Katzenellenbogen BS (1993) Powerful dominant negative mutants of the human estrogen receptor. *J Biol Chem* 268:14026-14032
- Ince BA, Schodin DJ, Shapiro DJ, Katzenellenbogen BS (1995) Repression of endogenous estrogen receptor activity in MCF-7 human breast cancer cells by dominant negative estrogen receptors. *Endocrinology* 136:3194-3199
- Jordan VC, Murphy CS (1990) Endocrine pharmacology of antiestrogens as antitumor agents. *Endocr Rev* 11:578-610
- Katzenellenbogen BS, Miller MA, Mullick A, Sheen YY (1985) Antiestrogen action in breast cancer cells: modulation of proliferation and protein synthesis, and interaction with estrogen receptors and additional antiestrogen binding sites. *Breast Cancer Res Treat* 5:231-243
- Katzenellenbogen BS, Norman MJ (1990) Multihormonal regulation of the progesterone receptor in MCF-7 human breast cancer cells: interrelationships among insulin/insulin-like growth factor-I, serum, and estrogen. *Endocrinology* 126:891-898
- Katzenellenbogen BS, Bhardwaj B, Fang H, Ince BA, Pakdel F, Reese JC, Schodin DJ, Wrenn CK (1993) Hormone binding and transcription activation by estrogen receptors: analyses using mammalian and yeast systems. *J Steroid Biochem Mol Biol* 47:39-48
- Katzenellenbogen BS, Montano MM, Le Goff P, Schodin DJ, Kraus WL, Bhardwaj B, Fujimoto N (1995) Antiestrogens: mechanisms and actions in target cells. *J Steroid Biochem Mol Biol* 53: (in press)
- Ke F, Ramirez VD (1987) Membrane mechanism mediates progesterone stimulatory effect on LHRH release from superfused rat hypothalamus in vitro. *Neuroendocrinol* 45:514-517
- Ke F, Ramirez VD (1990) Binding of progesterone to nerve cell membranes of rat brain using progesterone conjugated to ¹²⁵I-bovine serum albumin as a ligand. *J Neurochem* 54:467-472
- Kim K, Ramirez VD (1986) Effects of prostaglandin E₂, forskolin and cholera toxin on cAMP production and in vitro LH-RH release from the rat hypothalamus. *Brain Res* 386:258-265

- human estrogen receptor important in transcriptional activity of the receptor. *Mol Endocrinol* 7:1408-1417
- Pappas TC, Gametchu B, Watson CS (1995) Membrane estrogen receptors identified by multiple antibody labeling and impeded-ligand binding. *FASEB J* 9:404-410
- Reese JR, Katzenellenbogen BS (1991) Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor overexpressed in mammalian cells. *Nucleic Acids Res* 19:6595-6602
- Reese JC, Katzenellenbogen BS (1992a) Characterization of a temperature-sensitive mutation in the hormone binding domain of the human estrogen receptor: studies in cell extracts and intact cells and their implications for hormone-dependent transcriptional activation. *J Biol Chem* 267:9868-9873
- Reese JC, Katzenellenbogen BS (1992b) Examination of the DNA binding abilities of estrogen receptor in whole cells: implications for hormone-independent transactivation and the action of the pure antiestrogen ICI164,384. *Mol Cell Biol* 12:4531-4538
- Santen R, Manni A, Harvey H, Redmond C (1990) Endocrine treatment of breast cancer in women. *Endocr Rev* 11:221-265
- Sartorius CA, Tung L, Takimoto GS, Horwitz KB (1993) Antagonist-occupied human progesterone receptors bound to DNA are functionally switched to transcriptional agonists by cAMP. *J Biol Chem* 268:9262-9266
- Schüle R, Rangarajan P, Klierer S, Ransone LJ, Bolado J, Yang N, Verma IM, Evans RM (1990) Functional antagonism between oncoprotein C-jun and the glucocorticoid receptor. *Cell* 62:1217-1226
- Sheffield LG, Welsch CW (1985) Cholera-toxin-enhanced growth of human breast cancer cell lines in vitro and in vivo: interaction with estrogen. *Int J Cancer* 36:479-483
- Shemshedini L, Knaulthe R, Sassone-Corsi P, Ponnau A, Groneneyer H (1991) Cell-specific inhibitory and stimulatory effects of fos and jun on transcription activation by nuclear receptors. *EMBO J* 10:3839-3849
- Silberstein GB, Strickland PS, Trumbour V, Coleman S, Daniel CW (1984) cAMP stimulates growth and morphogenesis of mouse mammary ducts. *Proc Natl Acad Sci USA* 81:4950-4954
- Strähle U, Schmid W, Schütz G (1988) Synergistic action of the glucocorticoid receptor with transcription factors. *EMBO J* 7:3389-3395
- Sumida C, Pasqualini JR (1989) Antiestrogen antagonizes the stimulatory effect of epidermal growth factor on the induction of progesterone receptor in fetal uterine cells in culture. *Endocrinology* 124:591-597
- Sumida C, Pasqualini JR (1990) Stimulation of progesterone receptors by phorbol ester and cyclic AMP in fetal uterine cells in culture. *Mol Cell Endocrinol* 69:207-215

- Kraus WL, Montano MM, Katzenellenbogen BS (1993) Cloning of the rat progesterone receptor gene 5' region and identification of two functionally distinct promoters. *Mol Endocrinol* 7:1603-1616
- Kumar V, Green S, Staub A, Chambon P (1986) Localization of the oestrogen triol-binding and putative DNA binding domains of the human oestrogen receptor. *EMBO J* 5:2231-2236
- Kumar V, Green S, Stack G, Berry M, Jin JR, Chambon P (1987) Functional domains of the human estrogen receptor. *Cell* 51:941-951
- Lahooti H, White R, Danielian PS, Parker MG (1994) Characterization of ligand-dependent phosphorylation of the estrogen receptor. *Mol Endocrinol* 8:182-188
- Le Goff P, Montano MM, Schodin DI, Katzenellenbogen BS (1994) Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem* 269:4458-4466
- Lieberherr M, Grosse B, Kachkache M, Balsan S (1993) Cell signaling and estrogens in female rat osteoblasts: a possible involvement of unconventional nonnuclear receptors. *J Bone Miner Res* 8(11):1365-1376
- Montano MM, Muller V, Katzenellenbogen BS (1994) Role for the carboxy-terminal F domain of the estrogen receptor in transcriptional activity of receptor and in the effectiveness of antiestrogens as estrogen antagonists. *Endocrinology* [Suppl] 134:323 (abstract 491)
- Montano MM, Mueller V, Trobaugh A, Katzenellenbogen BS (1995) The carboxy-terminal F domain of the human estrogen receptor: role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 9:814-825
- Montminy MR, Sevarino KA, Wagner JA, Mandel G, Goodman RH (1986) Identification of a cyclic-AMP-responsive element within the rat somatostatin gene. *Proc Natl Acad Sci USA* 83:6682-6686
- Osborne CK, Coronado E, Allred DC, Wiebe V, DeGregorio M (1991) Acquired tamoxifen resistance: correlation with reduced breast tumor levels of tamoxifen and isomerization of trans-4-hydroxytamoxifen. *J Natl Cancer Inst* 83:1477-1482
- Pakdel F, Katzenellenbogen BS (1992) Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J Biol Chem* 267:3429-3437
- Pakdel F, Le Goff P, Katzenellenbogen BS (1993a) An assessment of the role of domain F and PEST sequences in estrogen receptor half-life and bioactivity. *J Steroid Biochem Mol Biol* 46:663-672
- Pakdel F, Reese JC, Katzenellenbogen BS (1993b) Identification of charged residues in an N-terminal portion of the hormone binding domain of the

- Sumida C, Leער F, Pasqualini JR (1988) Control of progesterone receptors in fetal uterine cells in culture: effects of estradiol, progestins, antiestrogens, and growth factors. *Endocrinology* 122:3-11
- Szego CM, Davis JS (1967) Adenosine 3',5'-monophosphate in rat uterus: acute elevation by estrogen. *Proc Natl Acad Sci USA* 58:1711-1718
- Tzukerman MT, Esty A, Santiso-Mere D, Danielian P, Parker MG, Stein RB, Pike JW, McDonnell DP (1994) Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol Endocrinol* 94:21-30
- Wrenn CK, Katzenellenbogen BS (1993) Structure-function analysis of the hormone binding domain of the human estrogen receptor by region-specific mutagenesis and phenotypic screening in yeast. *J Biol Chem* 268:24089-24098
- Yang-Yen H, Chambard J, Sun Y, Smeal T, Schmidt TJ, Drouin J, Karin M (1990) Transcriptional interference between C-jun and the glucocorticoid receptor: mutual inhibition of DNA binding due to direct protein-protein interaction. *Cell* 62:1205-1215

3 Analysis of Genetically Altered Mice Without Glucocorticoid Receptor

W. Schmid, T. Cole, J. Blendy, L. Montoliu, and G. Schütz

3.1	Introduction	51
3.2	Physiological Effects of Glucocorticoids	52
3.3	The Structure of the GR Gene and Its Disruption by Gene Targeting	54
3.4	Effects of Disruption of the GR Gene	56
3.5	Cell-Specific and Developmental Activation of a Glucocorticoid-Responsive Gene: The Tyrosine Aminotransferase Gene	58
3.6	Conclusion	61
	References	62

3.1 Introduction

Steroid hormones regulate a number of developmental and physiological processes in vertebrates by controlling the transcriptional activity of specific genes (Beato 1989, Tsai and O'Mally 1994). The ability of target cells to respond is attributed to the presence of specific receptors which mediate the action of the hormone within the cell. The receptors are localized within the nucleus in association with other proteins, which in absence of the hormone keep the receptor in an inactive state. After binding of the hormone, the hormone-receptor complex, as a dimer, binds to specific DNA sequences. The various functions of the receptor - DNA binding, ligand binding, transcriptional activation - have been assigned to separate domains of the receptor. The unliganded receptor is maintained in a nonfunctional form by oligomerization with

Different Regions in Activation Function-1 of the Human Estrogen Receptor Required for Antiestrogen- and Estradiol-dependent Transcription Activation*

(Received for publication, April 23, 1996, and in revised form, June 5, 1996)

Eileen M. McInerney^{‡§} and Benita S. Katzenellenbogen^{‡||}

From the [‡]Department of Molecular and Integrative Physiology, ^{||}Department of Cell and Structural Biology, University of Illinois, Urbana, Illinois 61801

The human estrogen receptor (ER) is a ligand-inducible transcription factor that contains two transcriptional activation functions, one located in the NH₂-terminal region of the protein (AF-1) and the second in the COOH-terminal region (AF-2). Antiestrogens, such as *trans*-hydroxytamoxifen (TOT), have partial agonistic activity in certain cell types, and studies have implied that this agonism is AF-1-dependent. We have made progressive NH₂-terminal and other segment deletions and ligations in the A/B domain, and studied the transcriptional activity of these mutant ERs in ER-negative MDA-MB-231 human breast cancer and HEC-1 human endometrial cancer cells. Using several estrogens and several partial agonist/antagonist antiestrogens, we find that estrogens and antiestrogens require different regions of AF-1 for transcriptional activation. Deletion of the first 40 amino acids has no effect on receptor activity. Anti-estrogen agonism is lost upon deletion to amino acid 87, while estrogen agonism is not lost until deletions progress to amino acid 109. Antiestrogen agonism has been further defined to require amino acids 41–64, as deletion of only these amino acids results in an ER that exhibits 100% activity with E₂, but no longer shows an agonist response to TOT. With A/B-modified receptors in which antiestrogens lose their agonistic activity, the antiestrogens then function as pure estrogen antagonists. Our studies show that in these cellular contexts, hormone-dependent transcription utilizes a range of the amino acid sequence within the A/B domain. Furthermore, the agonist/antagonist balance and activity of antiestrogens such as TOT are determined by specific sequences within the A/B domain and thus may be influenced by differences in levels of specific factors that interact with these regions of the ER.

The estrogen receptor (ER)¹ is a ligand-inducible transcrip-

tion factor that regulates gene expression through interaction with cis-acting DNA elements called estrogen response elements (EREs) (for reviews, see Refs. 1–5). Like other steroid hormone receptors, the ER contains specific domains responsible for functions leading to transcription of target genes, such as ligand binding, DNA binding, and transactivation (6–8). The ER contains two distinct, non-acidic activation functions, one activation function at the NH₂ terminus (AF-1) and a second, hormone-dependent activation function at the COOH terminus (AF-2), in the hormone binding domain (8–12). AF-2 is highly conserved among species and other nuclear hormone receptors (1, 12, 13), whereas the A/B domain at the amino terminus of the ER, which includes AF-1, is less well conserved among different species and other nuclear receptors (1, 13, 14). The activity of each activation function of ER is cell- and gene promoter-dependent. AF-1 can exhibit transcriptional activity in the absence of AF-2 (8) in some cell contents but, in most cell and promoter contexts, both AF-1 and AF-2 function in a synergistic manner and are required for full receptor activity (6, 8, 15–22).

Transactivation of estrogen-responsive genes by ER can be antagonized by antiestrogens such as *trans*-hydroxytamoxifen (TOT) and ICI 164,384 (18, 19). One mechanism by which these antiestrogens inhibit ER action is by competition with estradiol (E₂) for binding to the ER. Although antiestrogen-occupied ER binds estrogen response DNA elements in cells (23, 24), it is thought that antiestrogens promote a conformational change which is different from that induced by E₂ (24, 25). Some antiestrogens, like TOT, have partial agonistic activity in certain cells, such as chicken embryo fibroblasts (CEF) and MDA-231 human breast cancer cells (18, 26). The cell and promoter dependence of TOT agonism has been attributed to the cell and promoter specificity of AF-1 activity (15–18). Previous studies using chimeric receptors have shown that TOT is unable to induce AF-2 activity, but that TOT can be a strong agonist in cellular and promoter contexts where AF-1 is an efficient transcriptional activator (11, 18, 21).

We have investigated the A/B domain of the ER and its role in the transcriptional activity of ER elicited by estrogens and some antiestrogens, and we find that different regions within this domain are required for transcriptional stimulation by estrogen *versus* antiestrogen. In the studies presented, we demonstrate that a specific 24-amino acid region of AF-1 of the human ER is necessary for agonism by TOT and other partial agonist/antagonist antiestrogens, but is not required for E₂-dependent transactivation. As a consequence, the activity of estradiol and the estrogen agonist/antagonist character of TOT depended markedly, but not always concordantly, on the se-

* This work was supported in part by National Institutes of Health Grants 2R37CA18119 and CA60514 and United States Army Grant DAMD17-94-J-4205 (to B. S. K.). The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

§ Received partial support from National Institutes of Health Grant T32GM07238.

|| To whom correspondence should be addressed: Dept. of Molecular and Integrative Physiology, University of Illinois, 524 Burrill Hall, 407 S. Goodwin Ave., Urbana, IL 61801-3704. Fax: 217-244-9906; E-mail: katzenel@uiuc.edu.

¹ The abbreviations used are: ER, estrogen receptor; hER, human estrogen receptor; ERE, estrogen response element; E₂, 17 β -estradiol; TOT, *trans*-hydroxytamoxifen; AF, activation function; CAT, chloramphenicol acetyltransferase; HEC-1, human endometrial cancer cells; CEF, chicken embryo fibroblast cells; CMV, cytomegalovirus; BF,

2-phenylbenzofuran; BT, 2-phenylbenzothiophene; PCR, polymerase chain reaction.

quences present within the A/B domain in the ER. Our studies show that in the context of the full-length ER, hormone-dependent transcription utilizes a broad range of sequences within the A/B domain and suggest that differences in the agonist/antagonist character of antiestrogens observed in different cells could be due to altered levels of specific factors that interact with these regions.

MATERIALS AND METHODS

Chemicals and Materials—Cell culture media were purchased from Life Technologies, Inc. Calf serum was from Hyclone Laboratories (Logan, UT) and fetal calf serum was from Sigma. [¹⁴C]Chloramphenicol (50–60 Ci/mmol) was from DuPont NEN. The antiestrogens TOT and ICI 164,384 were kindly provided by Dr. Alan Wakeling, Zeneca Pharmaceuticals, Macclesfield, United Kingdom. The antiestrogens 2-phenylbenzofuran (BF) and 2-phenylbenzothiophene (BT) were generously provided by Dr. E. von Angerer, University of Regensburg, Germany.

Plasmid Constructions—The ER expression vectors, all containing human ER (hER), are derivatives of pCMV5-hER (27). NH₂-terminal deletion mutants N21 and E41 were constructed by replacement of the pCMV5-hER *Sst*II fragment with a PCR-generated fragment containing a new start codon and an *Sst*II site at amino acids 21 and 41, respectively. NH₂-terminal deletion mutants A87 and M109 were constructed by replacement of the pCMV5-hER *Sst*II/*Xma*III fragment with a PCR-generated fragment containing an *Sst*II site at amino acids 87 and 109, respectively. Estrogen receptor deleted of amino acids 41–64 (Δ 41–64) was constructed by replacing the *Sst*II fragment of pCMV5-hER (containing residues 1–64) with a PCR-generated fragment containing residues 1–40 with an *Sst*II site after amino acid 40. Δ 87–108 was constructed by inserting a PCR-generated fragment containing an *Sst*II site at amino acid 87 into the *Sst*II site of M109 and insertion of the *Xma*III fragment from this construct to replace the *Xma*III fragment of pCMV5-hER. 41–66-CDEF was constructed by replacing the *Xma*III fragment of E41 with a PCR-generated fragment containing an *Xma*III site at amino acid 180. 41–87-CDEF was constructed by replacing the *Xma*III fragment from pCMV5-hER with two PCR-generated fragments, amino acids 41–87 and amino acids 180–311 containing *Bgl*II sites at amino acids 88 and 179. 41–109-CDEF was constructed in a similar manner to 41–87-CDEF with a PCR-generated fragment, amino acids 41–109, containing a *Bgl*II site at amino acid 110. Δ AB ER was constructed as described previously (28). The sequences of all ER mutants utilized were confirmed by dideoxy sequencing methods to assure accuracy. The (ERE)₃-pS2-chloramphenicol acetyltransferase (CAT) reporter was constructed as described previously (27). The plasmid pCMV β , which contains the β -galactosidase gene, was used as an internal control for transfection. The plasmid pTZ19R, used as carrier DNA, was provided by Dr. Byron Kemper of the University of Illinois.

Cell Culture and Transient Transfections—MDA-MB-231 human breast cancer cells were maintained in Leibovitz's L-15 Medium with 10 mM HEPES, 5% calf serum, 100 units of penicillin/ml (Life Technologies, Inc.), 100 μ g of streptomycin/ml (Life Technologies, Inc.), 25 μ g of gentamicin/ml, 6 ng of bovine insulin/ml, 3.75 ng of hydrocortisone/ml, and 16 μ g of glutathione/ml. Human endometrial cancer (HEC-1) cells were maintained in minimum essential medium plus phenol red supplemented with 5% calf serum and 5% fetal calf serum, 100 units of penicillin/ml (Life Technologies, Inc.), and 100 μ g of streptomycin/ml (Life Technologies, Inc.). MDA-231 cells or HEC-1 cells were grown in minimum essential medium plus phenol red supplemented with 5% charcoal/dextran-treated calf serum for 2 days prior to transfection. Cells were plated at a density of 3×10^6 cells/100-mm dish in phenol red-free Improved minimal essential medium and 5% charcoal/dextran-treated calf serum and were given fresh medium 24 h before transfection. All cells for transfection were maintained at 37 °C in a humidified CO₂ atmosphere. Cells were transiently transfected by the CaPO₄ coprecipitation method (29). One ml of precipitate contained 0.8 μ g of pCMV β as internal control, 6 μ g of an ERE-containing reporter plasmid (ERE)₃-pS2-CAT, 100 ng of ER expression vector, and pTZ19R carrier DNA to a total of 15 μ g of DNA. Cells remained in contact with the precipitate for 4 h and were then subjected to a 2.5-min glycerol shock (20% in transfection medium). Cells were rinsed with Hanks' balanced salt solution and given fresh medium with hormone treatment as indicated.

Promoter Interference Assays—MDA-MB-231 cells were transiently transfected with 2 μ g of CMV-(ERE)₂-CAT reporter plasmid (23), 0.8 μ g of pCMV β , 12.2 μ g of pTZ19R, and 100 ng of ER expression vector/100-mm dish of cells. Cells were treated as described previously for

transient transfection, and CAT assays were performed on cell extracts.

Immunoblot Assays—COS-1 cells were transfected in 100-mm dishes with 10 μ g of expression vector for wild type ER or ER derivatives and 5 μ g of pTZ19R carrier plasmid. Whole cell extracts were collected by centrifugation and fractionated on a polyacrylamide gel. Proteins were transferred to nitrocellulose and immunoblots were performed using ER monoclonal antibody H222 as described previously (30).

RESULTS

Different Regions in the A/B Domain Are Important for Estradiol- and trans-Hydroxytamoxifen-dependent Transcriptional Activity—Our studies were aimed at identifying regions within the A/B domain that are responsible for E₂-dependent transcription and for antiestrogen agonism. We have generated ER derivatives that contain increasing NH₂-terminal deletions or other deletional changes in the A/B domain. Fig. 1 shows the structure of the ER derivatives used in this study and the relative expression levels of the receptors observed in cells. Western immunoblot analysis showed that receptors of the predicted sizes were being produced in the cells and that all of the A/B domain altered receptors (Fig. 1B) were expressed at levels very similar to that of the wild type ER.

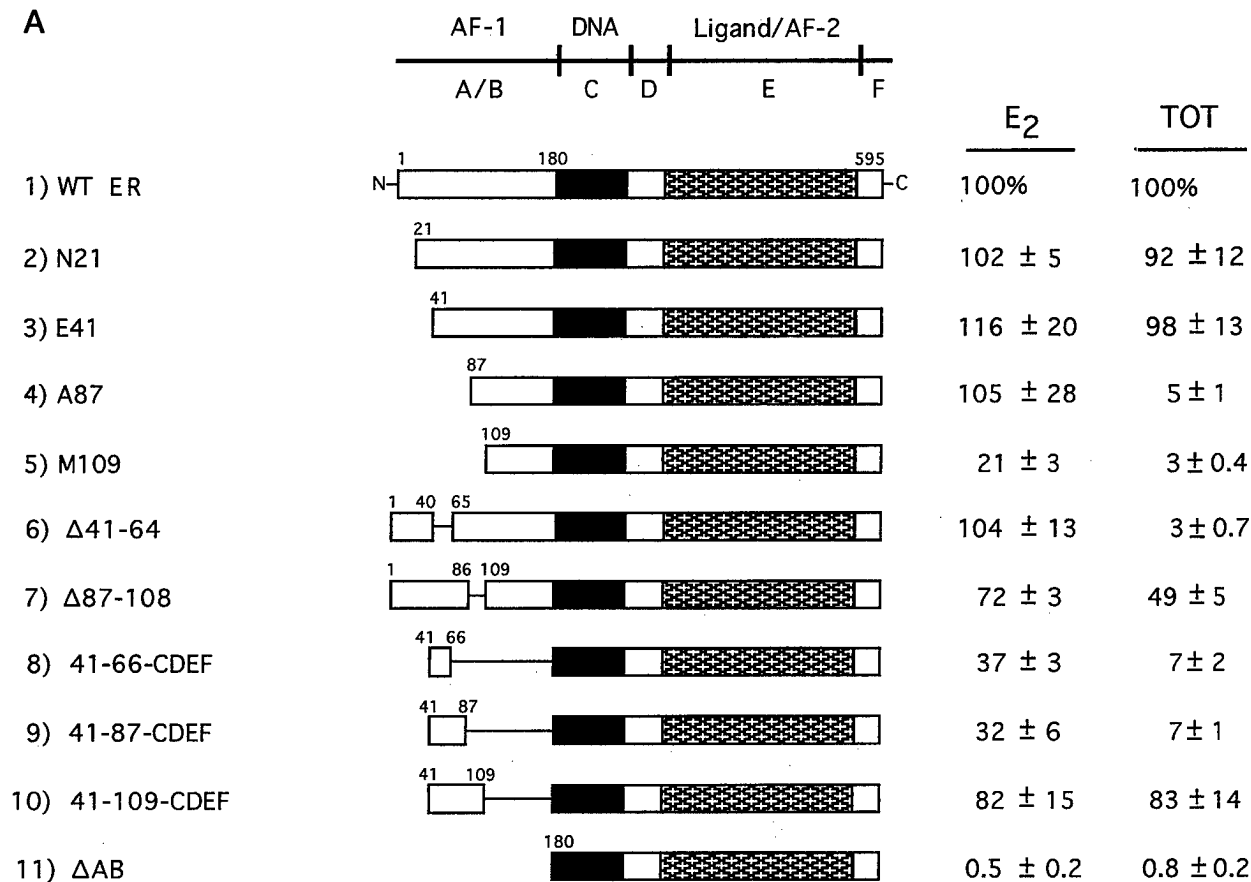
These ER mutants were then analyzed for their ability to transactivate an ERE-containing pS2 promoter-reporter gene in ER-negative MDA-231 human breast cancer cells. Wild type ER or receptors with deletions of amino acids 1–20 (N21), 1–40 (E41), 1–86 (A87), 1–108 (M109), or 1–179 (Δ AB) were transiently transfected into MDA-231 cells, and transcriptional activity was measured in response to increasing concentrations of E₂. ER mutants N21, E41, and A87 showed dose-response curves for transcriptional activity virtually identical to that observed with wild type ER (Fig. 2A). In contrast, deletion of the first 108 amino acids resulted in receptors that showed a great loss of activity; M109 receptors showed only about 20% of wild type ER transcriptional activity at 10^{–8} M E₂, suggesting that residues between amino acid 87 and 108 are important for estradiol-stimulated activity. Deletion of the complete A/B domain (amino acids 1–179) gave a receptor that showed no activity in this cell system.

Similar studies were conducted using the NH₂-terminal deletion mutants to examine transcriptional response to the triphenylethylene compound *trans*-hydroxytamoxifen, TOT (Fig. 2B). MDA-231 cells were again used in these studies, since with wild type ER, TOT behaves as a relatively strong agonist. TOT (10^{–7} M) stimulates transcriptional activity to approximately 30% the level evoked by maximal (10^{–8} M) E₂ stimulation. Compared with the wild type ER, deletion of amino acids 1–20 or 1–40 had no effect on either the E₂ response or TOT agonism. However, deletion of amino acids 1–86, which had no effect on E₂-induced activity, abolished TOT agonism completely (Fig. 2B). The further deleted mutant, M109, which was transcriptionally impaired in response to E₂ treatment, did not exhibit any measurable response to TOT. The loss of TOT agonism observed selectively with the A87 mutant suggested that sequences between 41 and 87 may be important contributors to TOT agonism, but are not essential for the response to E₂.

Deletion mutant Δ 41–64, which lacks only amino acids 41–64, was constructed and tested for its transactivation ability in response to E₂ and TOT. Δ 41–64 retained 100% of wild type E₂-dependent activity (Fig. 2C) yet displayed no measurable response to TOT (Fig. 2D). These results are consistent with the loss of TOT response with the A87 mutant as they implicate residues 41–64 as a major contributor to TOT agonism but not to E₂ response.

A/B Deletion Mutants Exhibit Differential Response to Other Estrogens and Antiestrogens—Further examination of the ligand-dependent transcriptional activity of these mutants revealed

A



B

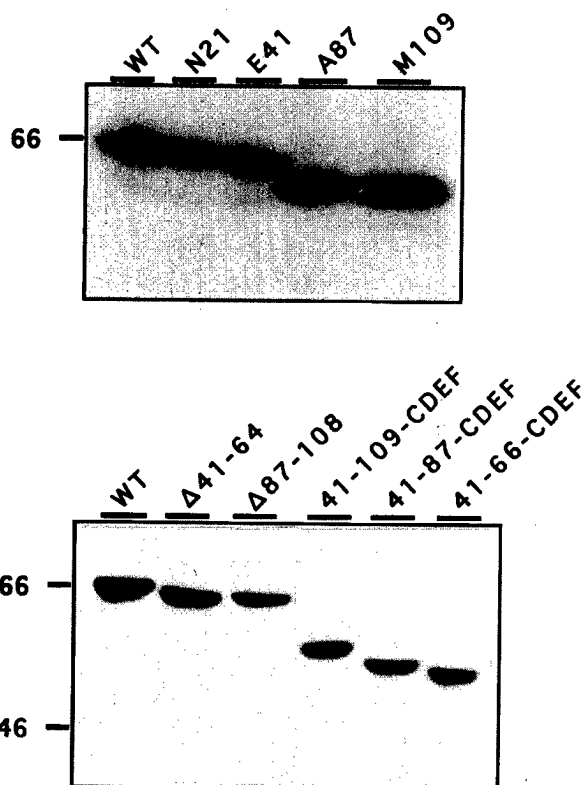
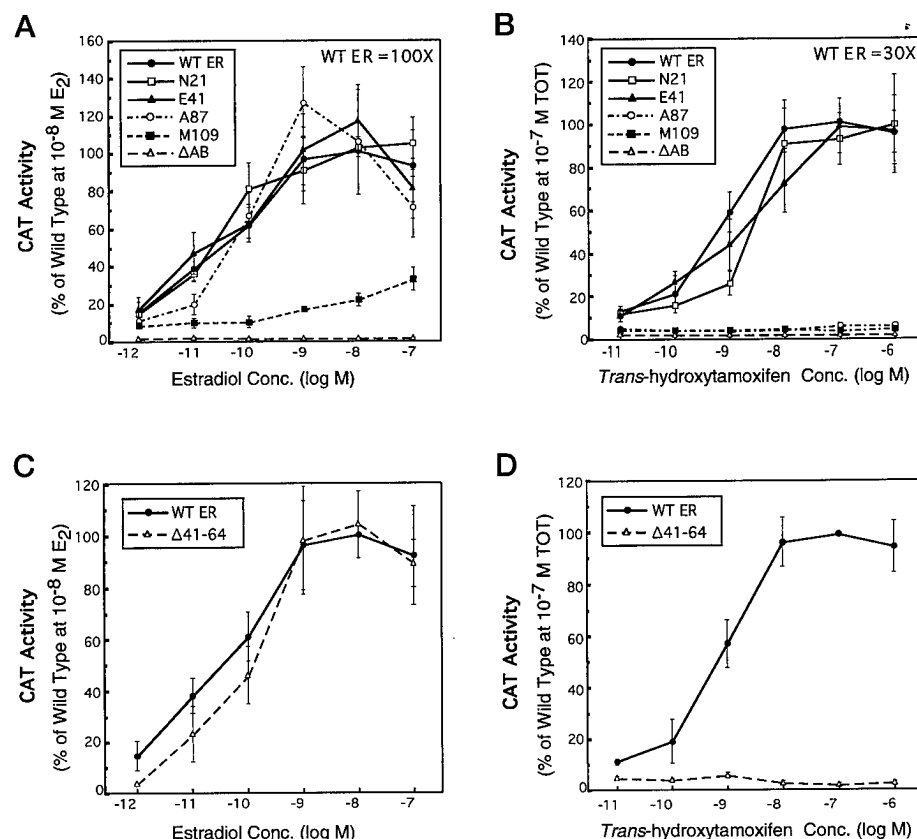


FIG. 1. Structure and expression of ER derivatives. A, the functional domains (A/B, C, D, E, F) and activation functions (AF-1 and AF-2) of ER are shown at the top along with schematics for the A/B domain mutants studied in this report. The values to the right of the receptor

FIG. 2. Transcriptional activation by wild type ER and A/B domain deletion ER mutants. ER-negative MDA-231 cells were transfected with expression vector for wild type or mutant ER and a (ERE)₃-pS2-CAT reporter gene. Cells were treated with increasing concentrations of E₂ (A and C) or TOT (B and D) for 24 h. CAT activity was normalized for β -galactosidase activity from an internal control plasmid. Values represent the mean \pm S.E. for three or more determinations and are expressed as a percentage of wild type ER response with 10⁻⁸ M E₂ or 10⁻⁷ M TOT. For some values, error bars are too small to be visible. Wild type ER showed a ~100-fold and 30-fold induction of CAT activity in response to 10⁻⁸ M E₂ or 10⁻⁷ M TOT, respectively.



that another full estrogen, the resorcylic lactone P1496 (31), showed a pattern of activity identical to that observed with E₂. Like E₂, transcriptional response to P1496 was fully retained in N21, E41, and A87 receptors, but was impaired with the deletion of the first 108 residues (Fig. 3A). Similar results to those seen with TOT were observed with the antiestrogen compounds BF and BT (32). Like TOT, these heterocycle-based antiestrogens were significant agonists, evoking transcriptional activity that was similar in magnitude to that obtained with TOT (~30% of E₂ stimulation). As seen in Fig. 3A, antiestrogen stimulation of CAT activity was lost with the mutants A87 and Δ 41-64 for the three antiestrogen compounds (TOT, BF, and BT), while estrogen (E₂ and P1496) stimulation of transcriptional activity was still maintained maximally in these two constructs. No stimulation of wild type ER or any ER mutants was seen with the pure antiestrogen ICI 164,384 (data not shown).

These A/B domain mutants were also tested in a different cell background utilizing an ER-negative human endometrial cancer cell line (HEC-1 cells). In these cells, wild type ER also responds to TOT as an agonist, showing about 30–40% of wild type E₂ response (Fig. 3B). Similar results to those seen previously in MDA-231 breast cancer cells were observed with the A/B domain deletion mutants in these endometrial cancer cells; both A87 and Δ 41-64 receptors retained full wild type transcriptional activity in response to E₂ but did not exhibit any response to TOT. These results demonstrate again that a region between amino acids 40 and 65 is critical for TOT agonism yet is not required for E₂-dependent transcription.

Specific Regions in the A/B Domain Are Required to Support TOT Agonism—Since TOT was not a full estrogen agonist in

these assays, and is known to show mixed estrogen agonist and antagonist activity in many cells (15–18), we also examined the antagonist activity of TOT and how this was impacted by changes in the A/B domain of ER (Fig. 4). TOT agonism was apparent in wild type ER, N21, and E41 receptors and, in these three receptors, TOT (at a 10-fold excess concentration relative to that of E₂) was also able to suppress E₂-stimulated activity to that of its own inherent level of agonism (*i.e.* approximately 30% of the E₂-stimulated level). Thus, with these receptors, this compound showed partial agonist and partial antagonist activity. Of interest, in the A87, M109, and Δ 41-64 receptors where TOT showed no agonistic activity, TOT behaved as a pure antiestrogen and was now a complete antagonist of the E₂ stimulation. Thus, the agonist/antagonist character of the antiestrogen TOT differed with the nature of the ER A/B domain.

Deletions in the A/B Domain Do Not Affect Receptor Level or DNA Binding—Since certain A/B deletion mutants exhibited a differential response to estrogens and antiestrogens, the levels of these receptors and the DNA binding abilities of these mutant ERs were determined following exposure to E₂ or TOT in order to determine whether differences in response to these two ligands might be attributable to ligand-induced alteration in receptor stability or DNA binding ability. As seen in Fig. 5A, levels of wild type ER, Δ 41-64 ER and A87 ER were similar following cell treatment with E₂ or TOT. Thus, differential turnover of these receptor proteins in response to TOT *versus* E₂ is not likely to explain the very different transcriptional response of these receptors to these two ligands.

DNA binding studies were conducted with several of the mutants by use of a promoter interference assay, in order to assess whether differences in DNA binding of the TOT-ER

schematics indicate the transcriptional activity of the receptors in response to 10⁻⁸ M E₂ or 10⁻⁷ M TOT and summarize data that are derived from dose response experiments detailed later in this paper. B, the expression of wild type and mutant estrogen receptors from cytomegalovirus promoter-containing expression vectors was determined following transfection into ER-negative COS-1 cells. Equal amounts of protein were used and immunoblotting was done with the anti-ER monoclonal antibody H222.

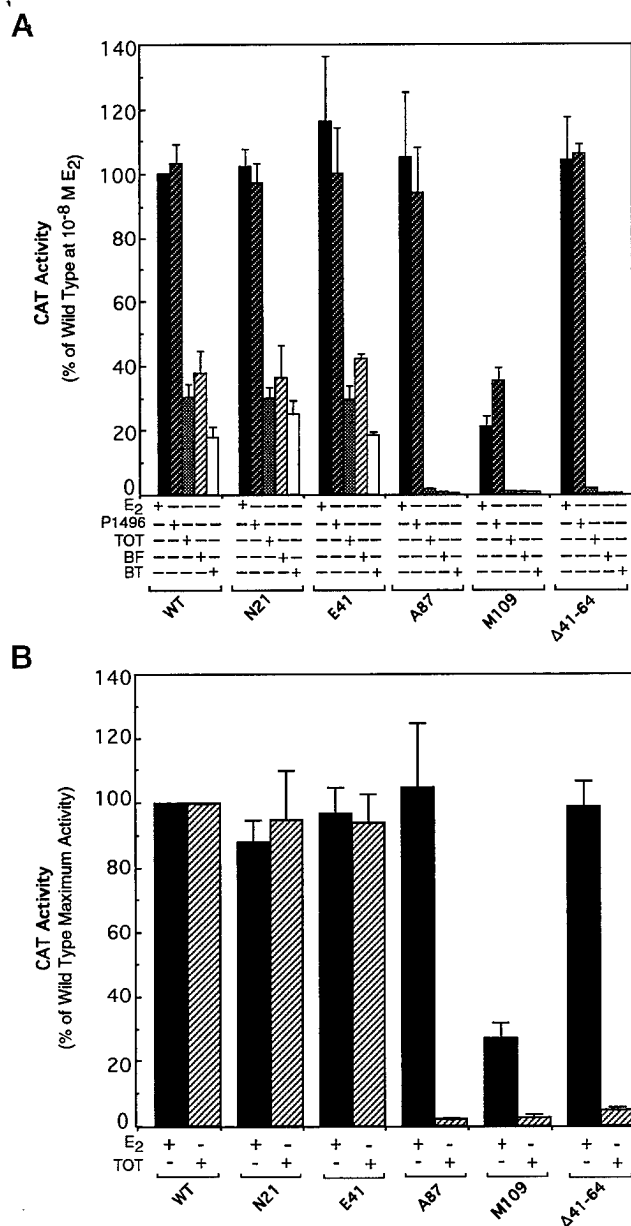


FIG. 3. Transcriptional activation by wild type ER and A/B domain deletion ER mutants in response to two estrogens and three antiestrogens. A, MDA-231 breast cancer cells were transfected with ER expression vectors and a (ERE)₃-pS2-CAT reporter gene. Cells were treated for 24 h with either 10^{-8} M E₂, 10^{-7} M P1496, 10^{-7} M TOT, 10^{-7} M BF, or 10^{-7} M BT as indicated. B, ER-negative HEC-1 human endometrial cancer cells were transfected with ER expression vectors and a (ERE)₃-pS2-CAT reporter gene and treated with either 10^{-8} M E₂ or 10^{-7} M TOT. CAT activity was determined as described in the legend to Fig. 2. Values are the mean \pm S.E. for three or more determinations from separate experiments. Some error bars are too small to be visible.

versus E₂-ER complexes might explain their different transcriptional efficacy (Fig. 5B). This promoter interference assay measures the ability of ER to bind to ERE DNA in intact cells (23). Binding of ER to the ERE is assayed by assessing the ability of ERE-bound ER to block transcription from the constitutively active cytomegalovirus (CMV) promoter, with the repression of CAT activity being a measure of the binding of ER to the ERE-containing promoter. A87, which responds to E₂ but not to TOT, and M109, which is impaired in both E₂- and TOT-dependent activity, were both able to bind to the EREs and to interfere with promoter activity to the same extent as the wild type ER (Fig. 5B). Therefore, differences in E₂- and

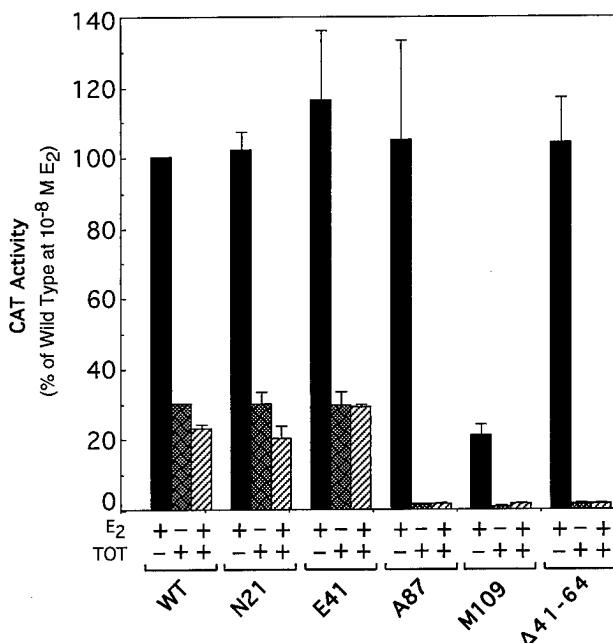


FIG. 4. The antiestrogen TOT is an estrogen agonist and antagonist, with its agonist/antagonist balance dependent on the particular ER protein. ER-negative MDA-231 cells were transfected with expression vector for wild type or A/B domain ER mutants and a (ERE)₃-pS2-CAT reporter gene. Cells were treated for 24 h with 10^{-8} M E₂ or 10^{-7} M TOT alone or in combination (10^{-8} M E₂ and 10^{-7} M TOT). CAT activity was analyzed as described in the legend to Fig. 2. Values are the mean \pm S.E. for three or more determinations from separate experiments. Some error bars are too small to be visible.

TOT-dependent transactivation exhibited by these ER derivatives do not appear to be caused by differences in receptor protein level or by differential DNA binding.

Residues 41–109 Encompass Sequences Important for Both Estradiol- and TOT-dependent Transcription—Additional analysis of the A/B region was made to further characterize sequences important for E₂- and TOT-dependent transcription. Since transcriptional response to E₂ was almost completely lost in going from the A87 to the M109 ER, we wished to directly assess the importance of amino acids 87–108 in E₂-dependent activity. To do so, we tested an ER mutant lacking only amino acids 87–108 (Δ 87–108). Full dose-response studies employing 10^{-12} to 10^{-7} M E₂ and 10^{-11} to 10^{-6} M TOT were conducted for this mutant and all other mutants described below, as done for the mutant ERs shown in Fig. 2. The dose-response curves are not shown, but the findings at 10^{-8} M E₂ and 10^{-7} M TOT are summarized in Fig. 1A. Deletion of residues 87–108 resulted in only a ~30% decrease in E₂-stimulated transcriptional activity (Fig. 1A, entry 7). From these results, it appears that E₂-dependent transcription is supported by sequences outside of the 87–108 region of the A/B domain, as deletion of only these amino acids is not sufficient to reduce the transcriptional activity to the level observed with M109.

Further analysis of the A/B region was made using segment ligated mutants (Fig. 1A, entries 8–10). To examine the region between residues 40 and 65, which were required for TOT agonism, we constructed a segment ligated ER derivative, 41–66-CDEF, containing only amino acids 41–66 of the A/B domain linked directly to the intact ER domains C through F and assayed this receptor for its ability to transactivate an ERE-containing reporter gene in the presence of E₂ or TOT. This mutant was surprising in its ability to activate the reporter gene to approximately 40% of the wild type ER in response to E₂ (Fig. 1A, entry 8), even though deletion of amino acids 41–64 resulted in no change in E₂-stimulated activity. The ER mutant

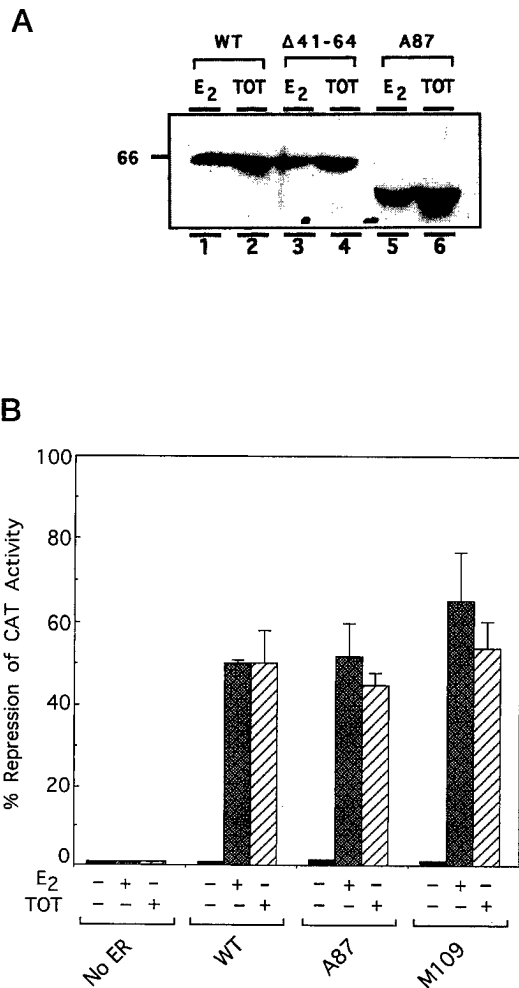


FIG. 5. Protein levels and DNA-binding abilities of wild type ER and ER mutants treated with estrogen or antiestrogen. A, levels of wild type ER and ER mutants were examined following transfection and treatment of COS-1 cells with either 10^{-8} M E_2 or 10^{-7} M TOT for 24 h. Immunoblotting was done with the anti-ER monoclonal antibody H222. B, MDA-231 cells were transfected with the constitutively active CMV-(ERE)₂-CAT promoter interference plasmid and wild type ER or mutant ERs. Cells were treated with control vehicle, 10^{-8} M E_2 , or 10^{-7} M TOT, and CAT activity was analyzed as described in the legend to Fig. 2. Values are the mean \pm S.E. for three or more determinations from separate experiments. For some values, error bars are too small to be visible.

41–66-CDEF, however, exhibited no measurable response to TOT. This suggests that residues 41–64 are necessary for TOT agonism, but that they alone are not sufficient for TOT-directed transcription. Extension of the A/B domain toward the COOH terminus (Fig. 1A, entry 9) to include amino acids 41–87 (41–87-CDEF) did not result in any increase in E_2 - or TOT-dependent transcription compared with 41–66-CDEF. However, extension to amino acid 109 (41–109-CDEF) did result in a 2-fold increase in E_2 -dependent transcriptional activity compared with 41–66-CDEF and a dramatic increase in TOT agonism such that the activity measured was approximately 80% of wild type ER activity for both E_2 and TOT (Fig. 1A, entry 10). This indicates that the region encompassing residues 41–109 contains almost all of the A/B domain sequence needed both for E_2 and TOT stimulatory activity.

Interestingly, the transcriptional activity of 80% observed with 41–109-CDEF is in agreement with the observation that only 20% of wild type E_2 -stimulated activity is retained upon deletion of the first 108 residues. These results suggest that residues 87–108 play a significant role in E_2 -stimulated tran-

scriptional activity but are supported by other sequences in the A/B domain. This is highlighted by the $\Delta 87$ –108 mutant (Fig. 1A, entry 7), which lacks residues 87–108 in the A/B domain. This mutant is only weakly impaired in response to E_2 and TOT compared with wild type ER, consistent with residues 41–109 being important for full AF-1 function. Together, these results demonstrate that E_2 - and TOT-dependent transcription utilizes other flanking sequences beyond amino acids 87–108 within the A/B domain to achieve full receptor activity. These required regions could serve as a portion of the activation function or could serve a structural purpose, perhaps maintaining proper three-dimensional structure of the receptor protein.

DISCUSSION

The human estrogen receptor contains two transcriptional activation functions, AF-1 located in the A/B domain and AF-2 in the hormone-binding domain. Both transcriptional activation functions act in a promoter- and cell type-dependent manner. The amino acid sequences of these activation functions are not similar to other known activation sequences, so elucidation of their precise mechanism of action is of interest. Our studies have defined AF-1 regions within the A/B domain of ER that support the transcriptional response to estrogens (E_2 , P1496) and those that support the transcriptional response to several antiestrogens. While considerable overlap in the transcription-supporting regions is observed for both categories of ligands, we found that there are some distinct sequence requirements.

There are limitations in the applications of mutational methods to precisely define regions of the A/B domain that support the transcriptional agonism of these different ligands, as these activities appear to be distributed over more than one discrete segment. To address these issues we have, in fact, made three different types of alterations in the A/B domain, namely progressive NH₂-terminal deletions, segmental deletions, and segmental ligations. In many cases, we obtained consistent results regarding the transcription-supporting role of a particular region of the A/B domain by making the different types of mutations; however, we did not always get identical results using all three approaches.

When making progressive NH₂-terminal deletions, TOT agonism is lost when the A/B domain is truncated from E41 to A87, whereas the effect of E_2 is reduced only upon further deletion to M109. Therefore, TOT agonism appears to require a region between residues 41–86, whereas E_2 induction requires the 87–108 sequence. Segmental deletion of residues 41–64 does, in fact, eliminate TOT agonism without affecting E_2 induction. However, the 87–108 segmental deletion, which has a limited effect on TOT agonism, causes only a modest reduction in E_2 induced transcription. Thus, whereas the region 87–108 appears to be critical to the E_2 effect in the absence of residues 1–86 (i.e. by progressive NH₂-terminal deletion), it appears that much of the E_2 effect can be supported by the 1–86 segment (perhaps together with the 109–180 segment) that is still present in the $\Delta 87$ –108 segment-deleted mutant. The segment ligation approach confirms the importance of the 41–109 region, as this segment alone restores most of the agonistic effect of TOT and gives nearly full induction with E_2 . It is clear from our findings that distinctly different regions of the A/B domain are responsible for supporting the transcriptional activation induced by E_2 and the agonism effected by TOT and that in certain situations these regions may act in concert with other A/B segments.

Metzger *et al.* (21) analyzed the role of A/B sequences in chicken embryo fibroblast (CEF) and yeast cells in which AF-1 is able on its own to stimulate transactivation. They observed in CEF cells that deletion of the first ~60 or 80 residues resulted in a decrease in E_2 -stimulated transcription of 40 and

70%, respectively. In our studies in 231 human breast cancer and HEC-1 human endometrial cancer cells, deletion of the first 40 amino acids, had no effect on transcriptional activity, while deletion of the first 108 amino acids nearly completely eliminated transcriptional response to E_2 . Response to E_2 was fully retained in our A87 mutant, yet this mutant lost its ability to respond to TOT. In this and some other A/B domain mutants, we observed considerable differences in the ability of TOT versus E_2 to stimulate transcription, whereas in the several mutants analyzed for response to E_2 and TOT in CEF cells, which contained deletions of only certain NH_2 - or $COOH$ -terminal portions of the A/B domain, differences between E_2 and TOT were not seen. The differences in our findings and those of Metzger *et al.* (21) may reflect differences in the cell types and promoters studied, but may also reflect the fact that deletions in only the central portion of the A/B domain were not studied by Metzger *et al.* (21).

Tamoxifen is well known to show cell- and gene-specific agonism, being a relatively pure estrogen antagonist in some cells, and a partial agonist/antagonist or a relatively strong agonist in others (5, 22). Our current findings suggest that cellular processes that impinge on the specific A/B domain sequences we have identified should be key determinants of whether ligands such as tamoxifen will function as agonists, antagonists, or partial agonists/antagonists in any specific cell system. In a recent study, we have shown that the binding of both estrogens and antiestrogens to ER promotes an interaction between AF-1 in the A/B domain and AF-2 in domain E (27). This AF-1/AF-2 interaction appears to be an essential prerequisite for the competence of ER-ligand complexes to induce transcription. It is known that there are conformational differences in ER-estrogen and ER-antiestrogen complexes (24, 25, 33), which are presumed to occur in the ligand binding AF-2 region. Since the interaction of AF-2 with AF-1 is required for optimal transcriptional activity in the cell contexts we have examined, it is not surprising that distinctly different sequences within AF-1 are involved in supporting the transcription activation induced by these different ligand classes.

The mechanisms by which ligand-induced AF-1/AF-2 interaction occurs or by which ER-ligand complexes are able to elicit gene transcription are not well understood. These activation functions have been shown to have squelching effects on their own activity and on acidic activators (9). This transcriptional interference provides evidence that AF-1 and AF-2 interact with a titratable cellular factor(s) indispensable for different classes of activation functions (8, 9). A number of activation function-interacting proteins may be involved in these processes (Ref. 22 and references therein) and may account, as well, for the varying levels of agonism that TOT displays in different cells and on different promoters. For example, in systems in which TOT has agonist activity, a co-regulator or transcription factor that interacts specifically with the 41–64 region of AF-1 in the ER-TOT complex may support transcription, whereas systems in which TOT is a pure antagonist may lack this factor. E_2 -induced transcription, which operates via somewhat different AF-1 sequences, may not utilize this factor or may utilize

other factors. Our identification of differences in the sequences within ER that are required for TOT versus estradiol agonism should aid in elucidating the underlying mechanisms regulating the cell-specific pharmacology and biocharacter of antiestrogens.

Acknowledgments—We are grateful to Drs. Edwin von Angerer and Alan Wakeling for providing antiestrogens and thank Dr. Geoffrey Greene and Abbott Laboratories for providing estrogen receptor antibodies.

REFERENCES

- Evans, R. M. (1988) *Science* **240**, 889–895
- Mangelsdorf, D. J., Thummel, C., Beato, M., Herrlich, P., Schutz, G., Umesono, K., Blumberg, B., Kastner, P., Mark, M., Chambon, P., and Evans, R. M. (1995) *Cell* **83**, 835–839
- Gronemeyer, H. (1991) *Annu. Rev. Genet.* **25**, 89–123
- Tsai, M. J., and O'Malley, B. W. (1994) *Annu. Rev. Biochem.* **63**, 451–486
- Katzenellenbogen, B. S. (1996) *Biol. Reprod.* **54**, 287–293
- Kumar, V., Green, S., Stack, G., Berry, M., Jin, J.-R., and Chambon, P. (1987) *Cell* **51**, 941–951
- Kumar, V., Green, S., Staub, A., and Chambon, P. (1986) *EMBO J.* **5**, 2231–2236
- Tora, L., White, J., Brou, C., Tasset, D., Webster, N., Scheer, E., and Chambon, P. (1989) *Cell* **59**, 477–487
- Tasset, D., Tora, L., Fromental, C., Scheer, E., and Chambon, P. (1990) *Cell* **62**, 1177–1187
- Lees, J. A., Fawell, S. E., and Parker, M. G. (1989) *Nucleic Acids Res.* **17**, 5477–5487
- Webster, N. J., Green, S., Jin, J. R., and Chambon, P. (1988) *Cell* **54**, 199–207
- Danielian, P. S., White, R., Lees, J. A., and Parker, M. G. (1992) *EMBO J.* **11**, 1025–1033
- White, R., Lees, J. A., Needham, M., Ham, J., and Parker, M. G. (1987) *Mol. Endocrinol.* **1**, 735–744
- Krust, A., Green, S., Argos, P., Kumar, V., Walter, P., Bornert, J. M., and Chambon, P. (1986) *EMBO J.* **5**, 891–897
- Metzger, D., Losson, R., Bornert, J. M., Lemoine, Y., and Chambon, P. (1992) *Nucleic Acids Res.* **20**, 2813–2817
- Tzukerman, M. T., Esty, A., Santiso-Mere, D., Danielian, P., Parker, M. G., Stein, R. B., Pike, J. W., and McDonnell, D. P. (1994) *Mol. Endocrinol.* **8**, 21–30
- Pham, T. A., Hwung, Y. P., Santiso-Mere, D., McDonnell, D. P., and O'Malley, B. W. (1992) *Mol. Endocrinol.* **6**, 1043–1050
- Berry, M., Metzger, D., and Chambon, P. (1990) *EMBO J.* **9**, 2811–2818
- Katzenellenbogen, B. S., Montano, M., LeGoff, P., Schodin, D. J., Kraus, W. L., Bhardwaj, B., and Fujimoto, N. (1995) *J. Steroid Biochem. Mol. Biol.* **53**, 387–393
- Bocquel, M. T., Kumar, V., Chambon, P., and Gronemeyer, H. (1989) *Nucleic Acids Res.* **17**, 2581–2595
- Metzger, D., Ali, S., Bornert, J.-M., and Chambon, P. (1995) *J. Biol. Chem.* **270**, 9535–9542
- Katzenellenbogen, J. A., O'Malley, B. W., and Katzenellenbogen, B. S. (1996) *Mol. Endocrinol.* **10**, 119–131
- Reese, J. C., and Katzenellenbogen, B. S. (1992) *Mol. Cell. Biol.* **12**, 4531–4538
- McDonnell, D. P., Clemm, D. L., Hermann, T., Goldman, M. E., and Pike, J. W. (1995) *Mol. Endocrinol.* **9**, 659–669
- Beekman, J. M., Allan, G. F., Tsai, S. Y., and O'Malley, B. W. (1993) *Mol. Endocrinol.* **7**, 1266–1274
- Montano, M. M., Muller, V., Trobaugh, A., Katzenellenbogen, B. S. (1995) *Mol. Endocrinol.* **9**, 814–825
- Kraus, W. L., McInerney, E. M., and Katzenellenbogen, B. S. (1995) *Proc. Natl. Acad. Sci. U. S. A.* **92**, 12314–12318
- LeGoff, P., Montano, M. M., Schodin, D. J., and Katzenellenbogen, B. S. (1994) *J. Biol. Chem.* **269**, 4458–4466
- Chen, C., and Okayama, H. (1987) *Mol. Cell. Biol.* **8**, 5323–5330
- Wrenn, C. K., and Katzenellenbogen, B. S. (1990) *Mol. Endocrinol.* **4**, 1647–1654
- Katzenellenbogen, B. S., Katzenellenbogen, J. A., and Mordecai, D. Z. (1979) *Endocrinology* **105**, 33–40
- von Angerer, E., Biberger, C., and Leichter, S. (1995) *Ann. N. Y. Acad. Sci.* **761**, 176–191
- Allan, G. F., Leng, X., Tsai, S. Y., Weigel, N. L., Edwards, D. P., Tsai, M.-J., and O'Malley, B. W. (1992) *J. Biol. Chem.* **267**, 19513–19520

Analysis of estrogen receptor transcriptional enhancement by a nuclear hormone receptor coactivator

(activation domain/nuclear receptor/coactivator/transcriptional synergism)

EILEEN M. MCINERNEY*, MING-JER TSAI†, BERT W. O'MALLEY†, AND BENITA S. KATZENELLENBOGEN*‡§

Departments of *Molecular and Integrative Physiology and †Cell and Structural Biology, University of Illinois, Urbana, IL 61801; and ‡Department of Cell Biology, Baylor College of Medicine, Houston, TX 77030

Contributed by Bert O'Malley, June 25, 1996

ABSTRACT The estrogen receptor (ER), a member of a large superfamily of nuclear hormone receptors, is a ligand-inducible transcription factor that regulates the expression of estrogen-responsive genes. The ER, in common with other members of this superfamily, contains two transcription activation functions (AFs)—one located in the amino-terminal region (AF-1) and the second located in the carboxyl-terminal region (AF-2). In most cell contexts, the synergistic activity of AF-1 and AF-2 is required for full estradiol (E₂)-stimulated activity. We have previously shown that a ligand-dependent interaction between the two AF-containing regions of ER was promoted by E₂ and the antiestrogen *trans*-hydroxytamoxifen (TOT). This interaction, however, was transcriptionally productive only in the presence of E₂. To explore a possible role of steroid receptor coactivators in transcriptional synergism between AF-1 and AF-2, we expressed the amino terminal (AF-1-containing) and carboxyl-terminal (AF-2-containing) regions of ER as separate polypeptides in mammalian cells, along with the steroid receptor coactivator-1 protein (SRC-1). We demonstrate that SRC-1, which has been shown to significantly increase ER transcriptional activity, enhanced the interaction, mediated by either E₂ or TOT, between the AF-1-containing and AF-2-containing regions of the ER. However, this enhanced interaction resulted in increased transcriptional effectiveness only with E₂ and not with TOT, consistent with the effects of SRC-1 on the full-length receptor. Our results suggest that after ligand binding, SRC-1 may act, in part, as an adapter protein that promotes the integration of amino- and carboxyl-terminal receptor functions, allowing for full receptor activation. Potentially, SRC-1 may be capable of enhancing the transcriptional activity of related nuclear receptor superfamily members by facilitating the productive association of the two AF-containing regions in these receptors.

The estrogen receptor (ER) is a 66-kDa, ligand-inducible transcription factor that regulates the transcription of estrogen-responsive genes (for reviews see refs. 1–3). Like other steroid hormone receptors, the ER is a modular protein that can be divided into separable domains with specific functions, such as ligand binding, dimerization, DNA binding, and transactivation (4–7). In addition to a centrally located C domain, corresponding to the DNA binding domain, the ER contains two distinct activation functions (AFs; refs. 6–9). The AF located in the amino-terminal A/B domain is termed AF-1, and a second, hormone-dependent AF (AF-2) is located in the E domain along with the hormone binding function of ER. AF-1 and AF-2 function in a synergistic manner and are required for full ER activity in most cell contexts (7, 10, 11). Like other activation domains, the AFs of ER are thought to be important targets for basal transcriptional factors or specific

cellular proteins that function as coactivators. The activity of each AF of ER varies in different cellular contexts, and these AFs have been shown to have squelching effects on their own activity and on the activity of other receptors (9), providing evidence that AF-1 and AF-2 interact with cellular proteins, which may be distinct from the basal transcription factors.

Previously, we have shown that when the amino-terminal region (ABCD) and the carboxyl-terminal region (EF) of the ER were expressed as separate polypeptides in mammalian cells, they were capable of interacting in an estradiol (E₂)-dependent manner to reconstitute the transcriptional activity of ER (12). Furthermore, we demonstrated that the interaction between ABCD and EF was also promoted by the antiestrogen *trans*-hydroxytamoxifen (TOT); however, this interaction was not transcriptionally productive. Although these studies provided information regarding ER transactivation through synergism between the two ER AFs, these studies were unable to determine whether the interaction between the amino- and carboxyl-terminal regions was direct or indirect, perhaps requiring intermediary proteins to promote the association of the AF-1- and AF-2-containing regions of the receptor. It is possible that the interaction between AF-1 and AF-2 requires accessory proteins, possibly a coactivator, to contribute to the transcriptionally productive association between the amino- and carboxyl-terminal regions of ER. We were interested in determining how coactivators, required for optimal ER transactivation, enhance receptor activity.

Using a yeast two-hybrid system, Oñate *et al.* (13) recently identified the steroid receptor coactivator-1 (SRC-1) protein, which interacted in a ligand-dependent manner with the hormone binding domain of the progesterone receptor. More recently, SRC-1 has been postulated to exist as a family of proteins related to p160 (ERAP160) (14, 15). SRC-1 was shown to significantly enhance the transcriptional activity of ER and other steroid hormone receptors. Overexpression of SRC-1 also reversed the squelching of progesterone receptor transcriptional activity upon coexpression of ligand-bound ER, demonstrating that SRC-1 is a genuine coactivator for steroid hormone receptors. It is unknown what precise function SRC-1 or other coactivators perform after binding to the receptor to result in enhanced transcriptional activity. In these studies, we use SRC-1, a coactivator for steroid hormone receptors, and examine its ability to enhance the ligand-dependent interaction of the amino- and carboxyl-terminal regions of ER, resulting in a more potent transcriptional response to estrogen.

Abbreviations: ER, estrogen receptor; AF, activation function; E₂, estradiol; TOT, *trans*-hydroxytamoxifen; SRC-1, steroid receptor coactivator-1; ERE, estrogen response element; CHO, Chinese hamster ovary; CAT, chloramphenicol acetyltransferase.

§To whom reprint requests should be addressed at: Department of Molecular and Integrative Physiology, University of Illinois, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, IL 61801-3704. e-mail: katzenel@uiuc.edu.

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. §1734 solely to indicate this fact.

MATERIALS AND METHODS

Chemicals and Materials. Cell culture media were purchased from GIBCO. Calf serum was from HyClone and fetal calf serum was from Sigma. ^{14}C -Chloramphenicol (50–60 Ci/mmol; 1 Ci = 37 GBq) was from DuPont/NEN.

Plasmids. The ER expression vectors (pCMV5-her) for full-length wild-type human ER (amino acids 1–595) and ER derivatives ABCD (amino acids 1–378), M109 (amino acids 109–595), M109CD (amino acids 109–378), EF (amino acids 312–595), and EF-VP16, were constructed as described (12). An expression vector encoding SRC-1 and an empty expression vector that lacks the SRC-1 cDNA have been described (13). ER-VP16 and M109-VP16 were generated by replacing the *BsmI*/*Bam*HI fragment of pCMV-her or pCMV-her(M109), respectively, with a PCR-generated fragment encoding 78 aa of the VP16 activation domain containing *BsmI*/*Bam*HI sites. The estrogen response element (ERE)-containing reporter plasmids were (ERE)₃-pS2-CAT, constructed as described (12), and (ERE)₄-TATA-CAT, which was provided by David J. Shapiro of the University of Illinois. Either the plasmid pCH110 (Pharmacia) or pCMV β (Clontech), which contains the β -galactosidase gene, was used as an internal control for transfection efficiency. pTZ19R carrier DNA was from Pharmacia.

Cell Culture and Transient Transfections. Chinese hamster ovary (CHO) cells were maintained and transfected as described (16). Cells were transiently transfected by CaPO₄ coprecipitation method and were given 400 μl of precipitate containing the following: either 10 ng of wild-type ER, ER-VP16, M109, or M109-VP16 or 500 ng of each ER-derivative expression vector; 2.0 μg of (ERE)₄-TATA-CAT reporter plasmid; 0.3 μg of pCH110 internal control plasmid; up to 6.0 μg of SRC-1 expression vector or empty vector; and pTZ19R carrier DNA to a total of 10 μg of DNA. After 12–16 h, cells were shocked with 20% glycerol/Hanks' balanced salt solution (HBSS) for 1.5 min, rinsed with HBSS, and given fresh medium and hormone treatment as indicated. 3T3 mouse fibroblast cells were maintained and transfected as described (12, 17). Cells were harvested 24 h after glycerol shock and hormone treatment, and extracts were prepared in 200 μl of 250 mM Tris-HCl (pH 7.5) using three freeze-thaw cycles. β -Galactosidase activity was measured to normalize for transfection efficiency and chloramphenicol acetyltransferase (CAT) assays were performed as described (16).

RESULTS

The present study was designed to aid in understanding how SRC-1 increases transcriptional activity of the ER and to determine if this involved enhancing the integration of activities of the two AFs of the receptor located in the amino- and carboxyl-terminal regions. The schematic in Fig. 1 shows the ER derivatives used in our studies. We first tested the effect of exogenous SRC-1 on the transcriptional activity of the full-length receptor in ER-negative CHO cells (Fig. 2). When expressed in cells in the absence of added SRC-1, the wild-type ER was able to induce transactivation of an ERE-containing CAT reporter gene \approx 12-fold in the presence of E₂. No transcriptional activation was observed with the wild-type ER upon treatment with the antiestrogen TOT. When SRC-1 was expressed alone in cells in the absence of ER, it was unable to evoke transcription in the presence or absence of any hormone treatment tested. However, when SRC-1 was coexpressed in increasing amounts along with wild-type ER, it enhanced transcriptional activity nearly 5-fold in the presence of E₂. No transcriptional activity was observed with TOT treatment even with high levels of SRC-1. In addition, enhancement of E₂-occupied wild-type ER transcriptional activity was due to SRC-1 and not to other elements in the plasmid, as there was

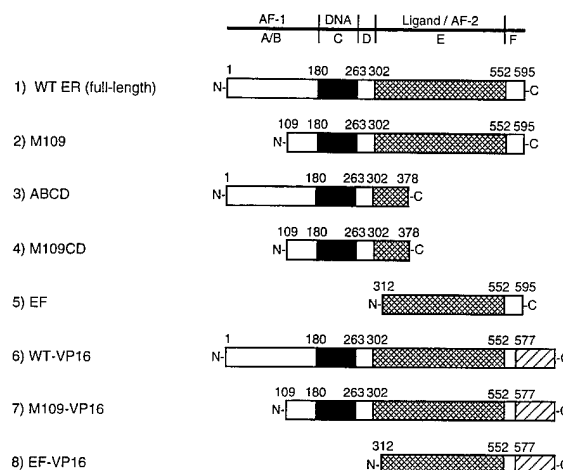


FIG. 1. Structure of ER derivatives used in this study. The structural domains of ER (A/B, C, D, E, and F), as well as the AF-1, AF-2, DNA-binding (solid boxes) and ligand-binding (cross-hatched boxes) functional domains, are shown above the schematics for the receptors. Hatched boxes represent the VP16 activation domain (residues 413–490).

no change in E₂-stimulated activity of wild-type ER when cotransfections used an empty expression vector lacking the SRC-1 cDNA (data not shown). The enhancement of E₂-dependent transcriptional activity of the ER with increasing amounts of SRC-1 implies that SRC-1 is a coactivator for E₂-dependent activity of ER, consistent with previous studies conducted in HeLa cells (13).

We then tested the ability of SRC-1 to enhance the transcriptionally productive interaction between the AF-1-containing, DNA-binding (ABCD) and the AF-2-containing, hormone-binding (EF) regions of ER (Fig. 3). Coexpression of SRC-1 with either ABCD or EF alone did not stimulate transcription of the reporter gene. When the ABCD and EF polypeptides were coexpressed in CHO cells in the absence of

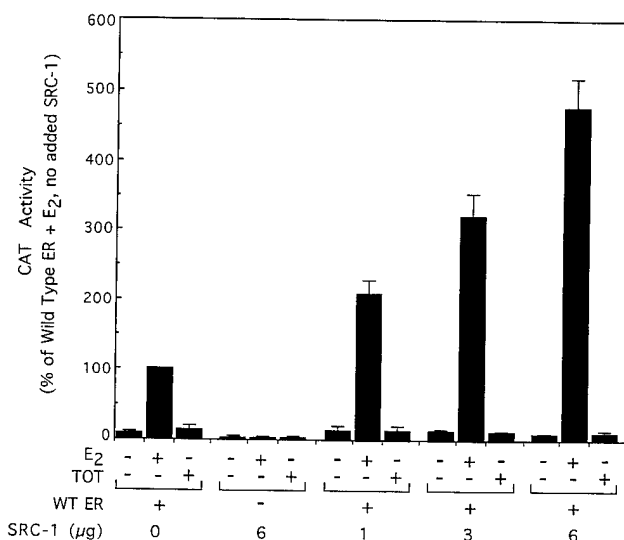


FIG. 2. Enhancement of wild-type ER transcriptional activity by SRC-1. ER-negative CHO cells were transfected with expression vectors for wild-type (WT) ER and SRC-1 as indicated, an internal control β -galactosidase plasmid, and an ERE-TATA-CAT reporter. Cells were treated with control (0.1% ethanol) vehicle, 10 nM E₂, or 1 μM TOT for 24 h. CAT activity was normalized for β -galactosidase activity from an internal control plasmid and analyzed. The CAT activity observed with wild-type ER plus E₂ but no added SRC-1 is set at 100%. Error bars represent the mean \pm SEM for three or more determinations. Some error bars are too small to be visible.

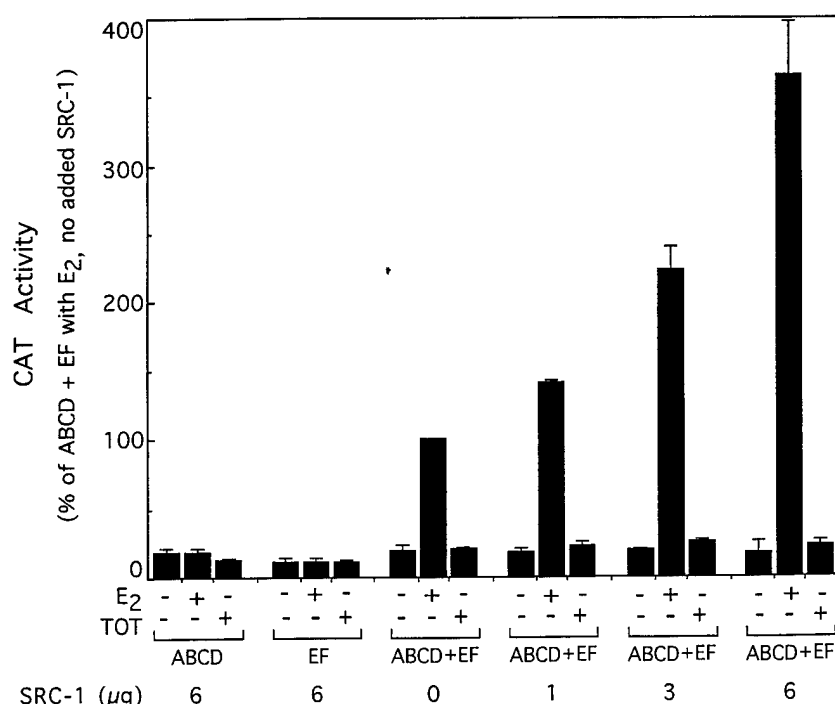


FIG. 3. E₂-dependent enhancement of the transcriptional activity of the amino- and carboxyl-terminal regions of ER by SRC-1. CHO cells were transfected with expression vectors for ER derivatives ABCD, EF, and SRC-1, as indicated, and an ERE-TATA-CAT reporter. Cells were treated with control vehicle, 10 nM E₂, or 1 μM TOT, and CAT activity, normalized for internal control β-galactosidase activity, was analyzed as described in the legend to Fig. 2.

added SRC-1, they were capable of interacting in a transcriptionally productive manner only in the presence of E₂, reconstituting ≈30% of the full-length receptor activity. When SRC-1 was coexpressed in increasing amounts with ABCD and EF, it enhanced the E₂-dependent, transcriptionally productive interaction without inducing any transcription in the absence of hormone or in the presence of TOT. These results show that coexpression of SRC-1 results in a significant increase in the transcriptional activity generated by the assembly of ABCD and EF in the presence of E₂ and not TOT, similar to the effects of SRC-1 on the full-length receptor seen in Fig. 2.

To determine if SRC-1 enhances integration of the transactivating functions of the amino- and carboxyl-terminal regions of ER, we coexpressed SRC-1 with ABCD and EF-VP16. The EF-VP16 fusion protein contains domains E and F of the human ER linked to the activation domain of the viral protein 16 (18). The constitutively active VP16 activation domain allows the detection of an interaction between ABCD and EF, even if the interaction is not transcriptionally productive. As shown in Fig. 4, coexpression of SRC-1 with either ABCD or EF-VP16 did not result in any significant transcriptional activity. When ABCD and EF-VP16 were expressed together in cells, stimulation of transcriptional activity was observed upon treatment with E₂ and to a lesser extent, TOT, indicating an interaction between ABCD and EF-VP16 in the presence of E₂ and TOT. However, when SRC-1 was coexpressed with ABCD and EF-VP16, the activity in the presence of E₂ and TOT was enhanced to ≈7-fold and ≈5-fold, respectively, above that in the absence of added SRC-1, and the enhancement occurred in an SRC-1 dose-dependent manner. In addition, when an amino-terminally truncated version of ABCD (M109CD), which lacks most of the A/B domain (i.e., lacks the first 108 aa of the receptor), was used in place of ABCD, it was unable to associate with EF-VP16 even at high levels of SRC-1, indicating that SRC-1 enhancement of ABCD and EF-VP16 activity requires an intact AF-1 region.

Similar results were obtained in the ER-negative 3T3 mouse fibroblast cell line using a different ERE-containing reporter (3ERE-pS2-CAT), where the association of the amino- and carboxyl-terminal regions of ER was enhanced ≈3-fold in the presence of E₂ or TOT with 3 or 6 μg of SRC-1 (data not presented). The magnitude of enhancement was less in the 3T3 cells compared with the CHO cells, possibly indicating higher levels of endogenous SRC-1 in the 3T3 cells.

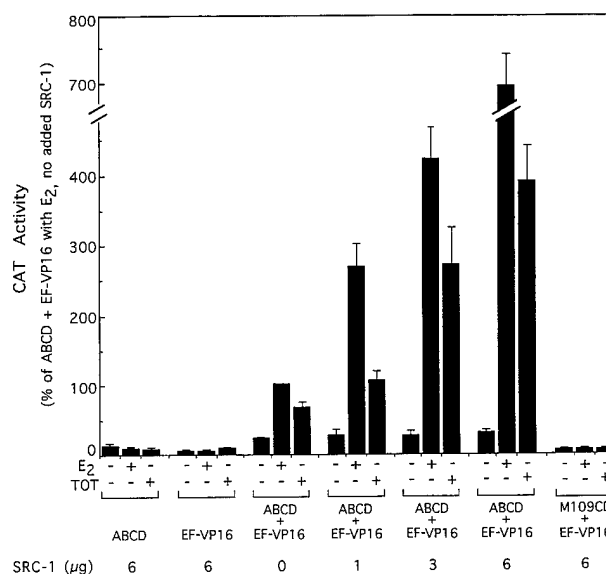


FIG. 4. Enhancement of the interaction of the amino- and carboxyl-terminal regions of ER by SRC-1. CHO cells were transfected with expression vectors for ER derivatives ABCD, M109CD, EF-VP16, and SRC-1, as indicated, and an ERE-TATA-CAT reporter. Cells were treated with control vehicle, 10 nM E₂, or 1 μM TOT, and CAT activity, normalized for internal control β-galactosidase activity, was measured as described in the legend to Fig. 2.

We also compared the effect of SRC-1 on transcriptional activity of the full-length ER or the full-length ER linked to the VP16 activation domain (ER-VP16) in the presence of E_2 or TOT. As expected, the E_2 -dependent transcriptional activity of wild-type ER was enhanced by the coexpression of SRC-1 (Fig. 5A Left). In contrast to the wild-type ER, ER-VP16 alone stimulated substantial transcription in the absence of hormone (Fig. 5A Right), and this transcriptional activity was not enhanced by coexpression of SRC-1. With E_2 in the absence of added SRC-1, ER-VP16 activity was twice that seen with no hormone addition, indicating that ER-VP16 is brought more effectively to the DNA when it is liganded. SRC-1 enhanced ER-VP16 transcriptional activity in the presence of E_2 , and the ≈ 4 -fold enhancement by SRC-1 was similar in magnitude to that seen with the E_2 -occupied wild-type ER. These results suggest that the increased transcription by ER-VP16 with E_2 is likely due to transcriptional enhancement of ER AF-1/AF-2 activity by SRC-1. In the presence of TOT, no transcriptional enhancement was observed when ER-VP16 was coexpressed with SRC-1. Since there is no transcription by AF-1 and AF-2 in the presence of TOT, it is perhaps not surprising that SRC-1 does not affect ER-VP16 liganded with TOT. Together, these results indicate that in this cellular context, an E_2 -ER complex is needed for SRC-1 enhancement, and the VP16 activation domain was not significantly affected by SRC-1. The lack of enhancement of the VP16 activation domain by SRC-1 was not likely due to competition for limiting cellular factors required for transcription, as similar results were obtained using significantly lower (i.e., 10- or 20-fold lower) levels of ER-VP16 expression plasmid (data not shown).

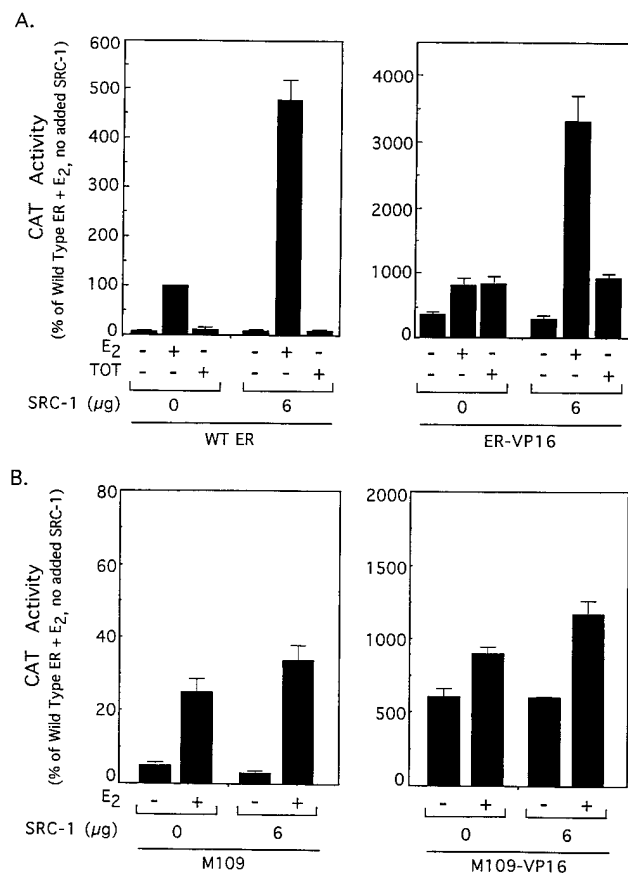


FIG. 5. Effects of SRC-1 on ER-VP16 fusion proteins. CHO cells were transfected with expression vectors for (A) wild-type ER or ER-VP16 or (B) M109 or M109-VP16, SRC-1, as indicated, and an ERE-TATA-CAT reporter. Cells were treated with control vehicle, 10 nM E_2 , or 1 μ M TOT, and CAT activity was analyzed as described in the legend to Fig. 2.

In related studies, we used the ER mutant, M109, which lacks most (the first 108 aa) of the A/B domain. M109 was transcriptionally impaired compared with the wild-type ER, stimulating only $\approx 30\%$ of wild-type ER activity in the presence of E_2 (Fig. 5B Left). Upon coexpression of SRC-1, there was minimal change in the E_2 -dependent transcriptional activity of M109. Similar results were obtained with M109-VP16 (Fig. 5B Right) in that there was little enhancement of E_2 -dependent transcription upon coexpression of SRC-1. Therefore, in this cell system, deletion of AF-1 nearly fully abolished the enhancement of receptor activity by SRC-1 with both M109 and M109-VP16 in the presence of E_2 . Presumably, SRC-1 still interacts with these A/B deletion receptors through the intact AF-2 region; however, the transcriptional enhancement of ER by SRC-1 requires an intact AF-1 containing A/B domain.

DISCUSSION

Our results provide one potential mechanism by which coactivators promote the full transcriptional activity of ER. The enhancement of a transcriptionally productive association of the amino- and carboxyl-terminal regions of ER through the influence of SRC-1 may be an essential step in activated transcription by hormone-occupied ER. Because of the complexity of receptor-mediated transcription, the detailed events that lead to hormone-dependent transactivation are not yet well understood. However, it is known that, after hormone binding, the ER undergoes a conformational change that is thought to allow the displacement of repressor proteins associated with the ER and to make the receptor accessible for interaction with coactivators (19, 20). The activated receptor has been postulated to aid in the stabilization of the preinitiation complex (3, 20, 21) and to play a role in the alteration of chromatin structure (1-3, 22). Our studies investigate two important aspects leading to ER-mediated transcription—namely, the conformational change in ER that is induced by ligand binding and the interaction of ER with coactivators. In this report, we have demonstrated that the ligand-induced conformational change promotes the interaction between the amino- and carboxyl-terminal regions of ER, when expressed as separate polypeptides in cells, and that this interaction is facilitated by the coactivator SRC-1. The next step, enhancement of transcriptional activity by SRC-1, requires that the ER be liganded with hormone (E_2), and not antihormone (TOT), for the integrated functions of the AF-1- and AF-2-containing regions of the ER to be transcriptionally productive. These results help in providing a clearer picture of the molecular events that occur after ligand binding to result in an activated receptor.

SRC-1 was first isolated through its ability to bind to the AF-2-containing, ligand-binding domain of progesterone receptor (13). Our results suggest that SRC-1 can act, at least in part, to functionally enhance ER activity by promoting the association between the amino- and carboxyl-terminal regions of ER. SRC-1 did not stimulate TOT-dependent wild-type ER activity and did not promote the transcriptionally productive assembly of ABCD and EF in the presence of TOT, because AF-2 is not functional when liganded with TOT (7, 23). However, SRC-1 did evoke increased activity measured with ABCD and EF-VP16 in the presence of TOT (Fig. 4 versus Fig. 3), indicating that SRC-1 promotes the functional interaction of ABCD and EF-VP16. The absence of SRC-1 stimulation of full-length ER activity when occupied with TOT highlights the important role of ligand character in the response of the receptor to SRC-1. In the cellular contexts examined, SRC-1 enhanced transcriptional effectiveness only of the E_2 -AF-1/AF-2 complex, perhaps by facilitating the interaction of the two AF-containing regions of the receptor with the basal transcription complex.

Multiple proteins have been identified which interact with ER in a ligand-dependent manner (15, 24–27); however, most have not yet been shown to enhance ER-stimulated transcription. An exception is the cAMP response element-binding protein (CREB) coactivator, CREB-binding protein (CBP), another recently reported coactivator for the steroid receptor superfamily (14). SRC-1 has been shown to significantly increase the transcriptional activity of progesterone receptor and other steroid hormone receptors, including ER. Potentially, SRC-1 may function to enhance the transcription of other members of the steroid hormone receptor superfamily by a mechanism analogous to our findings. The conservation of an amino- and a carboxyl-terminal activation domain among steroid hormone receptors (2, 3) and the ability of SRC-1 to act as a coactivator for several steroid hormone receptors together suggest a general mechanism for coactivator action on steroid hormone receptors that may involve facilitation of the productive association of the two AF containing regions of these receptors, enabling optimal stimulation of transcription. At present, however, we do not have evidence that the functional interaction of AF-1 and AF-2 promoted by SRC-1 is direct. In fact, the receptor complex appears to include at least SRC-1 and CBP, and the complexity is likely to grow with the verification of functional interactions of other receptor binding proteins. Any one of these molecules could interact with the receptor, directly or indirectly, to promote the cooperative actions of AF-1 and AF-2. Continued investigation of steroid hormone receptor–coactivator complexes and their interaction with the transcription apparatus should aid in elucidating further aspects of the detailed biochemical mechanism of activated transcription.

We thank W. Lee Kraus for making some of the ER expression vectors. We also thank Byron Kemper, David Shapiro, and John Katzenellenbogen for helpful discussions and critical reading of the manuscript. This research was supported by National Institutes of Health Grants CA18119 and CA60514 (to B.S.K.) and a National Institutes of Health grant to B.W.O.

1. Evans, R. M. (1988) *Science* **240**, 889–895.
2. Mangelsdorf, D. J., Thummel, C., Beato, M., Herrlich, P., Schutz, G., Umesono, K., Blumberg, B., Kastner, P., Mark, M., Chambon, P. & Evans, R. M. (1995) *Cell* **83**, 835–839.
3. Tsai, M. J. & O'Malley, B. W. (1994) *Annu. Rev. Biochem.* **63**, 451–486.
4. Kumar, V., Green, S., Stack, G., Berry, M., Jin, J. R. & Chambon, P. (1987) *Cell* **51**, 941–951.
5. Kumar, V., Green, S., Staub, A. & Chambon, P. (1986) *EMBO J.* **5**, 2231–2236.
6. Lees, J. A., Fawell, S. E. & Parker, M. G. (1989) *Nucleic Acids Res.* **17**, 5477–5487.
7. Tora, L., White, J., Brou, C., Tasset, D., Webster, N., Scheer, E. & Chambon, P. (1989) *Cell* **59**, 477–487.
8. Webster, N. J., Green, S., Jin, J. R. & Chambon, P. (1988) *Cell* **54**, 199–207.
9. Tasset, D., Tora, L., Fromental, C., Scheer, E. & Chambon, P. (1990) *Cell* **62**, 1177–1187.
10. Tzukerman, M. T., Esty, A., Santiso-Mere, D., Danielian, P., Parker, M. G., Stein, R. B., Pike, J. W. & McDonnell, D. P. (1994) *Mol. Endocrinol.* **8**, 21–30.
11. Pham, T. A., Hwung, Y. P., Santiso-Mere, D., McDonnell, D. P. & O'Malley, B. W. (1992) *Mol. Endocrinol.* **6**, 1043–1050.
12. Kraus, W. L., McInerney, E. M. & Katzenellenbogen, B. S. (1995) *Proc. Natl. Acad. Sci. USA* **92**, 12314–12318.
13. Oñate, S. A., Tsai, S. Y., Tsai, M. J. & O'Malley, B. W. (1995) *Science* **270**, 1354–1357.
14. Kamei, Y., Xu, L., Heinzel, T., Torchia, J., Kurokawa, R., Glass, B., Lin, S.-C., Heyman, R. A., Rose, D. W., Glass, C. K. & Rosenfeld, M. G. (1996) *Cell* **85**, 403–414.
15. Halachmi, S., Marden, E., Martin, G., MacKay, H., Abbondanza, C. & Brown, M. (1994) *Science* **264**, 1455–1458.
16. Reese, J. C. & Katzenellenbogen, B. S. (1991) *J. Biol. Chem.* **266**, 10880–10887.
17. Montano, M. M., Muller, V., Trobaugh, A. & Katzenellenbogen, B. S. (1995) *Mol. Endocrinol.* **9**, 814–825.
18. Sadowski, I., Ma, J., Triezenberg, S. & Ptashne, M. (1988) *Nature (London)* **335**, 563–564.
19. McDonnell, D. P., Clemm, D. L., Hermann, T., Goldman, M. E. & Pike, J. W. (1995) *Mol. Endocrinol.* **9**, 659–669.
20. Beekman, J. M., Allan, G. F., Tsai, S. Y. & O'Malley, B. W. (1993) *Mol. Endocrinol.* **7**, 1266–1274.
21. Klein-Hitpass, L., Tsai, S. Y., Weigel, N. L., Allan, G. F., Riley, D., Rodriguez, R., Schrader, W. T., Tsai, M. J. & O'Malley, B. W. (1990) *Cell* **60**, 247–257.
22. Pham, T. A., Hwung, Y. P., McDonnell, D. P. & O'Malley, B. W. (1991) *J. Biol. Chem.* **266**, 18179–18187.
23. Berry, M., Metzger, D. & Chambon, P. (1990) *EMBO J.* **9**, 2811–2818.
24. Cavailles, V., Dauvois, S., Danielian, P. S. & Parker, M. G. (1994) *Proc. Natl. Acad. Sci. USA* **91**, 10009–10013.
25. LeDourarin, B., Zechel, C., Garnier, J.-M., Lutz, Y., Tora, L., Pierrat, B., Heery, D., Gronemeyer, H., Chambon, P. & Losson, R. (1995) *EMBO J.* **14**, 2020–2033.
26. Cavailles, V., Dauvois, S., L'Horsset, F., Lopez, G., Hoare, S., Kushner, P. J. & Parker, M. G. (1995) *EMBO J.* **14**, 3741–3751.
27. Katzenellenbogen, J. A., O'Malley, B. W. & Katzenellenbogen, B. S. (1996) *Mol. Endocrinol.* **10**, 119–131.

Constitutively Active Human Estrogen Receptors Containing Amino Acid Substitutions for Tyrosine 537 in the Receptor Protein

Karen E. Weis, Kirk Ekena, James A. Thomas,
Gwendal Lazennec, and Benita S. Katzenellenbogen

Department of Molecular and Integrative Physiology
(K.E.W., K.E., G.L., B.S.K.)
Department of Cell and Structural Biology (J.A.T., B.S.K.)
University of Illinois and College of Medicine
Urbana, Illinois 61801

To better understand structure-activity relationships in the human estrogen receptor (ER), we examined the role of tyrosine 537 in the transcriptional response of the receptor, since this residue is close to a region of the hormone-binding domain shown previously to be important in hormone-dependent transcriptional activity and because this amino acid has been proposed to be a tyrosine kinase phosphorylation site important in the activity of the ER. We substituted five amino acids at this position (alanine, phenylalanine, glutamic acid, lysine, or serine) and screened these mutants for their biological activities in the presence and absence of estradiol. Two of the ER mutants, Y537A and Y537S, displayed estrogen-independent constitutive activity that was approximately 20% or 100%, respectively, of the activity of the wild type receptor with estradiol, when assessed in two different cell backgrounds using three different estrogen-responsive promoters. In some circumstances, the Y537E and Y537K proteins also exhibited some low level of constitutive activity. The constitutive activity of the mutants, as well as their activity in the presence of E_2 , was fully suppressed by antiestrogen. The extent of interaction of the constitutively active ERs with the steroid receptor coactivator-1 (SRC-1) closely parallel the magnitude of transcriptional activity of the receptor. Whereas wild type ER showed interaction with SRC-1 only in the presence of estrogen, Y537A and Y537S ER showed moderate or full interaction in the absence of ligand, an interaction that was blocked by antiestrogen, and the magnitude of interaction was increased to or remained at 100% upon estradiol treatment, implying that the ability

of an ER to associate with SRC-1 is a good indicator of a transcriptionally active conformational state of the receptor. Our findings indicate that tyrosine 537 is in a region important in the ligand regulation of ER transcriptional activity and that the presence of certain amino acids at this position can shift ER into a conformation that is active even without ligand. However, tyrosine is not required at this site for estrogen binding or transcriptional response to estrogen in the systems investigated. Our findings, interpreted in light of the recently published x-ray crystal structure of the ligand-binding domains of three related receptors of the nuclear receptor superfamily, suggest that some of the amino acid substitutions introduced at position 537 may facilitate the shift of helix 12 of the ER into an active conformation and/or allow for differential stabilization of the receptor in its active form. (*Molecular Endocrinology* 10: 1388–1398, 1996)

INTRODUCTION

The human estrogen receptor (ER) and related members of the nuclear steroid receptor superfamily regulate the complex pathway of transcriptional activation for many biologically important genes. Upon ligand binding, ER undergoes a conformational change allowing the receptor to activate transcription of target genes (1–5). Numerous factors regulate the activity of ER, such as the nature of the ligand bound to the receptor, the phosphorylation state of the ER, and interactions with coactivator proteins (4–7).

The ER is comprised of several functionally distinct domains (2, 8–12). The N-terminal A/B domain contains the transcription activation function-1 (AF-1). The highly conserved C domain is the site of DNA binding to estrogen-response elements, whereas the D do-

main appears to function as a hinge region. Domains E and F, at the C terminus of the receptor, are the regions of ligand binding and recognition and contain the ligand-dependent transcription activation function-2 (AF-2).

We have been interested in ER structure-activity relationships and, particularly, in identifying regions of domain E crucial in ligand binding and interpretation (7, 13). Because ER bioactivity is also known to be significantly regulated by phosphorylation, we and others (14–23) have examined the role of some serine and tyrosine residues as sites of phosphorylation. Recently, by alanine-scanning mutagenesis across a 21-amino acid region from residue 515 to 535 in the ER hormone-binding domain, we identified several amino acids between 520 and 530 as being sites of contact between ER and the hormone estradiol (E_2) (24). Because tyrosine phosphorylation may be important in the activity of the ER and, in particular, in the ability of growth factors such as insulin-like growth factor I and epidermal growth factor to synergize with estrogen and enhance ER transcriptional activity (25–30), we have, in this report, extended our structure-function analysis of the ER to examine tyrosine 537 and its potential role in the bioactivity of the ER. In addition to substituting alanine for this tyrosine, we also substituted four other amino acids at this position and screened the mutants for their activities in the presence and absence of estradiol. The amino acid substitutions for tyrosine 537 were as follows: alanine, a relatively conservative substitution; phenylalanine, the most conservative change from tyrosine; glutamic acid, which mimics tyrosine phosphorylation in providing the same charge on the receptor as phosphorylation; lysine, an opposite charge from phosphorylation; and serine, a different, potentially phosphorylatable residue. Several of the ER mutants we generated displayed estrogen-independent constitutive transcriptional activity and estrogen-independent association with the nuclear receptor coactivator SRC-1 and, in all cases, the receptors were capable of good transcriptional activity in the presence of estradiol. Our findings are interpreted in light of the recently published x-ray crystal structures of the ligand-binding domains of three related receptors of the nuclear receptor superfamily (31–34).

RESULTS

Ligand-Dependent and Ligand-Independent Transcriptional Activities of Tyrosine 537 Mutant Receptors

Using mutant oligonucleotides, we prepared ERs containing five different amino acid substitutions for tyrosine at position 537 of the human ER (hER). All ER mutations were confirmed by restriction digests and dideoxy-nucleotide sequencing. To assess transcriptional ability of the Y537A mutant ER in which alanine

was substituted for tyrosine, we transiently transfected ER-negative MDA-MB-231 breast cancer cells with Y537A ER expression vector and an estrogen-responsive promoter-reporter construct, 2ERE-pS2-CAT, containing two estrogen-response elements, the pS2 gene promoter, and the chloramphenicol acetyl transferase (CAT) reporter gene. Cells were treated with either control ethanol vehicle, estradiol (E_2), or the antiestrogen *trans*-hydroxytamoxifen (TOT), and CAT activity was measured. For these assays we used 1×10^{-8} M E_2 and 100 ng ER expression vector, since wild type ER reached maximal activity at this concentration of E_2 , and under these conditions the level of activation was independent of the amount of transfected ER DNA over the range of 50–400 ng (data not shown). CAT activity was very low in wild type receptor treated with control 0.1% ethanol vehicle and was induced 100- to 200-fold by the addition of E_2 (Fig. 1).

The Y537A mutant exhibited activity quite similar to the wild type ER over a range of E_2 concentrations (Fig. 1). Interestingly, however, the alanine mutant also possessed some constitutive activity in the absence of ligand, approximately 20% of the maximal activity achieved by the wild type ER with E_2 treatment. Using the minimal TATA promoter, in a 2ERE-TATA-CAT reporter gene, we observed a similar level of constitutive activity from Y537A expressed in Chinese hamster ovary (CHO) cells (data not shown). Consistent with the dose-response profiles seen in Fig. 1, the Y537A ER demonstrated very similar E_2 binding affinity compared with wild type ER. From radiolabeled E_2 hormone-binding assays performed over a broad range of hormone concentrations (3×10^{-11} M to 2×10^{-8} M), calculated equilibrium dissociation constants (K_d) were 0.27 nM and 0.29 nM for Y537A and wild type ER, respectively, similar to previously reported values for wild type ER (24, 35).

We next compared the transcriptional activity of Y537A with receptors in which four different amino acids were substituted for tyrosine 537 in the ER (Y537F, Y537K, Y537E, and Y537S). Assays were conducted using several different promoter and cell backgrounds. Initially, the mutant ERs were screened in MDA-MB-231 cells using the estrogen-responsive 2ERE-pS2-CAT gene construct. Transfected cells were treated with control ethanol vehicle, E_2 at 1×10^{-8} M, TOT at 1×10^{-6} M, or with both E_2 and TOT (Fig. 2). All of the receptors showed good transactivation activity in the presence of E_2 ; whereas Y537K, Y537E, Y537A, and Y537S reached full wild type activation, Y537F reached only 70% of wild type activity. Unexpectedly, the Y537S mutant was fully active in the absence of ligand, making it a stronger constitutively active mutant than Y537A. Treatment with TOT alone blocked the constitutive activity seen for both the Y537A and Y537S mutants, and all of the receptors demonstrated antagonism of E_2 induction by TOT. Similar antagonism of receptor transactivation was observed with the pure antiestrogen ICI 164,384 (data not shown).

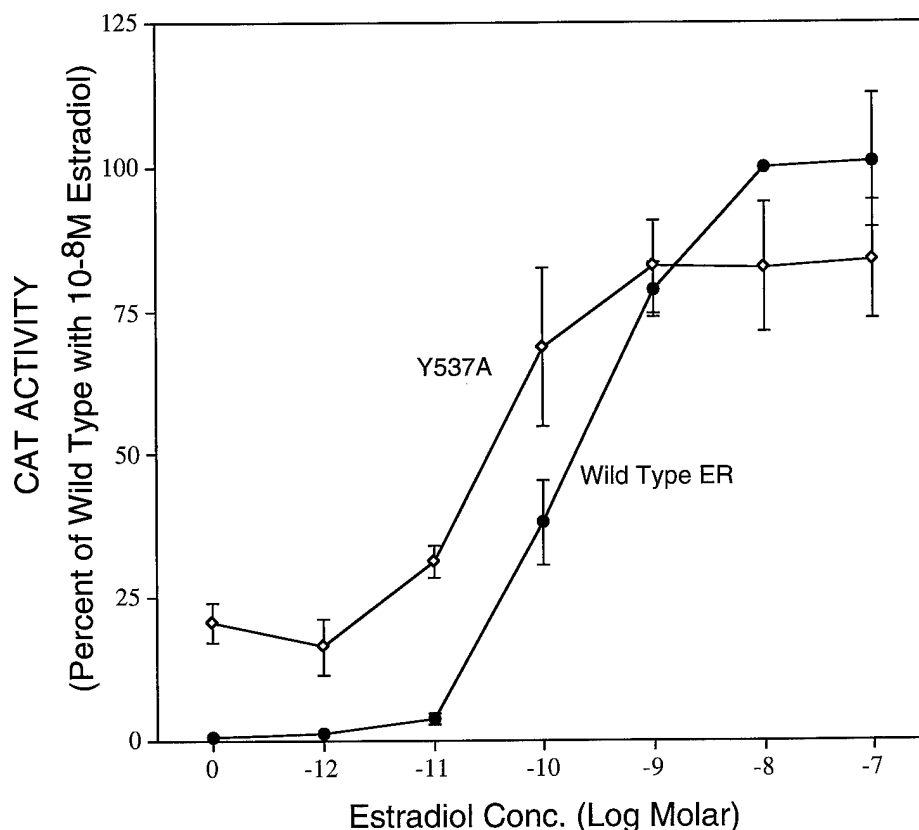


Fig. 1. Transactivation Ability of Y537A ER vs. Wild Type (wt) ER in the Presence and Absence of E_2

MDA-MB-231 cells were transfected with Y537A or wild type ER expression vector, 2ERE-pS2-CAT reporter plasmid, and a β -galactosidase internal reporter to correct for transfection efficiency. Before harvesting, cells were treated for 24 h with E_2 from $1 \times 10^{-12}M$ to $1 \times 10^{-7}M$ or with control (0.1% ethanol) vehicle (zero E_2). Transactivation was determined by CAT activity, normalized to the internal β -galactosidase control, and is expressed as percent of wild type activity at $1 \times 10^{-8}M E_2$. Each point represents the mean \pm SD of determinations from two to four individual experiments.

We further examined all of the Y537 mutants in 231 cells using a different promoter, namely the thymidine kinase driven-CAT construct, 2ERE-TK-CAT (Fig. 3). Once again, the Y537A and Y537S mutants exhibited substantial ligand-independent transactivation. In the absence of ligand, the Y537A and Y537S mutants showed $\sim 30\%$ and $\sim 120\%$ of wild type E_2 -stimulated activity, respectively. Moreover, the Y537K and Y537E mutants also showed statistically significant activity in the absence of added ligand (~ 10 - 15% of wild type ER + $1 \times 10^{-8}M E_2$). In the presence of $1 \times 10^{-8}M E_2$, all mutant receptors showed activity similar to that of wild type ER. Treatment with TOT at $1 \times 10^{-6}M$ brought transcriptional activity of all the unliganded mutant receptors or E_2 -occupied receptors (data not shown) to the low level observed for the wild type receptor treated with TOT.

Y537S ER Shows Full Constitutive Transcriptional Activity Over a Broad Range of Receptor Concentrations

As shown in Fig. 4, the Y537S receptor showed E_2 -independent constitutive activity over a very

broad range of transfected ER plasmid. At all amounts of ER plasmid used, the Y537S receptor without any added ligand showed transcriptional activity indistinguishable in magnitude from that of the wild type receptor plus E_2 . These curves were also the same as that obtained for Y537S plus E_2 , indicating that this receptor was fully active without E_2 and that treatment with E_2 did not change activity of this receptor, whereas wild type receptor was dependent on E_2 for stimulation of its transcriptional activity.

The Tyrosine 537 Mutant ERs Have Similar Phenotypes in Two Different Cell Backgrounds

To determine whether the observed activities of the mutant ERs depended on cell type, transfections were also conducted in ER-negative CHO cells (Fig. 5). In CHO cells treated with E_2 , all of the Y537 mutant receptors induced transcription to near wild type levels except for Y537F, which achieved about half-maximal activity. The Y537A and Y537S mutants displayed constitutive activity, as seen previously in 231 cells (Figs. 2 and 3); however, the magnitude of ligand-

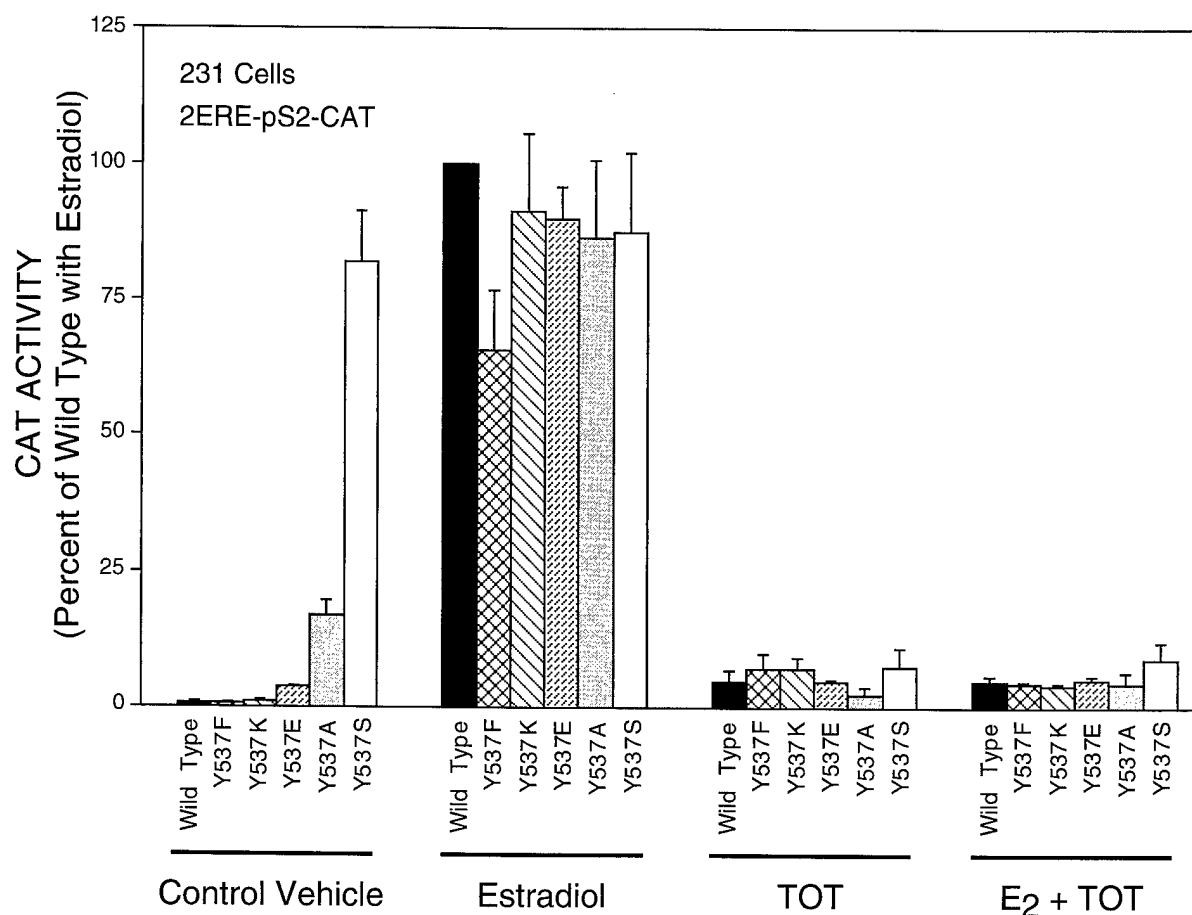


Fig. 2. Transactivation Ability of Various Y537 Mutant Receptors Treated with No Ligand or with Estrogen and Antiestrogen. MDA-MB-231 cells were transfected with wild type or mutant ER expression vector Y537F; Y537K; Y537E; Y537A or Y537S, the 2ERE-pS2-CAT reporter plasmid, and a β -galactosidase internal reporter. Before harvesting, cells were treated for 24 h with control ethanol vehicle, E_2 at $1 \times 10^{-8}M$, TOT at $1 \times 10^{-6}M$, or E_2 plus TOT at $1 \times 10^{-8}M$ and $1 \times 10^{-6}M$, respectively. Transactivation was determined by CAT activity normalized to the internal β -galactosidase control and is expressed as percent of the wild type receptor activity at $1 \times 10^{-8}M$ E_2 . Each bar represents the mean \pm SD of determinations from two to four individual experiments.

independent activity was slightly lower in the CHO cells. The addition of TOT reduced the constitutive activity of Y537A and Y537S to the level of the wild type receptor treated with TOT.

Expression of the Tyrosine 537 Mutant Proteins in Cells

To verify levels of protein expression, the Y537 mutant receptors were expressed in 231 cells and analyzed by Western blotting. Whole cell extracts were prepared, separated by SDS-PAGE, and probed with the ER-specific antibody H226, which detects an epitope in the N-terminal region of the receptor, far from the amino acid 537 region (Fig. 6). All of the mutant ERs were present at levels either equal to that of the wild type ER, or in the case of Y537K and Y537S receptors, at somewhat higher levels than those seen for the wild type receptor.

Interaction of the Constitutively Active Receptors Y537A and Y537S with the Steroid Receptor Coactivator (SRC-1) Protein

Since Y537S and Y537A showed substantial constitutive activity, we analyzed the ligand-dependent and -independent interaction of Y537A and Y537S ERs with the steroid receptor coactivator protein SRC-1, which has been shown to be a coregulator that enhances ER transactivation (36). *In vitro* transcribed and translated SRC-1 was incubated with glutathione-S-transferase (GST) fusion proteins of each mutant ER, or with wild type ER for comparison, in the presence or absence of ligand (Fig. 7).

The wild type receptor showed a distinct E_2 -dependent association with SRC-1, which was not seen by treatment with the antiestrogen TOT. Interestingly, the Y537A and Y537S mutants both exhibited ligand-independent association with SRC-1. While the Y537A

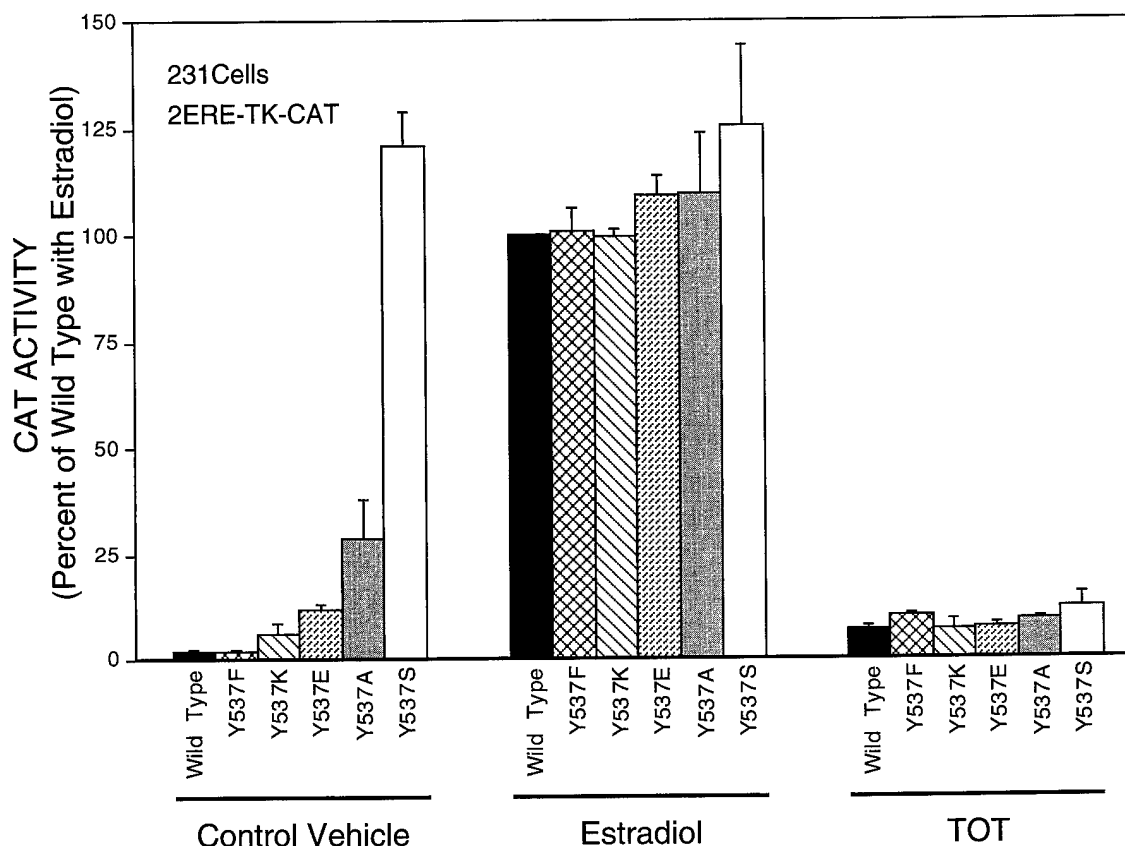


Fig. 3. Transactivation Ability of Y537 Mutant Receptors, Treated with No Ligand or with E_2 or TOT, Using an Estrogen-Responsive Thymidine Kinase Promoter Reporter Construct

MDA-MB-231 cells were transfected with wild type or mutant ER expression vector, the 2ERE-TK-CAT reporter plasmid, and a β -galactosidase internal reporter plasmid. Before harvesting, cells were treated for 24 h with control ethanol vehicle, E_2 at $1 \times 10^{-8}M$, or TOT at $1 \times 10^{-6}M$. Transactivation was determined by CAT activity normalized to the internal β -galactosidase control and is expressed as percent of the wild type receptor activity at $1 \times 10^{-8}M$ E_2 . Each bar represents the mean \pm SD of determinations from two to four individual experiments.

ER fusion protein exhibited a moderate constitutive interaction with SRC-1 in the absence of any ligand (Fig. 7, control vehicle lane), the Y537S mutant associated strongly with SRC-1 in the absence of E_2 , and treatment with the antiestrogen TOT completely eliminated this ligand-independent interaction of the Y537A and Y537S ERs. Interaction of Y537A receptor with SRC-1 was increased to that of the wild type ER in the presence of E_2 whereas treatment with E_2 did not further increase association of the Y537S receptor with SRC-1, which was already maximal. Thus there was a good correlation between magnitudes of interaction with this coregulator and transcriptional activity of these receptors.

DISCUSSION

Our findings show that position 537 of the ER is important in ligand regulation of ER transcriptional activity and that certain amino acid substitutions for ty-

rosine at this position can result in a receptor that is fully active in the absence of ligand. Notably, all amino acid changes resulted in receptors that showed good activity in the presence of E_2 . Two of the mutant ERs, Y537A and Y537S, were able to induce transcription independently of ligand to approximately 20% and 100% of wild type maximal E_2 -induced activity, respectively. The constitutive activity of Y537A and Y537S was observed in the several different cell and promoter contexts investigated, and this constitutive activity was blocked by the addition of antiestrogen. Y537S ER appears, therefore, to be in a fully active conformation while Y537A ER, which shows only partial constitutive activity, demonstrates a dose-response to E_2 that is similar to that of the wild type ER. The ligand-independent transcriptional activity of Y537S was observed over a broad range of receptor concentrations and, notably, its activity without ligand was of the same magnitude as that of wild type ER with E_2 at all expression plasmid concentrations, even very low nanogram amounts (Fig. 4). In some circum-

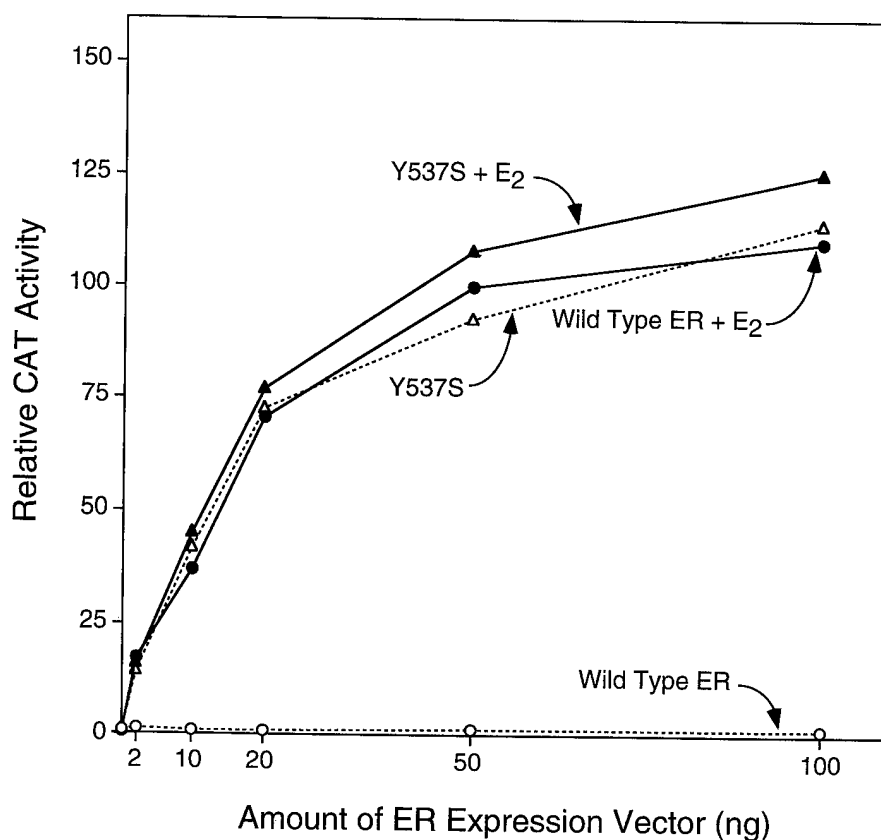


Fig. 4. Transactivation Activity of Wild Type and Y537S ERs as a Function of Amount of Transfected Receptor Expression Vector. MDA-MB-231 cells were transiently transfected with the wild type or mutant ER expression vector at the amount indicated, 2ERE-TK-CAT reporter plasmid, and a β -galactosidase internal reporter plasmid. Before harvesting, cells were treated for 24 h with control ethanol vehicle or E_2 at $1 \times 10^{-8}M$. Transactivation was determined by CAT activity normalized to the internal β -galactosidase control and is expressed as a percent of the wild type receptor activity with 100 ng wild type ER plasmid and $1 \times 10^{-8}M E_2$, which is set at 100%. Each point represents the mean of closely corresponding determinations from two individual experiments.

stances, two other mutant proteins, Y537E and Y537K, also exhibited some low level of constitutive activity.

Rather remarkably, the magnitude of estrogen-independent transcriptional activity of the Y537A and Y537S receptors paralleled very closely the magnitude of their interaction with the steroid receptor coactivator, SRC-1, in the absence of estrogen. In addition, the extent of E_2 stimulation of transactivation by these two receptors in the presence of E_2 was also mirrored in their extent of association with SRC-1, implying that the ability of an ER to associate with SRC-1 is a good indicator of a transcriptionally active conformational state of the ER, be it constitutive or ligand-induced. The presence of serine at amino acid 537 in the receptor fully shifts the receptor into an activated state, whereas alanine at this site results in only a partial achievement of this activated state. As might be expected, antiestrogen reduced both the constitutive and estrogen-stimulated transcriptional activities and SRC-1 interacting abilities of these receptors. These findings and our direct hormone-binding studies, conducted with the Y537A and wild type ERs, are consis-

tent with previous observations that amino acids most important in E_2 binding in this region of the receptor span from approximately amino acids 520 to 530 but do not extend to residues immediately carboxyl- or amino-terminal of this region (9, 13, 24, 35). Of note, none of our amino acid substitutions destroyed ER-transcriptional activity, implying considerable permissiveness in the character of the amino acid that can be tolerated at position 537. In fact, tyrosine is not conserved at this position among other members of the nuclear receptor superfamily (31–34). Phosphorylation of this particular tyrosine thus appears not to be necessary for good receptor activity, at least in the cell and promoter contexts we have examined.

To date, the three-dimensional structure of the ER has not been determined. However, by analogy to recently published crystal structures of other nuclear receptors (31–34), we can predict some features for ER. In Fig. 7, the ER amino acid sequence is displayed in alignment with human retinoic acid receptor- γ (hRAR γ), rat thyroid hormone receptor $\alpha 1$ (rTR $\alpha 1$), and human retinoid X receptor- α (hRXR α). The α -helical character of liganded RAR γ and TR $\alpha 1$ and unliganded

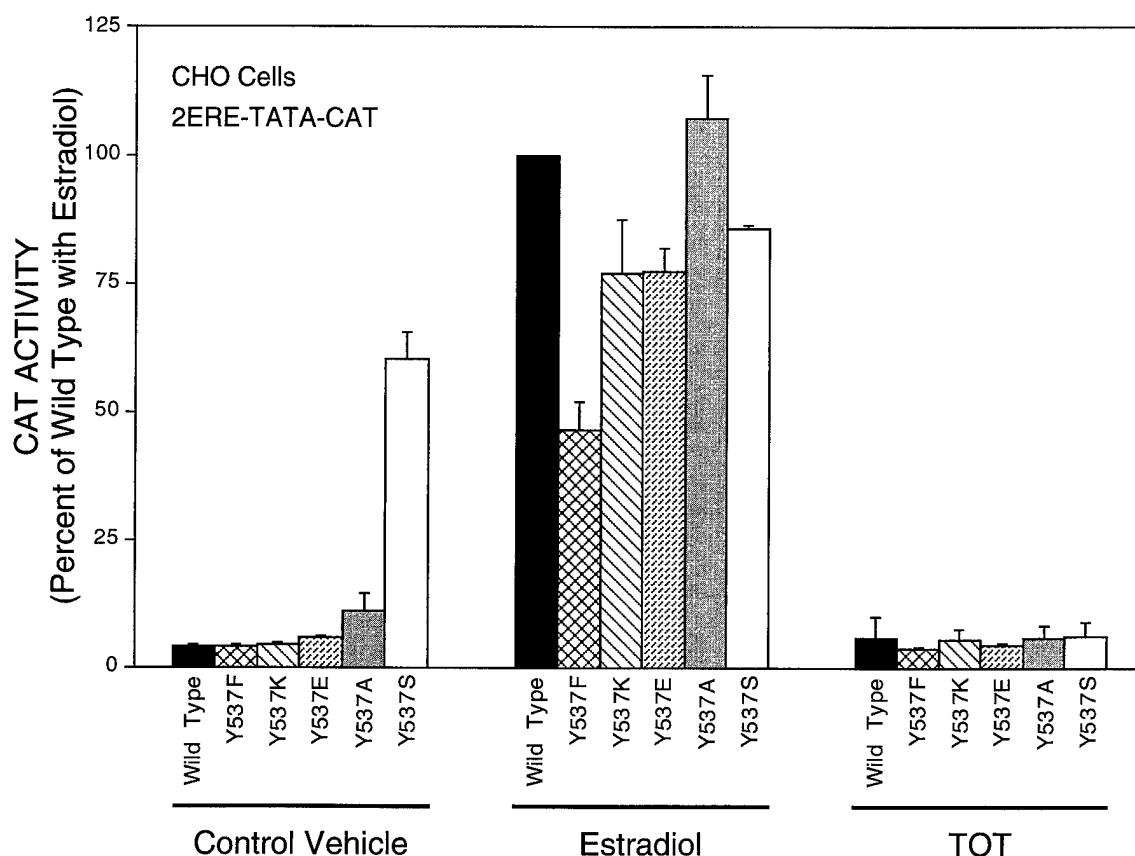


Fig. 5. Transactivation Ability of Y537 Mutant Receptors in CHO Cells with a Minimal Promoter

CHO cells were transiently transfected with wild type or mutant ER expression vector, 2ERE-TATA-CAT reporter, and a β -galactosidase internal reporter plasmid. Before harvesting, cells were treated for 24 h with ethanol control vehicle, E_2 at 1×10^{-9} M, or TOT at 1×10^{-6} M. Transactivation was determined by CAT activity normalized to the internal β -galactosidase control and is expressed as a percent of the wild type receptor activity with E_2 . Each bar represents the mean \pm SD of determinations from two to four individual experiments.

RXR α is depicted, based on their crystal structures. When aligned with the unliganded RXR α molecule, the tyrosine at 537 of ER would reside in an α -helix designated helix 12. In contrast, in RAR γ and TR α 1, structures crystallized with ligand, the location of helix 12 has shifted downstream so that the Y537 of ER now lies at the very end of a loop region, at the start of helix 12. Renaud *et al.* (32) suggest that a conformational change upon ligand binding shifts helix 12 toward the N-terminal portion of the RAR γ ligand-binding domain, creating a transcriptionally active receptor. This finding, in light of the ER amino acid sequence alignments, implies that even small modifications at position 537 might elicit alterations in the three-dimensional structure of hER that could have profound effects on the constitutive and hormone-dependent transcriptional activities of the receptor. One explanation of our data is that some of the amino acid substitutions introduced at position 537, such as Y537A and Y537S, might facilitate the shift of helix 12 into an active conformation and/or allow for differential stabilization of the receptor in its active form, enabling a transcrip-

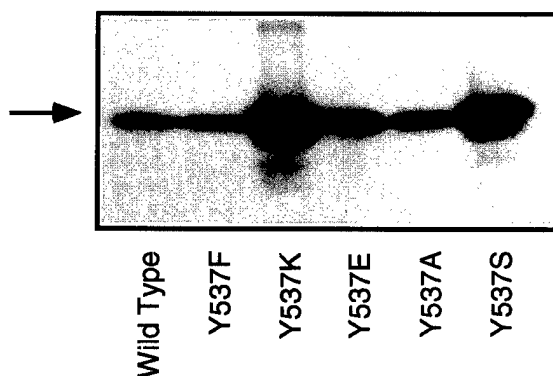


Fig. 6. Western Immunoblot Analysis of Wild Type and Y537 Mutant ER Expression Levels

Whole-cell extracts were prepared from MDA-MB-231 cells transfected with wild type or the indicated mutant ER expression plasmid. At 24 h after transfection, extracts were prepared and ~ 150 μ g of total protein were loaded per lane and separated by SDS-PAGE. The 66-kDa ER protein (denoted by arrow) was detected using the anti-ER antibody H226.



Fig. 7. Interaction of Y537A, Y537S, and Wild Type ERs with Steroid Receptor Coactivator Protein-1

SRC-1 was made by *in vitro* transcription and translation incorporating [35 S]methionine and was incubated with GST-ER fusion protein that had been adsorbed onto glutathione-Sepharose resin. Incubations were conducted in the presence of control 0.1% ethanol vehicle (C), 1×10^{-6} M estradiol (E), or 1×10^{-6} M *trans*-hydroxytamoxifen (T). Fusion proteins were GST alone (no ER), or GST fused to the ligand-binding domains (amino acids 282–595) of wild type ER, Y537A ER, or Y537S ER. After incubation for 2.5 h at 4°C, the resins were extensively washed, and retained SRC-1 was then eluted, separated by SDS-PAGE, and visualized after autoradiography.

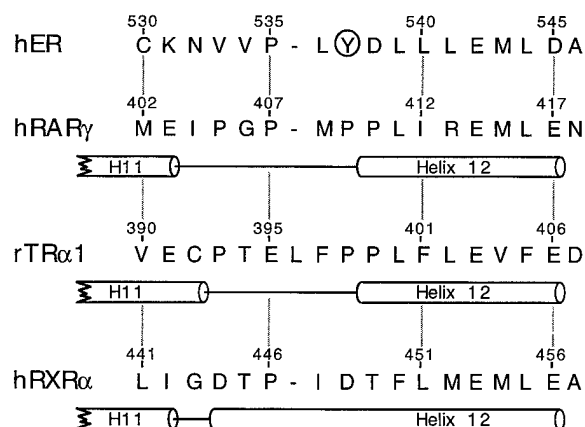


Fig. 8. Location of Residue Y537 in the ER Aligned Relative to Related Nuclear Hormone Receptors

The hER amino acid sequence was aligned to the sequences of hRAR γ , rTR α 1, and hRXR α (taken from Ref. 34). Representations of secondary structure from x-ray crystallography are shown *beneath* the amino acid sequences. Residue Y537 of the hER is *circled* to show its position relative to the α -helical region (helix 12) for liganded hRAR γ and rTR α 1, or unliganded hRXR α .

tionally active conformation even in the absence of ligand.

Constitutive transcriptional activity of nuclear receptors is a rare occurrence. If these mutant ERs were to arise due to mutations in estrogen-responsive cells, including breast or uterine cells for example, they would likely be quite detrimental. Previously, our laboratory reported on a different mutation in the ER ligand-binding domain, substitution of glutamine for glutamic acid at position 380 of the ER, which resulted in a mutant receptor displaying substantial constitutive

transcriptional activity in the absence of estrogen (37). Thus, changing the charge of E380 elicited transcriptional activity similar to that of the alanine and serine mutations at position 537 of the receptor. With regard to the hRAR γ structure, Renaud *et al.* (32) report that upon ligand binding, helix 12 is stabilized in an active conformation by an important salt bridge with residue K264. Based on sequence comparison, E380 of the ER aligns with K264 of RAR γ (34), which suggests that, in three-dimensional space, Y537 and E380 are within secondary structural elements that come together when the receptor is activated.

Although previous reports have suggested that tyrosine 537 was necessary for the ability of the receptor to bind hormone (19–21) or bind to estrogen-response element DNA in *in vitro* gel shift assays (23), our studies in intact cells do not support these conclusions. The earlier studies examined only tyrosine 537 changed to phenylalanine and were based on hormone binding from receptors made in Baculovirus or from *in vitro* produced receptor which, in some cases (21), also contained an incorrect amino acid (valine) at residue position 400. Possibly, this *in vitro* produced protein did not fold correctly or was unstable. Our experiments show that ERs containing several different amino acids substituted for tyrosine 537, when made and tested in intact cells, are fully able to activate ERE-dependent transcription in the presence, and sometimes even in the absence, of E $_2$. In addition, suppression of the activity of these receptors by antiestrogens implies that the transcriptional activity is indeed receptor mediated. While we have demonstrated that the potentially phosphorylatable tyrosine 537 is not required for E $_2$ -induced transcriptional activity of the ER, it is possible that this tyrosine might be a target for other signaling pathways such that the character of this residue might affect the ability of some growth factors to regulate ER activity, possibly in a tissue-specific manner. Additional studies will be needed to address these aspects.

We conclude, therefore, that Y537 is in a region of the receptor that is critical for ligand regulation of transcriptional activities, such that small changes in receptor structure (by point mutation and possibly other modifications) can impact greatly on the biological activity of the receptor, especially in its unliganded state.

MATERIALS AND METHODS

Plasmids and General Reagents

The plasmids 2ERE-pS2-CAT (38), 2ERE-TK-CAT (39), 2ERE-TATA-CAT (40), pCMV5 hER (41), pCH110 (35), and pCMV β (35, 41) (Clontech, Palo Alto, CA) have been previously described. The plasmid encoding SRC-1 (36) was kindly provided by Drs. Ming Tsai and Bert O'Malley (Baylor College of Medicine, Houston, TX). The plasmid pGEX-2TK-ER, which contains the hER spanning amino acids 282–595 (42) was kindly provided by Dr. Myles Brown (Harvard Med-

ical School, Boston, MA). The vector pTZ19R was kindly provided by Dr. Byron Kemper (University of Illinois, Urbana, IL) and pBluescript II SK⁺ was from Stratagene (La Jolla, CA). Plasmids were purified for transfection using either CsCl gradient centrifugation or a plasmid preparation kit according to the manufacturer's instructions (Qiagen, Chatsworth, CA). Restriction enzymes were purchased from GIBCO BRL (Gaithersburg, MD) and New England Biolabs (Beverly, MA). Cell culture media, calf serum, and other reagents for cell culture were purchased from GIBCO BRL and Sigma Chemical Co. (St. Louis, MO). For Western analysis, nitrocellulose membrane was obtained from Millipore (Marlborough, MA), the H226 antibody was kindly provided by Dr. Geoffrey Greene (University of Chicago, Chicago, IL), and rabbit anti-rat IgG was purchased from Zymed (San Francisco, CA). Radioisotopes for CAT assays, sequencing, hormone-binding assays, and Western blotting were purchased from Dupont NEN (Boston, MA) and Amersham (Arlington Heights, IL). E₂ was from Sigma, and TOT and ICI 164,384 were kindly provided by Zeneca Pharmaceuticals (Macclesfield, U.K.).

Oligo-Directed Mutagenesis

The 1.8-kb ER-containing *Bam*HI fragment from pCMV5 hER was cloned into the *Bam*HI site of pBluescript II SK⁺. Site-directed mutagenesis was then performed according to Kunkel *et al.* (43) using the following oligo-nucleotides:

GTGGTGCCCTCGCAGATCTGCTGCTGGAG, Y537A;
GAACGTGGTACCCCTCTTCGACCTGCTGCTGG, Y537F;
GGTGCCCTCAAAGATCTGCTGCTGG, Y537K;
GGTGCCCTCGAGGACCTGCTGCTGG, Y537E; and
GGTGCCCTCTCAGATCTGCTGCTGG, Y537S. Oligonucleotides were purchased from GIBCO BRL. Screening for the desired ER mutations was done by restriction enzyme analysis via silent mutations that incorporated a *Bgl*II site into Y537A, Y537K, and Y537S; an *Xho*I site into Y537E; and a *Kpn*I site into Y537F. After mutagenesis, the ER cDNAs were excised from pBluescript II SK⁺ using *Bam*HI and ligated into the *Bam*HI site of the cytomegalovirus-driven expression vector, pCMV5, kindly provided by Dr. David Russell (University of Texas, Dallas, TX) (44). The GST-ER mutant plasmids for Y537A and Y537S were constructed by digesting the pGEX-2TK-ER hormone-binding domain wild type construct with *Eag*I/*Bsm*I to excise a fragment of the ER. We then inserted *Eag*I/*Bsm*I fragments, which contained the mutations for Y537A or Y537S into the digested pGEX-2TK construct. All ER mutations were then confirmed by dideoxy sequence analysis using a Sequenase 2.0 kit from Amersham.

Cell Culture and Transfections

Transfections were done in either ER-negative human breast cancer MDA-MB-231 cells or CHO cells. Cells were maintained and transfected as previously described (35, 41, 45). 231 cells were plated for transfection at a density of 3×10^6 cells/100 mm dish and incubated for 40–48 h at 37 C with 5% CO₂. Transfections were performed using 2.0 μ g of either 2ERE-pS2-CAT or 2ERE-TK-CAT, 0.8 μ g of the internal reference β -galactosidase reporter plasmid pCMV β , 0.1 μ g ER expression vector, and pTZ19R carrier plasmid to 15 μ g total DNA per 100-mm diameter dish of cells. Cells were incubated with calcium phosphate-precipitated DNA for 4 h and then subjected to a 2.5-min glycerol shock, using 20% glycerol in growth medium, followed by a 2.5-min rinse in HBSS. Ligand treatment was then added in growth medium. CHO cells were plated at 1×10^5 cells per 100-mm dish and transfected with 1.6 μ g 2ERE-TATA-CAT reporter plasmid, 0.3 μ g of the β -galactosidase reporter plasmid pCH110, 0.01 μ g ER expression vector, and pTZ19R carrier plasmid to 8 μ g total DNA per 60-mm diameter dish of cells. Cells were incubated with calcium phosphate-precipitated DNA for 14 h and then sub-

jected to a 1.5-min glycerol shock, using 20% glycerol in HBSS, followed by a 1.0-min rinse in HBSS. Ligand treatment was then added in growth medium. In each case, cells were harvested 24 h after ligand treatment and lysed by three cycles of freezing on dry ice and thawing at 37 C. Transactivation ability as determined by CAT activity of the whole-cell lysates was assayed as described previously (35, 41). CAT assays were normalized to β -galactosidase activity from the cotransfected internal control plasmid.

Western Analysis

231 cells were transfected in 100-mm dishes with 10 μ g ER expression vector and 5 μ g pTZ19R carrier plasmid DNA. After harvesting in cold HBSS, the cells were centrifuged at $200 \times g$ for 5 min and resuspended in 20 mM Tris (pH 7.4), 0.5 M NaCl, 1.0 mM dithiothreitol, 10% glycerol (vol/vol), 50 μ g/ml leupeptin, 50 μ g/ml aprotinin, 2.5 μ g/ml pepstatin, and 0.2 mM phenylmethylsulfonylfluoride. Whole cell extracts were obtained by subjecting cells to three rounds of freezing on dry ice and thawing on wet ice followed by centrifugation at $15,000 \times g$ to remove cell debris. Equal amounts of total protein were loaded on a 10% SDS polyacrylamide gel. Electrophoresis and Western blotting were done according to standard methods (35). Nitrocellulose blots were probed with the hER-specific primary antibody H226 at 2.0 μ g/ml, then incubated with rabbit anti-rat IgG (1.0 μ g/ml), and detected with ¹²⁵I-conjugated protein A.

Hormone-Binding Assays

Binding assays for E₂ and Scatchard analysis were performed as described previously (35). 231 cells were transfected and whole-cell extracts were prepared as for Western blot analysis. Cell extracts were then incubated with concentrations of [³H]E₂ from 3×10^{-11} M to 2×10^{-9} M in the presence or absence of a 100-fold excess of radioinert E₂ to determine nonspecific and total binding, respectively. Ligand was diluted in 10 mM Tris (pH 7.4), 1.5 mM EDTA so that the final ethanol concentration in the reactions did not exceed 1.0% (vol/vol). Whole-cell extracts and ligand were incubated together at 4 C overnight, and unbound E₂ was removed from the samples by treatment with dextran-treated charcoal for 15 min at 4 C. Approximately 1.0 μ g of total protein was assayed at each concentration of hormone. Equilibrium dissociation constants (K_d) for the wild type and mutant ERs were determined by Scatchard analysis (46).

In Vitro Translation of SRC-1 and Assays of Interaction With ERs

In vitro translation of SRC-1 (36) was performed using the Promega TNT kit (Madison, WI). Briefly, 1 μ g SRC-1 vector was mixed with 25 μ l TNT rabbit reticulocyte lysate, 2 μ l TNT buffer, 1 μ l T3 RNA polymerase (20 U/ μ l), and 4 μ l [³⁵S] methionine (15 μ Ci/ μ l) (ICN, Costa Mesa, CA). The final reaction of 50 μ l was incubated for 90 min at 30 C. The translation efficiency was checked by analyzing 1 μ l of lysate by SDS-PAGE. Glutathione Sepharose (Pharmacia, Piscataway, NJ) was equilibrated with binding buffer (25 mM Tris-HCl (pH 7.9), 10% (vol/vol) glycerol, 0.1% NP-40, 0.5 mM dithiothreitol, 100 mM KCl). Five hundred micrograms of *Escherichia coli* bacterial crude extract containing GST ER hormone-binding domain (amino acids 282–595) fusion proteins were incubated at 4 C with 25 μ l of Sepharose beads for 2.5 h in the presence of control (0.1% ethanol) vehicle or hormone (E₂ or TOT at 1 μ M concentration). After three washes, the beads were incubated with 5 μ l of *in vitro* translated SRC-1 for 2.5 h in the presence of control vehicle or hormone at 4 C. The beads were washed three times with 1 ml binding buffer and two times with 1 ml of binding buffer containing 300 mM KCl. After

washing, the beads were boiled in SDS sample buffer, and one-fourth of each protein sample was analyzed by SDS-PAGE. The gel was dried and detected by autoradiography.

Acknowledgments

We thank Ming Tsai, Bert O'Malley, and Myles Brown for providing plasmids, Geoffrey Greene and Abbott Laboratories for providing ER antibody, and Alan Wakeling and Zeneca Pharmaceuticals for providing antiestrogens.

Received July 1, 1996. Revision received and accepted August 16, 1996.

Address requests for reprints to: Dr. Benita Katzenellenbogen, Department of Molecular and Integrative Physiology, University of Illinois, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, Illinois 61801-3704.

This work was supported by NIH Grant CA-18119 (to B.S.K.) and in part by NIH Postdoctoral Fellowship 1 F32 CA-68653 (to K.E.).

REFERENCES

- Evans RM 1988 The steroid and thyroid hormone receptor superfamily. *Science* 240:889-895
- Gronemeyer H 1991 Transcription activation by estrogen and progesterone receptors. *Annu Rev Genet* 25:89-123
- Truss M, Beato M 1993 Steroid hormone receptors: interaction with deoxyribonucleic acid and transcription factors. *Endocr Rev* 14:459-479
- Tsai M-J, O'Malley BW 1994 Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 63:451-486
- Beato M, Herrlich P, Schütz G 1995 Steroid hormone receptors: many actors in search of a plot. *Cell* 83:851-857
- Katzenellenbogen JA, O'Malley BW, Katzenellenbogen BS 1996 Tripartite steroid hormone receptor pharmacology: interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. *Mol Endocrinol* 10:119-131
- Katzenellenbogen BS 1993 Estrogen receptors: bioactivities and interactions with cell signaling pathways. *Biol Reprod* 54:287-293
- Kumar V, Green S, Stack G, Berry M, Jin JR, Chambon P 1987 Functional domains of the human oestrogen receptor. *Cell* 51:941-951
- Fawell SE, Lees JA, White R, Parker MG 1990 Characterization and colocalization of steroid binding and dimerization activities in the mouse estrogen receptor. *Cell* 60:953-962
- Kumar V, Green S, Staub A, Chambon P 1986 Localisation of the estradiol-binding and putative DNA-binding domains of the human oestrogen receptor. *EMBO J* 5:2231-2236
- Tora L, White J, Brou C, Tasset D, Webster N, Scheer E, Chambon P 1989 The human estrogen receptor has two nonacidic transcriptional activation functions. *Cell* 59:477-487
- Webster NJ, Green S, Jin JR, Chambon P 1988 The hormone-binding domains of the estrogen and glucocorticoid receptors contain an inducible transcription activation function. *Cell* 54:199-207
- Katzenellenbogen BS, Montano M, Le Goff P, Schodin DJ, Kraus WL, Bhardwaj B, Fujimoto N 1995 Antiestrogens: mechanisms and actions in target cells. *J Steroid Biochem Mol Biol* 53:387-393
- Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS 1994 Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examinations of their influence on transcriptional activity. *J Biol Chem* 269:4458-4466
- Ali S, Metzger D, Bornert J-M, Chambon P 1993 Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J* 12:1153-1160
- Denton RR, Koszewski NJ, Notides AC 1992 Estrogen receptor phosphorylation: hormonal dependence and consequence on specific DNA binding. *J Biol Chem* 267:7263-7268
- Lahooti H, White R, Danielian PS, Parker MG 1994 Characterization of ligand-dependent phosphorylation of the estrogen receptor. *Mol Endocrinol* 8:182-188
- Arnold SF, Obourn JD, Jaffe H, Notides AC 1994 Serine 167 is the major estradiol-induced phosphorylation site on the human estrogen receptor. *Mol Endocrinol* 8:1208-1214
- Castoria G, Migliaccio A, Green S, Di Domenico M, Chambon P, Auricchio F 1993 Properties of a purified estradiol-dependent calf uterus tyrosine kinase. *Biochemistry* 32:1740-1750
- Auricchio F, Migliaccio A, Di Domenico M, Nola E 1987 Oestradiol stimulates tyrosine phosphorylation and hormone binding activity of its own receptor in a cell-free system. *EMBO J* 6:2923-2929
- Migliaccio A, Di Domenico M, Green S, de Falco A, Kajtaniak EL, Blasi F, Chambon P, Auricchio F 1989 Phosphorylation on tyrosine of *in vitro* synthesized human estrogen receptor activates its hormone binding. *Mol Endocrinol* 3:1061-1069
- Arnold SF, Obourn JD, Jaffe H, Notides AC 1995 Phosphorylation of the human estrogen receptor on tyrosine 537 *in vivo* and by src-family tyrosine kinase *in vitro*. *Mol Endocrinol* 9:24-33
- Arnold SF, Vorobjikina DP, Notides AC 1995 Phosphorylation of tyrosine 537 on the human estrogen receptor is required for binding to an estrogen response element. *J Biol Chem* 270:30205-30212
- Ekena K, Weis KE, Katzenellenbogen JA, Katzenellenbogen BS 1996 Identification of amino acids in the hormone binding domain of the human estrogen receptor important in estrogen binding. *J Biol Chem* 271:20053-20059
- Aronica SM, Katzenellenbogen BS 1991 Progesterone receptor regulation in uterine cells: stimulation by estrogen, cyclic adenosine 3',5'-monophosphate and insulin-like growth factor I and suppression by antiestrogens and protein kinase inhibitors. *Endocrinology* 128:2045-2052
- Aronica SM, Katzenellenbogen BS 1993 Stimulation of estrogen receptor-mediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic adenosine monophosphate, and insulin-like growth factor-I. *Mol Endocrinol* 7:743-752
- Cho H, Katzenellenbogen BS 1993 Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Mol Endocrinol* 7:441-452
- Cho H, Aronica SM, Katzenellenbogen BS 1994 Regulation of progesterone receptor gene expression in MCF-7 breast cancer cells: a comparison of the effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology* 134:658-664
- Sumida C, Lecerf F, Pasqualini JR 1988 Control of progesterone receptors in fetal uterine cells in culture: effects of estradiol, progestins, antiestrogens, and growth factors. *Endocrinology* 122:3-11
- Ignar-Trowbridge DM, Nelson KG, Bidwell MC, Curtis SW, Washburn TF, McLachlan JA, Korach KS 1992 Coupling of dual signaling pathways: epidermal growth factor action involves the estrogen receptor. *Proc Natl Acad Sci*

- USA 89:4658-4662
31. Bourguet W, Ruff M, Chambon P, Gronemeyer H, Moras D 1995 Crystal structure of the ligand-binding domain of the human nuclear receptor RXR- α . *Nature* 375:377-382
 32. Renaud JP, Rochel N, Ruff M, Vivat V, Chambon P, Gronemeyer H, Moras D 1995 Crystal structure of the RAR- γ ligand-binding domain bound to all-trans retinoic acid. *Nature* 378:681-689
 33. Wagner RL, Apriletti JW, McGrath ME, West BL, Baxter JD, Fletterick RL 1995 A structural role for hormone in the thyroid hormone receptor. *Nature* 378:690-697
 34. Wurtz JM, Bourguet W, Renaud JP, Vivat V, Chambon P, Moras D, Gronemeyer H 1996 A canonical structure for the ligand binding domain of nuclear receptors. *Nature Struct Biol* 3:87-94
 35. Pakdel F, Katzenellenbogen BS 1992 Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J Biol Chem* 267:3429-3437
 36. Oñate SA, Tsai SY, Tsai M-J, O'Malley BW 1995 Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270:1354-1357
 37. Pakdel F, Reese JC, Katzenellenbogen BS 1993 Identification of charged residues in an N-terminal portion of the hormone binding domain of the human estrogen receptor important in transcriptional activity of the receptor. *Mol Endocrinol* 7:1408-1417
 38. Montano MM, Ekena K, Krueger KD, Keller AL, Katzenellenbogen BS 1996 Human estrogen receptor ligand activity inversion mutants: receptors that interpret antiestrogens as estrogens and estrogens as antiestrogens and discriminate among different antiestrogens. *Mol Endocrinol* 10:230-242
 39. Kraus WL, Montano MM, Katzenellenbogen BS 1994 Identification of multiple, widely spaced estrogen-responsive regions in the rat progesterone receptor gene. *Mol Endocrinol* 8:952-969
 40. Ince BA, Zhuang Y, Wrenn CK, Shapiro DJ, Katzenellenbogen BS 1993 Powerful dominant negative mutants of the human estrogen receptor. *J Biol Chem* 268:14026-14032
 41. Reese JC, Katzenellenbogen BS 1991 Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor overexpressed in mammalian cells. *Nucleic Acids Res* 19:6595-6602
 42. Halachmi S, Marden E, Martin G, MacKay H, Abbondanza C, Brown M 1994 Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription. *Science* 264:1455-1458
 43. Kunkel TA, Roberts JD, Zakour RA 1987 Rapid and efficient site-specific mutagenesis without phenotypic selection. *Methods Enzymol.* 154:367-382
 44. Andersson S, Davis DL, Dahlback H, Jornvall H, Russel DW 1989 Cloning, structure, and expression of the mitochondrial cytochrome P-450 sterol 26-hydroxylase, a bile acid biosynthetic enzyme. *J Biol Chem* 264:8222-8229
 45. Montano MM, Müller V, Trobaugh A, Katzenellenbogen BS 1995 The carboxy-terminal F domain of the human estrogen receptor: role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 9:814-825
 46. Scatchard C 1949 The attractions of protein for small molecules and ions. *Ann NY Acad Sci* 51:660-672



Identification of a Novel Transferable *cis* Element in the Promoter of an Estrogen-Responsive Gene that Modulates Sensitivity to Hormone and Antihormone

Monica M. Montano, W. Lee Kraus*, and
Benita S. Katzenellenbogen

Department of Molecular and Integrative Physiology
(M.M.M., W.L.K., B.S.K.)
Department of Cell and Structural Biology (B.S.K.)
University of Illinois,
Urbana, Illinois 61801

The estrogen receptor (ER) is a ligand-regulated transcription factor that acts at the promoters of estrogen-regulated genes to modulate their expression. In the present study, we examined three estrogen-regulated promoters, namely the rat progesterone receptor gene distal (PR_D) and proximal (PR_P) promoters and the human pS2 gene promoter, and observed marked differences in their sensitivity to stimulation by estrogen and repression of estrogen-stimulated transcription by antiestrogen (AE)-occupied ER. ER-containing MCF-7 human breast cancer cells were transfected with reporter gene constructs containing estrogen response elements upstream of the three gene promoters. In this system, PR_P and PR_D showed similar dose-response curves for stimulation by estradiol whereas pS2 was activated by even lower concentrations of estradiol. By contrast, PR_D was much less sensitive to repression of estrogen-stimulated activity by all AEs studied, relative to the PR_P and the pS2 promoters. Using deletion and mutational analysis, we have identified a transferable *cis* element at -131 to -94 bp in PR_D that is involved in modulating the sensitivity of this promoter to both estrogens and AEs. The element reduced the magnitude of estrogen-stimulated activity, enhanced the ability of AEs to repress estrogen-stimulated activity, and elicited similar effects when transferred to the promoter of another estrogen-responsive gene. Thus, removal of this region from PR_D further accentuated the insensitivity of this promoter to AE while enhancing its sensitivity (both EC₅₀ and fold induction) to es-

trogen. Gel mobility shift assays showed that proteins from nuclear extracts of MCF-7 cells interact with this element and that the binding of these proteins is inversely correlated with the transcriptional effectiveness of the ER. The findings demonstrate that a specific *cis* element from the promoter of an estrogen-responsive gene can alter the transcriptional activity of hormone and antihormone-occupied receptor bound at its response element near the promoter. Such ligand response modulatory elements, and changes in the levels and activity of factors that bind to such elements, may underlie the different sensitivities of steroid hormone-regulated genes to both hormones and antihormones. (Molecular Endocrinology 11: 330-341, 1997)

INTRODUCTION

Steroid hormones, such as estrogen, modulate gene expression via intracellular receptors that belong to a large superfamily of hormone-regulated transcription factors. In the case of the estrogen receptor (ER), the binding of estrogen initiates a process of receptor activation that includes the high-affinity binding of ER to specific DNA sequences, termed estrogen response elements (EREs). The interaction of ER with EREs results in the modulation of specific gene expression, through which the physiological actions of estrogens are manifested (for reviews, see Refs. 1-5). The regulatory actions of estrogens on gene expression, which are generally stimulatory, can be inhibited by potent synthetic ER antagonists (6) termed antiestrogens (AEs).

The promoters of many known estrogen-regulated genes are complex, with binding sites for other transcription factors in addition to ER. Positive and negative interactions between ER and these transcription factors, which may be promoter- or cell-specific, provide an important step at which ER function may be regulated (reviewed in Refs. 1, 4, and 5). A number of studies from this laboratory and others have demonstrated the significance of promoter and cell context in modulating responses to both estrogens and AEs (7–9).

In the present study, we observed marked differences in the sensitivities of three estrogen-regulated promoters to repression by AEs, suggesting the involvement of promoter-specific factors capable of modulating the activity of the ER. Using several approaches, we have identified a transferable *cis* element in the rat progesterone receptor (PR) gene distal promoter that is involved in modulating promoter sensitivity to both estrogens and AEs. Gel mobility shift assays have been used to show that proteins from nuclear extracts of MCF-7 human breast cancer cells interact with this ligand response modulatory element (LRME) and that the binding of these proteins is inversely correlated with the transcriptional effectiveness of ER. Our results demonstrate that *cis*-acting elements in the promoter region of estrogen-responsive genes can alter the transcriptional activity of estrogen- and AE-occupied ER bound at its response element near a promoter. Such ligand response modulatory elements may be broadly applicable in the actions of many nuclear receptors in which gene-specific modulation of hormonal induction is known, but the underlying basis is poorly understood.

RESULTS

Examination of the Differential Sensitivity of Several Estrogen-Stimulated Promoters to Repression by AEs

As shown in Fig. 1, we analyzed three estrogen-regulated promoters [the PR gene distal and proximal promoters (PR_D and PR_P) and the promoter of the human pS2 gene; pS2] for their relative sensitivity to the stimulatory actions of estrogen and the repressive actions of AEs. We previously cloned the 5'-flanking region of the rat PR gene and demonstrated the presence of two promoters, a distal promoter (–131 to +65; PR_D) and a proximal promoter (+461 to +675; PR_P) (10), and we have shown that these promoters are functionally distinct with respect to activation by ER-dependent pathways (11). The PR distal and proximal promoters control production of the B and A isoforms of the PR (ca. 120- and 90-kDa, respectively), and the pS2 promoter regulates production of a growth factor-like secreted protein whose function is not completely known (12).

For the studies in Fig. 1, MCF-7 human breast cancer cells, which contain high levels of endogenous ER,

were transfected with chloramphenicol acetyltransferase (CAT) reporter constructs containing two consensus EREs upstream of the pS2, PR_P or PR_D promoters. Extracts from the cells were analyzed for CAT activity after treatment with the lowest maximally stimulatory concentration of estradiol (E₂; 10^{–9} M) in the absence or presence of a 500- or 1000-fold excess of the AEs, ICI 164,384 (ICI) or LY 117018 (LY), respectively. As shown in Fig. 1, E₂ stimulated large (i.e. 50- to 130-fold) increases in the activity of the three promoter-reporter gene constructs. The AEs (ICI and LY) alone evoked essentially no activity, and they were able to repress greater than 90% of the E₂-stimulated CAT activity from either the pS2 or PR_P promoter-containing reporters. Of note, the AEs were substantially less effective at repressing E₂-stimulated activity from the PR_D-containing reporter (Fig. 1; and further investigated in Fig. 3 below).

Characterization of a Region of the PR_D Promoter that Modulates Sensitivity to Estrogen and AE: Deletion and Mutational Analyses

We further analyzed PR_D to identify region(s) involved in modulating the sensitivity of the promoter to the suppressive effects of AE on E₂-stimulated activity. Reporter constructs containing two consensus EREs upstream of the full-length PR_D, or deletion mutants of PR_D, were analyzed for stimulation by E₂ and repression by ICI. The results are shown in Fig. 2A. Deletion of the –131 to –94 *XmnI*/*BsmI* fragment from PR_D (to generate a truncated promoter denoted PR_{D, B/N}), since this truncated promoter contains the region –94 to +65, which spans from the *BsmI* to the *NheI* restriction sites) resulted in approximately 2-fold higher induction by E₂ than was observed with the full-length promoter (Fig. 2A; compare line 2 with line 1). Surprisingly, we also observed a 2-fold decrease in the sensitivity of the deleted promoter to the inhibitory actions of ICI relative to the full-length promoter (Fig. 2A). This indicates, as shown in Fig. 3 also (see below), that the –131 to –94 region plays a role in modulating the sensitivity of PR_D to both estrogens and AEs and that in its absence (as in PR_{D, B/N}), PR_D gene responsiveness to estrogen and AE is even more discordant than that of PR_P and pS2.

Further deletion of the PR_D promoter to –67 resulted in a substantial loss of estrogen-inducible activity (line 3). Deletion from the 3'-end of PR_D (the +25 to +65 region) similarly reduced promoter activity (line 4 vs. line 1), and further deletion to +1 almost fully destroyed PR_D activity (line 5), as expected. We also tested PR_D in the context of the natural estrogen-responsive sequences of the PR gene (line 6). With the ERE-like sequences contained in the five estrogen-responsive fragments of the rat PR gene linked together and placed upstream of PR_D [5E-PR_D-CAT (11)], we observed good stimulation by E₂ [50% of the magnitude observed with (ERE)₂-PR_D-CAT, line 1] and the same poor sensitivity to repression by ICI (Fig. 2A,

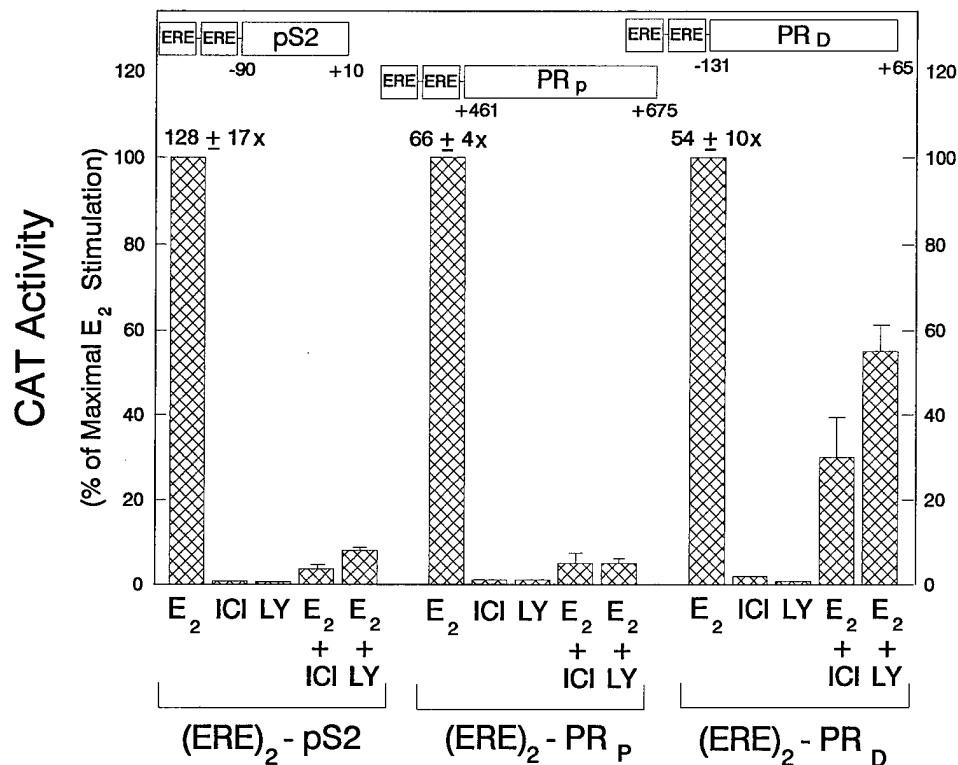


Fig. 1. Differential Sensitivity of Three Promoters to the Inhibitory Actions of AEs

MCF-7 cells were transfected with CAT reporter constructs containing two consensus EREs upstream of the pS2 promoter [−90 to +10; (ERE)₂-pS2-CAT], the proximal promoter of the rat PR gene [−131 to +65; (ERE)₂-PR_p-CAT], or the distal promoter of the rat PR gene [+461 to +675; (ERE)₂-PR_D-CAT], and a β-galactosidase expression plasmid, used as an internal control to correct for transfection efficiency, as described in *Materials and Methods*. The cells were then treated for 24 h with the estrogen E₂ (10^{−9} M), or the AE ICI 164,384 (ICI, 5 × 10^{−7} M) or LY 117018 (LY, 10^{−6} M) alone or in the combinations as indicated. Cell extracts were prepared and analyzed for CAT activity as described in *Materials and Methods*. The activity for each construct was expressed as a percent of the activity observed with E₂ treatment alone, which is set at 100%. Each bar represents the mean of three or more separate determinations + SEM. The numbers above the E₂ bar show the fold induction observed with E₂ alone for each of the three promoter constructs.

line 6, only 60% repression) as seen with 2ERE-PR_D-CAT (line 1). This indicates that the unusual resistance of PR_D to AE inhibition is a function of the 5′-flanking region to −131 and not the nature of the estrogen-response element regions.

Mutagenesis of the −131 to −84 region of PR_D (Fig. 2B) identified a nucleotide sequence that appears to be involved in conferring differential sensitivity to stimulation by estrogen and repression by AEs. Introduction of mutations at −115 to −110 of the PR gene distal promoter (Mut3) increased the magnitude of the response to E₂ (2-fold) and decreased the ability of ICI to suppress E₂-mediated transactivation with respect to the wild type promoter construct, reproducing what was observed upon deletion of the −131 to −94 region of PR_D (Fig. 2A, line 2). Mutations in the nucleotide sequence corresponding to the putative CTF/NF-1 site (Mut 4 and Mut 5) decreased E₂-mediated transactivation from PR_D to 40–50% of the wild type promoter, suggesting an involvement of CTF/NF-1 nuclear factors in estrogen regulation of PR_D, possibly similar to that observed previously for the vitellogenin

B1 gene (13). Mutations at other sites within the −131 to −84 region (Mut 1, 2, and 6) had relatively little effect on the response to E₂ or ICI.

Analysis of the Relationship between Estrogen Stimulation of the Three Promoters and the Sensitivity of the Three Promoters to AE Repression

We performed E₂ and AE (ICI) dose-response experiments using the estrogen-responsive reporter constructs containing the three different promoters (PR_D, PR_p, and pS2). We also assessed whether the decreased sensitivity of the −131 to −94 deleted PR_D (i.e. the PR_{D,B/N} promoter) to AE was attributable to the greater sensitivity of PR_{D, B/N} to stimulation by E₂ relative to PR_D.

The PR_p and PR_D gene promoter constructs showed similar dose-response curves for stimulation by E₂ (Fig. 3A), with half-maximal stimulation at approximately 3 × 10^{−11} M E₂. The pS2 promoter-containing construct [(ERE)₂-pS2-CAT] showed a 2- to

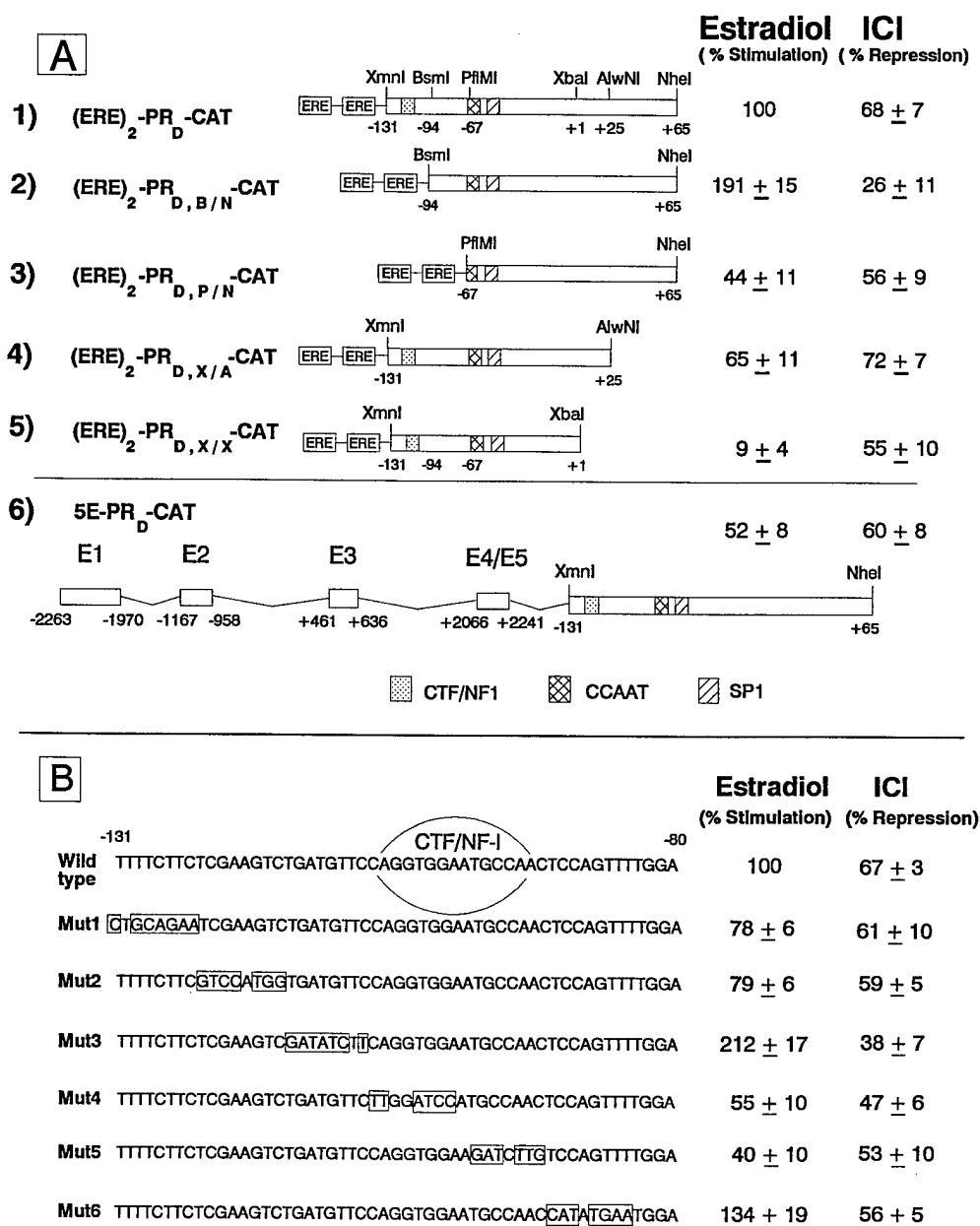


Fig. 2. Identification of a Region of PR_D Involved in ER-Ligand Sensitivity

Panel A, Estrogen stimulation and AE (ICI) repression were examined using (ERE)₂-PR_D-CAT or CAT reporter constructs containing two EREs upstream of PR_D deletion mutants (lines 1–5), or CAT reporter constructs containing the five natural ERE-containing estrogen-responsive regions of the PR gene linked together and placed upstream of PR_D (line 6), which were transfected into MCF-7 cells as described in the legend of Fig. 1. Differential responsiveness of the reporter constructs to E₂ (10⁻⁹ M) and to repression of E₂-stimulated activity by ICI (5 × 10⁻⁷ M) was monitored. The magnitude of transactivation of the full-length PR_D in response to E₂ was set at 100%. Percent repression with ICI indicates the percent inhibition of E₂-stimulated activity observed for each reporter construct upon cotreatment with 10⁻⁹ M E₂ and 5 × 10⁻⁷ M ICI. Each value represents the mean of three or more separate determinations ± SEM. Panel B, Mutagenesis of the region from -131 to -80 encompassing the XmnI/BsmI fragment of PR_D results in differential responsiveness to E₂ (10⁻⁹ M) and repression of E₂-stimulated activity by ICI (5 × 10⁻⁷ M). Shown is the nucleotide sequence of the -131 to -80 region of the PR_D promoter. The mutated nucleotides are indicated by the boxed regions in mutants 1 to 6. The CCAAT motif, Sp1-binding site, and putative binding site for CTF/NFI are also indicated. CAT reporter constructs containing two EREs upstream of the mutated PR_D were examined for E₂ responsiveness and for suppression of E₂-mediated transactivation by ICI as described in the legend of Fig. 1. The magnitude of transactivation of wild type PR_D in response to E₂ was set at 100%. Percent repression with ICI indicates the percent inhibition of E₂-stimulated activity observed for each reporter construct upon cotreatment with 10⁻⁹ M E₂ and 5 × 10⁻⁷ M ICI. Each value represents the mean of three or more separate determinations ± SEM.

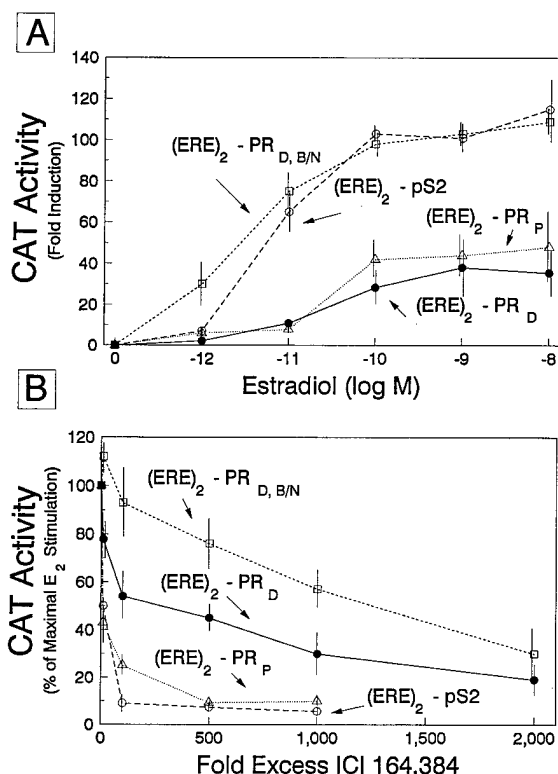


Fig. 3. Dose-Dependent Stimulatory Effects of E₂ (Panel A) and Dose-Dependent Inhibitory Effects of ICI on E₂ (10⁻⁹ M)-Stimulated CAT Activity (Panel B) Using the Three Estrogen-Responsive Promoter-Reporter Constructs

Constructs are defined in Figs. 1 and 2. The truncated PR_D is denoted PR_{D, B/N} and lacks the -131 to -94 *XmnI/BsmI* fragment of the PR_D promoter. Each value represents the mean of three or more separate determinations \pm SEM.

3-fold greater maximal CAT activity, and half-maximal activity required 5- to 10-fold less E₂ ($\sim 5 \times 10^{-12}$ M E₂, Fig. 3A). The pS2 and PR_P promoter constructs were equally and highly sensitive to suppression of E₂-stimulated CAT activity by the AE ICI, whereas the PR_D promoter was much less sensitive to ICI suppression over the entire concentration range tested (Fig. 3B), consistent with the data shown in Fig. 1, in which only a single concentration of ICI was used to inhibit E₂ activation. Thus, promoters that have similar sensitivities to the stimulatory actions of estrogens can have very different sensitivities to the inhibitory actions of AEs.

Shown in Fig. 3, panels A and B, is our observation that deletion of the -131 to -94 portion of the PR_D promoter [to give (ERE)₂-PR_{D, B/N}-CAT] resulted in a 3-fold increase in the magnitude of CAT activity in response to E₂ (Fig. 3A) and also resulted in a reduced sensitivity to suppression by ICI relative to that shown by the intact PR_D promoter construct [(ERE)₂-PR_D-CAT] (Fig. 3B). In addition, the truncated promoter construct [(ERE)₂-PR_{D, B/N}-CAT] required 10-fold less E₂ (i.e. $\sim 3 \times 10^{-12}$ M E₂) for half-maximal activity

compared with the PR_D-containing reporter [(ERE)₂-PR_D-CAT] (Fig. 3A).

It is of note that although the PR_{D, B/N} and the pS2 promoter constructs required similar E₂ concentrations for half-maximal activity, they had very different dose-response curves for suppression by ICI (Fig. 3B). These results illustrate the lack of correlation between the estrogen and AE sensitivities of a particular promoter and suggest that the decreased sensitivity of PR_{D, B/N} to repression by ICI relative to PR_D was not related to its increased sensitivity to the stimulatory actions of E₂.

Our initial expectation in deletion and mutagenesis studies in the PR_D promoter was that we would identify a region conferring the resistance that this promoter shows to AE antagonism. We failed to find such a region as far as we were able to study through deletions and mutations. Although this aspect merits further study, we have found that further deletions (Fig. 2, lines 3-5) reduced estrogen responsiveness of the promoter, complicating such an approach. On the other hand, in the PR_D gene, we have made the unusual observation that a small region in PR_D has a strong modulatory effect on estrogen and AE responsiveness, and its presence confers higher estrogen sensitivity and higher AE repression, even though overall the PR_D is less estrogen and AE responsive than other genes such as pS2 and PR_P. Therefore, we investigated this modulatory element further.

Evaluation of the -131 to -94 Region of the PR_D Promoter as a Transferable *cis* Element

To determine whether the -131 to -94 *XmnI/BsmI* fragment of PR_D had the properties of a *cis* element, one or two copies of the fragment were cloned 50 bp upstream of the EREs in both the (ERE)₂-PR_D-CAT and (ERE)₂-PR_{D, B/N}-CAT reporter constructs (Fig. 4). In the context of the full-length PR_D, which contains the -131 to -94 region in its natural location, the presence of additional *XmnI/BsmI* (i.e. -131 to -94) fragments resulted in no substantial change in magnitude of E₂-stimulated activity or ICI repression (Fig. 4, lines 1-3). Deletion of the -131 to -94 fragment from PR_D caused a doubling of the level of induction by E₂ and a reduction in the magnitude of repression by ICI (Fig. 4, line 4), as noted earlier in Figs. 2 and 3. The addition of one *XmnI/BsmI* fragment 50 bp upstream of the two EREs in (ERE)₂-PR_{D, B/N}-CAT reduced E₂-inducibility by 50% with no change in AE repression (Fig. 4, line 5 vs. line 4). Two *XmnI/BsmI* fragments had the same effect on the level of E₂ induction and gave a greater (67%) repression by the AE ICI (Fig. 4, line 7, as seen with the intact PR_D (Fig. 4, line 1). The effect was not seen with the mutated (Mut 3) form of the *XmnI/BsmI* fragment (Fig. 4, line 6). Therefore, the effect of the -131 to -94 fragment was to reduce E₂ stimulation and increase ICI repression.

The activity of the -131 to -94 fragment was also observed in the context of the promoter of another

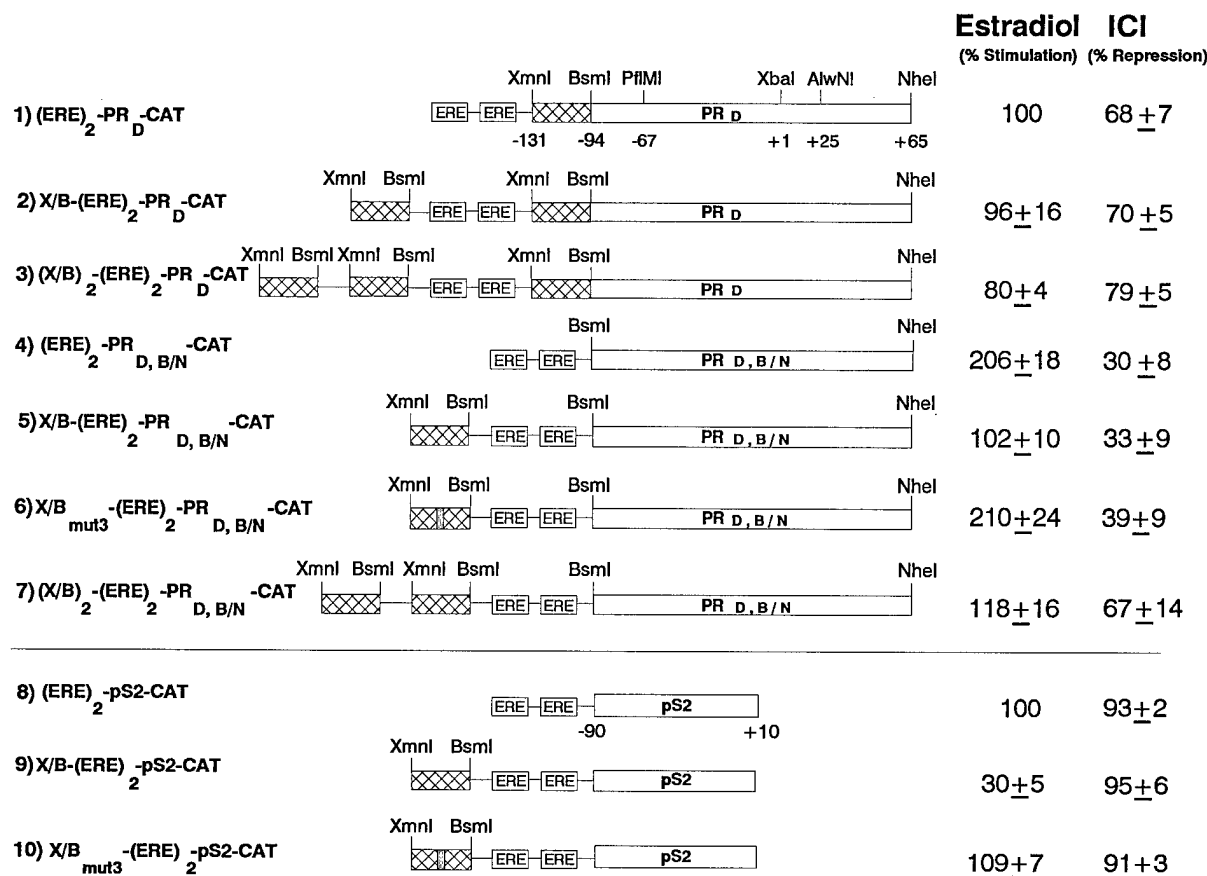


Fig. 4. The -131 to -94 Region of PR_D Acts as a Transferable *cis* Element

CAT reporter constructs containing two EREs upstream of either PR_D (-131 to +65 bp) or the truncated PR_D (-94 to +65 bp) denoted PR_{D, B/N} [lines 1-7] or pS2 (-90 to +10 bp) [lines 8-10] with one or two copies of the -131 to -94 XmnI/BsmI fragment of PR_D cloned 50 bp upstream of the EREs, were examined for E₂ responsiveness and for repression of E₂-stimulated CAT activity by ICI in MCF-7 cells as described in the legend of Fig. 1. For line 6 and line 10, one copy of the -131 to -94 XmnI/BsmI fragment containing the mutated nucleotides in mut3 (see Fig. 2B) was cloned upstream of ERE-containing PR_{D, B/N} or pS2, respectively. The magnitude of transactivation of wild type PR_D or wild type pS2 (without an XmnI/BsmI fragment cloned upstream of the ERE) was set at 100%. The percent repression with ICI indicates the percent inhibition of E₂-stimulated activity observed for each construct upon cotreatment with 10⁻⁹ M E₂ and 5 × 10⁻⁷ M ICI. Each value represents the mean of three or more separate determinations ± SEM.

estrogen-responsive gene. When the fragment was cloned upstream of the EREs in the (ERE)₂-pS2-CAT reporter construct, E₂-stimulated CAT activity was reduced to approximately 30% when compared with (ERE)₂-pS2-CAT lacking the fragment (Fig. 4, lines 8-9). ICI suppression, which in (ERE)₂-pS2 itself was greater than 90%, remained strong and unaffected, as expected (since ICI repression was complete and could not be increased further). This effect on E₂ stimulation was specific for the intact -131 to -94 XmnI/BsmI fragment and was not observed when the XmnI/BsmI fragment contained the Mut3 mutations (Fig. 4, line 10). Thus, the -131 to -94 XmnI/BsmI fragment satisfies two criteria of a regulatory *cis* element in that it is positionally independent and is transferable, being active in the context of a heterologous promoter, in this case, the promoter of another estrogen-responsive gene.

Analysis of the Effects of the -131 to -94 Region of the PR_D Promoter on Another Mediator of ER Action

The preceding studies examined the effects of the -131 to -94 region of PR_D on the actions of a typical positive regulator of ER function, namely E₂, and a typical fully negative regulator of ER function, namely the AE ICI. Since the ligand *trans*-hydroxytamoxifen (TOT) is a partial agonist/antagonist that can exhibit agonistic activity in certain promoter contexts (5-9), we wanted to determine whether the -131 to -94 region of PR_D could also modulate the agonistic effect of TOT.

As shown in Fig. 5, we examined the ability of the -131 to -94 region of PR_D to reduce the agonistic actions of TOT on different promoters, similar to the way it reduced the agonistic actions of E₂ as described

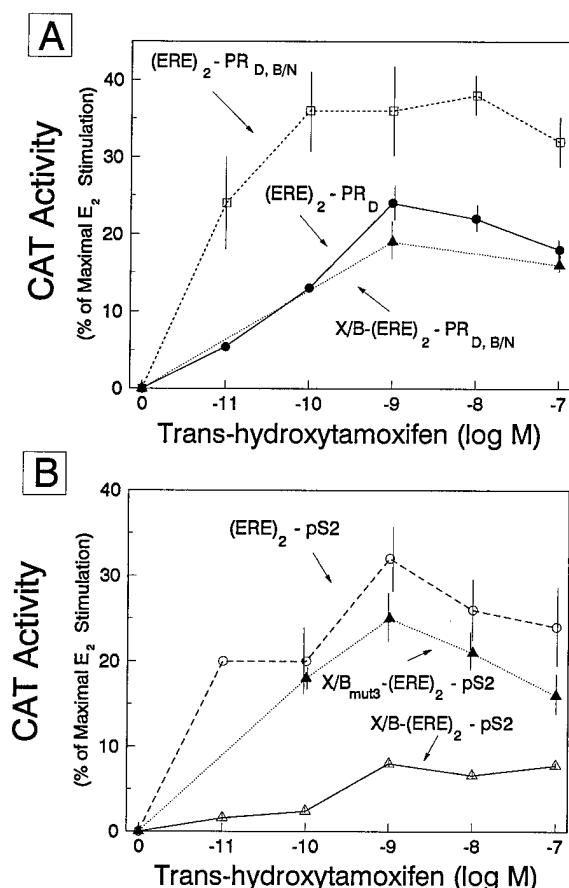


Fig. 5. The Effects of the -131 to -94 Region of PR_D on the Estrogen Agonist Activity of TOT

The (ERE)₂-PR_D-CAT, (ERE)₂-PR_{D, B/N}-CAT, (ERE)₂-PR_P-CAT, or (ERE)₂-pS2-CAT reporters were examined for responsiveness to the stimulatory (agonistic) actions of TOT (panels A and B) using MCF-7 cells as described in *Materials and Methods*. The activity for each construct is expressed as a percent of the maximal stimulation observed with 10⁻⁹ M E₂. Each value represents the mean of three or more separate determinations ± SEM.

above. Reporter constructs lacking the -131 to -94 *XmnI/BsmI* fragment from PR_D [i.e. (ERE)₂-PR_{D, B/N}-CAT and (ERE)₂-pS2-CAT] showed dose-dependent increases in CAT activity in response to treatment with TOT (Fig. 5, panels A and B) that were greater in magnitude than that of constructs containing the -131 to -94 *XmnI/BsmI* fragment [(ERE)₂-PR_D-CAT, Fig. 5A]. In addition, deletion of the -131 to -94 fragment from PR_D resulted in a promoter construct in which the EC₅₀ was lowered about 10-fold, from 10⁻¹⁰ M for (ERE)₂-PR_D-CAT to approximately 10⁻¹¹ M for (ERE)₂-PR_{D, B/N}-CAT. Also, addition of the *XmnI/BsmI* fragment to PR_{D, B/N} (to give X/B-(ERE)₂-PR_{D, B/N}) reduced TOT stimulation back to that of the intact PR_D construct (Fig. 5A). The same effect was seen with a different promoter (the pS2 gene promoter, Fig. 5B), in which addition of the *XmnI/BsmI* fragment [i.e. X/B-(ERE)₂-pS2-CAT] resulted in a great reduction in the

response to TOT, whereas addition of a mutated *XmnI/BsmI* fragment [X/B_{mut3}-(ERE)₂-pS2-CAT] resulted in virtually no change in the response to TOT (Fig. 5B). Thus, the -131 to -94 region of PR_D reduced the ability of ER to respond positively to at least two different types of stimulatory signals, namely E₂ and TOT acting as an estrogen agonist.

Interaction of MCF-7 Cell Factors with the -131 to -94 Region of PR_D

The identification of a region of PR_D (i.e. the -131 to -94 region) that can alter the sensitivity of the promoter to the stimulatory actions of estrogens and the inhibitory actions of AEs suggested the presence of a specific *trans*-acting factor(s) that could interact with this region. One major band (indicated in Fig. 6B) was detected in gel mobility shift assays using extracts from MCF-7 cells and a radiolabeled double-stranded oligomer containing the -131 to -94 sequence (shown in Fig. 6A). The band was competed by a 50-fold or 25-fold excess amount of unlabeled oligomer (Fig. 6B, lanes 7 and 11 vs. no competitor, lanes 6 and 10), but not by an excess of unlabeled -131/-94 oligonucleotide with a 6-bp mutation from -115 to -110 (mut3) (Fig. 6B, lane 8), indicating that the protein-DNA interaction producing the band was specific. Since a portion of the -131 to -94 sequence shows some homology to an NF-1 binding site (see Fig. 2B), an unlabeled double-stranded oligonucleotide containing an NF-1 binding site was also tested for its ability to compete for binding to the -131 to -94 oligomer. However, the complex was not competed by the NF-1 oligomer (not shown).

Interestingly, although the -131 to -94 sequence shares no homology with an ERE, the band was competed by a 25-fold excess of unlabeled ERE (Fig. 6B, lane 12) and more fully by a 50-fold excess of unlabeled ERE (lane 9), but was not competed by 200-fold excess mutated ERE (lane 5) or consensus glucocorticoid response element (GRE) (lane 4). The band was not supershifted in the presence of an anti-ER antibody (not shown), suggesting that ER was not present in the complex. These results suggest that a protein in the complex is also capable of interacting with ERE or with ER when it is bound to its response element in a manner that disrupts the binding of the labeled oligo. Because ER is present in the MCF-7 cell nuclear extract, this protein may be titrated away from the complex upon the addition of excess competing ERE.

The effect of ligand treatments on the formation of the DNA-protein complex was also examined (Fig. 6B). There was no difference in complex formation using nontreated (lanes 6 and 10) vs. ICI-treated cell extracts (Fig. 6, lane 2). Notably, using cell extracts treated with E₂, there was a marked decrease in the intensity of the shifted complex (lane 1). Cell extracts treated with TOT (lane 3) showed a gel shift pattern similar to, but slightly less strong than, that observed with ICI treatment. Therefore, differential binding of factors to the

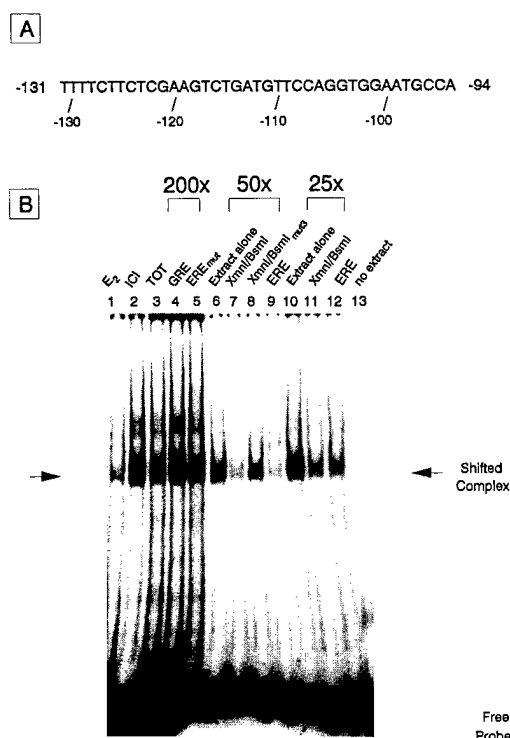


Fig. 6. Analysis of Protein Interactions with the -131 to -94 Region of PR_D

A, The sequence of the coding strand of the -131 to -94 region of PR_D and of the double-stranded oligomer used in the gel mobility shift assay. B, Gel mobility shift assays were performed using a double-stranded oligomer containing the -131 to -94 sequence of PR_D and extracts from MCF-7 cells as described in *Materials and Methods*. With extracts treated with E₂ (10⁻⁸ M, lane 1), or ICI (10⁻⁷ M, lane 2), or TOT (10⁻⁷ M, lane 3) for 15 min on ice; with extract + 200-fold excess unlabeled GRE (lane 4); with extract + 200-fold excess unlabeled ERE (lane 5); extract alone (lane 6); with extract + 50-fold excess unlabeled -131 to -94 double stranded oligo (lane 7); with extract + 50-fold excess unlabeled -131 to -94 double-stranded oligo with a 6-bp mutation from -115 to -110 (mut3, lane 8); with extract + 50-fold excess unlabeled ERE (lane 9); extract alone (lane 10); with extract + 25-fold excess unlabeled -131 to -94 double-stranded oligo (lane 11); with extract + 25-fold excess unlabeled ERE (lane 12); no extract (lane 13). The positions of the shifted complex and the free probe are indicated. The autoradiograph is representative of three separate experiments.

*Xmn*I/*Bsm*I fragment may occur in the presence of estrogens vs. AEs, and disappearance of the DNA-protein complex is correlated with the presence of transcriptionally productive, E₂-liganded ER.

DISCUSSION

Differential Sensitivity of Estrogen-Stimulated Promoters to the Inhibitory Actions of AEs

The experiments described herein have demonstrated the differential sensitivity of a number of estrogen-

regulated promoters, namely PR_D, PR_P and the pS2 promoter, to the actions of estrogens and AEs. In general, there was no correlation between the sensitivity of a given promoter to stimulation by estrogens when compared with its sensitivity to inhibition by AEs. For example, although the PR_D and the pS2 promoters required relatively comparable levels of E₂ for half-maximal stimulation, they showed markedly different dose-response profiles to the inhibitory actions of AEs. These findings suggest that the magnitude of estrogen responsiveness of a particular promoter is intrinsic to the nature of the promoter and that promoter responsiveness to the actions of estrogen- and AE-occupied ERs are separable. Furthermore, they implicate the involvement of inhibitory *cis* elements and promoter-specific factors acting to modulate the response of each promoter to different ER-ligand complexes. These findings are consistent with earlier reports in which it has been noted that reporter constructs containing EREs upstream of different promoters are differentially activated by estrogen in transient transfection assays, even when other experimental variables remain constant (10, 14), and with increasing evidence for promoter-specific actions of estrogens and AEs (5, 7-9, 14, 15).

Identification of an Inhibitory *cis* Element that Modulates the Sensitivity of Promoters to Estrogen- and AE-Occupied ER

Our search for ligand response modulatory elements began with our observation that the PR_D promoter showed reduced sensitivity to suppression by the AE-ER complex relative to the two other promoters, PR_P and pS2, examined. Deletion and mutational analyses led to the identification of a region in PR_D that, in fact, made the PR_D and a different estrogen-responsive (pS2) promoter more sensitive to inhibition by AE. Although these studies have allowed us to identify this novel element, which directs the AE sensitivity that PR_D does have, it is evident that the reduced AE sensitivity of PR_D overall must derive from activities from other portions of the promoter, which may normally act in concert with this element. Further analysis of PR_D would be of interest but may be complicated by our observation that more extensive deletions in the 5'-flanking region reduced activity altogether.

The identified element in PR_D had the following properties: 1) it reduced the magnitude and sensitivity of estrogen-stimulated activity, 2) it enhanced the ability of AEs to repress estrogen-stimulated activity, and 3) it elicited similar effects when transferred to the promoter of another estrogen-responsive gene. This LRME appears to have no clear homology to previously identified *cis* elements. Gel mobility shift assays showed that a cellular factor or factors were capable of binding to the element. Although we know very little about the nature of these factors, changes in the level or activity of these *trans*-acting factors would be predicted to play important roles in the gene-selective

actions of hormone- and antihormone-receptor complexes.

Relevant to our findings are reports from the Simons' laboratory (16, 17) of a *cis*-acting glucocorticoid modulatory element that, like the element we identify here in the PR gene, alters the sensitivity of the tyrosine aminotransferase (TAT) gene to glucocorticoid and to mixed agonist/antagonist antiglucocorticoids. The element differs, however, from the one we have identified in that it is located much further away from the promoter (3646 bp upstream of the start of TAT gene transcription). In addition, our LRME reduces the magnitude of E₂ stimulation or TOT agonism and increases the EC₅₀ for E₂ stimulation or TOT agonism, while the glucocorticoid-modulatory element enhances the sensitivity of the TAT gene to glucocorticoid (lower EC₅₀, *i.e.* left-shifted dose-response curve) and confers greater agonistic activity with partial agonist/antagonist antiglucocorticoids. However, the magnitudes of the shift in the dose-response curves (~10-fold) and the maximum activity levels (~2- to 3-fold) effected by the glucocorticoid-modulatory element and our LRME are very similar.

Of note, PR_D is a TATA-less promoter. However, the reduced AE sensitivity and the activity of the -131/-94 element is not exclusive to TATA-less promoters. For example PR_P, which is also TATA-less, shows strong sensitivity to AEs. Furthermore, the -131/-94 element can be transferred to the pS2 promoter, which is TATA-containing, and elicits the same activity.

Implications for Gene-Specific Regulation by Estrogens and AEs

Our results suggest that the sensitivity of a given promoter to the stimulatory actions of estrogens is not necessarily correlated with its sensitivity to the inhibitory actions of AEs. Furthermore, we have demonstrated that the presence of a modulatory *cis* element in the promoter region of a gene can dramatically influence the response of that promoter to agonist- and antagonist-occupied receptor. Ligand response-modulatory elements, such as we have identified in the PR_D promoter, may participate in regulating the activity of different estrogen-responsive genes by altering the pharmacology of estrogen and AE ligands that regulate these genes. They may thus be important in selectively modulating the properties of gene induction by estrogen agonists and antagonists and may underlie the known differences in dose-response curves for estrogen induction of different genes (5).

A BLAST search for this 38-bp *cis* element sequence in other genes revealed a related sequence (26-bp sequence, 84% identity) in the vinculin gene. Interestingly, vinculin, which encodes an actin-binding cytoskeletal protein, is also known to be under estrogen regulation (18). Thus, this sequence may potentially influence estrogen and AE sensitivity of several genes.

Our findings add to the growing list of modulators of ER activity. Modulators at almost every step in the

process of transcriptional activation by ER have been identified: the type of ligand, receptor phosphorylation (19-21), the sequence of the estrogen response element (Refs. 22 and 23 for reviews), coactivator proteins (such as TIF-1, SRC-1, SPT-6, and others) (24-29), some other nuclear hormone receptors (30-33), and chromatin structure (34, 35). The identification of so many potential modulators of ER activity suggests that transcriptional activation by ER is not a simple process and that there are many checkpoints in the process suitable for regulation.

The modulatory *cis* element that we have identified, which is capable of increasing the sensitivity of a promoter to the inhibitory actions of AEs, is especially interesting in light of the therapeutic uses of AEs in the treatment of breast cancer. A detailed understanding of the mechanisms by which this element and the factors that bind to it alter responsiveness to AEs may assist ultimately in the development of more effective therapeutic agents. In addition, since the ER is a member of a large superfamily of structurally and functionally related ligand-activated transcription factors, it is likely that similar *cis* elements, as identified previously in the glucocorticoid-regulated TAT gene (16, 17), will be found to modulate the sensitivity of genes regulated by other steroid receptors, and thyroid and retinoic acid receptors, to their agonist and antagonist ligands.

MATERIALS AND METHODS

Reagents and Radioisotopes

Cell culture media and antibiotics were purchased from GIBCO (Grand Island, NY). Calf serum was from Hyclone Laboratories (Logan, UT) and FCS from Sigma Chemical Company (St. Louis, MO). ³⁵S- and ³²P-radiolabeled nucleotides and [dichloroacetyl-1,2-¹⁴C]-chloramphenicol (50-60 Ci/mmol) were from Dupont/NEN Research Products (Boston, MA). Custom oligonucleotides were purchased from National Biosciences Inc. (Plymouth, MN). DNA restriction and modifying enzymes were from New England Biolabs (Beverly, MA), GIBCO/Bethesda Research Laboratory (Gaithersburg, MD), and U.S. Biochemicals (Cleveland, OH). DNA sequencing reagents were from U.S. Biochemicals. E₂ was from Sigma. The AEs ICI 164,384 and TOT were kindly provided by Alan Wakeling (Zeneca Pharmaceuticals, Macclesfield, U.K.). The AE LY 117,018 was kindly provided by Eli Lilly & Co. (Indianapolis, IN). All general reagents were of molecular biology grade and were purchased from Sigma Chemical Co., U.S. Biochemicals, or Fisher Scientific (Houston, TX).

Plasmid Construction and Mutagenesis

All cloning was done using standard techniques (36, 37). When necessary to make termini compatible, 3'- and 5'-overhangs generated by restriction digestion were blunted with T4 DNA polymerase and the Klenow fragment of *Escherichia coli* DNA polymerase, respectively. The insertion of double-stranded oligonucleotides and the deletion of DNA fragments were confirmed by dideoxy chain termination DNA sequencing. Other manipulations were confirmed by restriction digest analyses.

The construction of pTZ-TK-CAT, PR_D-CAT, PR_P-CAT, and (ERE)₂-PR_P-CAT (10), and 5E-PR_D-CAT (11), as well as

that of pS2-CAT (38), has been described previously. PR_{D,B/N}-CAT was constructed by releasing and blunting the *BsmI/NheI* fragment from the rat PR genomic clone EE(3.1)3Z (10) and cloning it into *SalI/BglII*-digested pTZ-TK-CAT. (ERE)₂-PR_{D,B/N}-CAT and (ERE/*HindIII*)₂-PR_D-CAT were made by annealing the single-stranded oligomers 5'-AATTAGT-CAGGTACAGTGACCTGATC-3' and 5'-AATTGATCAGGT-CACTGTGACCTGACT-3' and cloning two copies of the resultant double-stranded oligomer into the *HindIII* sites of PR_{D,B/N}-CAT and PR_D-CAT, respectively. (ERE)₂-pS2-CAT was made by annealing the single-stranded oligomers 5'-GATCCAAAGTCAGGTACAGTGACCTGATCAAAGA-3' and 5'-GATCTCTTTGATCAGGTCACTGTGACCTGACTT-TG-3' and cloning two copies of the resultant double-stranded oligomer into the *BamHI* site of pS2-CAT. (ERE/*HindIII*)₂-pS2-CAT was made by replacing the *BamHI/NcoI* fragment from (ERE/*HindIII*)₂-PR_D-CAT with the *BamHI/NcoI* fragment from pS2-CAT.

(X/B)_N-(ERE)₂-PR_D-CAT and (X/B)_N-(ERE)₂-PR_{D,B/N}-CAT were made by annealing the single-stranded oligomer 5'-TTTTCTTCTCGAAGTCTGATGTTCCAGGTGGAATGCC-3' with its complement and cloning one or two copies of the resultant double-stranded oligomer into *EagI*-digested and blunted (ERE/*HindIII*)₂-PR_D-CAT and (ERE/*HindIII*)₂-PR_{D,B/N}-CAT, respectively. (X/B)₂-(ERE)₂-pS2-CAT was made by replacing the *BamHI/NcoI* fragment from (ERE)₂-PR_D-CAT with the *BamHI/NcoI* fragment from pS2-CAT.

Six reporter constructs, each containing 6-bp mutations introduced sequentially from the -131 to -84 region of the rat PR gene distal promoter, were constructed by site-directed mutagenesis (39) with modifications (40). The *EcoRI* fragment of (ERE/*HindIII*)₂-PR_D-CAT was first inserted into the *EcoRI* site of Bluescript II SK⁺ (Stratagene, La Jolla, CA) to make (ERE/*HindIII*)₂-PR_D-BSK⁺. Mutagenic oligonucleotides were then annealed to single-stranded DNA generated using the f1 origin of replication in Bluescript II SK⁺. The mutagenic oligonucleotides used in six separate mutagenesis reactions were:

5'-ATCAGACTTCGATTCTGACGTCGACTCTAGAG-3'
5'-CCTGGAACATCACCATGGACGAAGAAAATCGA-3'
5'-GCATTCCACCTGAAGATATCGACTTCGAGAAG-3'
5'-TGGAGTTGGCATGGATCCAAGAACATCAGACT-3'
5'-TCCAAAACCTGGACAAGATCTCCACCTGGAAC-3'
5'-TGGCGAGATCCATTCATATGGTTGGCATTCCA-3'

To make each of the six (ERE/*HindIII*)₂-PR_{D,mut}-CAT reporter constructs, the *EcoRI/EcoRI* fragment of (ERE/*HindIII*)₂-PR_D-CAT was then replaced with the mutated *EcoRI/EcoRI* fragment of (ERE/*HindIII*)₂-PR_{D,mut}-BSK⁺. To simplify the reporter construct nomenclature used, we will refer to (ERE/*HindIII*)₂-PR_D-CAT, (ERE/*HindIII*)₂-PR_{D,B/N}-CAT, (ERE/*HindIII*)₂-PR_{D,mut}-CAT, and (ERE/*HindIII*)₂-pS2-CAT as (ERE)₂-PR_D-CAT, (ERE)₂-PR_{D,B/N}-CAT, (ERE)₂-PR_{D,mut}-CAT, and (ERE)₂-pS2-CAT, respectively.

The plasmid pCMVβ, which constitutively expresses β-galactosidase, was obtained from Clontech (Palo Alto, CA) and was used as an internal control for transfection efficiency in all experiments. The plasmid pTZ19, used as a carrier DNA, was provided by Dr. Byron Kemper of the University of Illinois.

DNA Preparation

Plasmid DNA for transfections was prepared on CsCl gradients as previously described (8, 37) or with a plasmid preparation kit (Qiagen, Chatsworth, CA).

Cell Culture and Transfections

MCF-7 cells (K1 subline, see Ref. 41) were maintained in MEM plus phenol red supplemented with 5% calf serum. For transfection experiments, the cells were switched to MEM plus phenol red supplemented with 5% charcoal-dextran-treated calf serum for 2 days, and then to MEM without

phenol red plus 5% charcoal-dextran-treated calf serum for 6 days before plating for transfection. All media included penicillin (100 U/ml) and streptomycin (100 μg/ml). For transfections, the cells were plated at a density of 3.5 × 10⁶ per 100-mm diameter dish and were given fresh medium about 30 h after plating. The cells were transfected by the calcium phosphate coprecipitation method (42) 16 h later with 15 μg of CAT reporter plasmid plus 400 ng of pCMVβ. The cells remained in contact with the precipitates for 6 h and were then subjected to a 3-min shock (25% glycerol in culture medium), which was followed by a rinse with HBSS. Treatments were added in fresh medium after the rinse.

β-Galactosidase and CAT Assays

All cells were harvested 24 h after hormone treatment. Extracts were prepared in 200 μl of 250 mM Tris-HCl (pH 7.5) using three freeze-thaw cycles. β-Galactosidase activity, which was measured to normalize for transfection efficiency, and CAT activity were assayed as previously described (43).

Gel Shift Assays

Whole cell extracts from MCF-7 cells for use in the gel shift assays were prepared by freeze-thaw lysis as described previously for transfected COS-1 cells (44). The single-stranded oligomer 5'-TTTTCTTCTCGAAGTCTGATGTTCCAGGTGGAATGCC-3', which represents the -131 to -94 region of the rat PR gene, was annealed with its complement. The resultant double-stranded oligomer was gel purified on a nondenaturing 10% polyacrylamide gel run in 0.5× Tris-borate-EDTA. The ability of extract protein(s) to bind to the -131 to -94 fragment was analyzed using standard gel mobility shift assays. Briefly, 2 μl (~5 μg) of MCF-7 whole cell extract was incubated with 1 ng of end-labeled -131/-94 oligomer, under conditions described previously (11). The specificity of binding was assessed by competition with excess unlabeled double-stranded -131/-94 oligomer or with excess unlabeled double-stranded -131/-94 oligonucleotide with a 6-bp mutation from -115 to -110 (mut3; single-stranded oligomer 5'-TTTTCTTCTCGAAGTCTGATGTTCCAGGTGGAATGCC-3' annealed to its complement) as well as with excess unlabeled double-stranded oligomers containing the consensus ERE, mutated ERE, or consensus GRE sequence. The nondenaturing gels used to analyze the protein-DNA complexes were run as described previously (11, 44).

Received October 14, 1996. Revision received December 17, 1996. Accepted December 19, 1996.

Address requests for reprints to: Dr. Benita S. Katzenellenbogen, Department of Molecular and Integrative Physiology, University of Illinois, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, Illinois 61801.

This work was supported by NIH Grant CA-18119 and US Army Grant DAMD17-94-J-4205. M.M.M. is the recipient of a postdoctoral fellowship from the Susan G. Komen Foundation.

*Present address: Department of Biology, University of California, San Diego, La Jolla, California 92093.

REFERENCES

1. Beato M, Herrlich P, Schütz G 1995 Steroid hormone receptors: many actors in search of a plot. *Cell* 83:851-857
2. Green S, Chambon P 1991 The oestrogen receptor: from perception to mechanism. In: Parker M (ed) *Nuclear Hor-*

- mone Receptors. Academic Press, New York, pp 15-38
3. Katzenellenbogen BS, Bhardwaj B, Fang H, Ince BA, Pakdel F, Reese JC, Schodin DJ, Wrenn CK 1993 Hormone binding and transcription activation by estrogen receptors: analyses using mammalian and yeast systems. *J Steroid Biochem Mol Biol* 47:39-48
4. Tsai M-J, O'Malley BW 1994 Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 63:451-486
5. Katzenellenbogen JA, O'Malley BW, Katzenellenbogen BS 1996 Tripartite steroid hormone receptor pharmacology: interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. *Mol Endocrinol* 10:119-131
6. Katzenellenbogen BS, Montano M, Le Goff P, Schodin DJ, Kraus WL, Bhardwaj B, Fujimoto N 1995 Antiestrogens: mechanisms and actions in target cells. *J Steroid Biochem Mol Biol* 53:387-393
7. Berry M, Metzger D, Chambon P 1990 Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-2818
8. Montano MM, Müller V, Trobaugh A, Katzenellenbogen BS 1995 The carboxyl-terminal F domain of the human estrogen receptor: role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 9:814-825
9. Tzukerman MT, Esty A, Santiso-Mere D, Daniellian P, Parker MG, Stein RB, Pike JW, McDonnell DP 1994 Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol Endocrinol* 8:21-30
10. Kraus WL, Montano MM, Katzenellenbogen BS 1993 Cloning of the rat progesterone receptor gene 5' region and identification of two functionally distinct promoters. *Mol Endocrinol* 7:1603-1616
11. Kraus WL, Montano MM, Katzenellenbogen BS 1994 Identification of multiple, widely spaced estrogen-responsive regions in the rat progesterone receptor gene. *Mol Endocrinol* 8:952-969
12. Brown AMC, Jeltsch JM, Roberts M, Chambon P 1984 Activation of pS2 gene transcription is a primary response to estrogen in the human breast cancer cell line MCF-7. *Proc Natl Acad Sci USA* 81:6344-6348
13. Corthesy B, Cardinaux JR, Claret FX, Wahli W 1989 A nuclear factor I-like activity and a liver-specific repressor govern estrogen-regulated *in vitro* transcription from the *Xenopus laevis* vitellogenin B1 promoter. *Mol Cell Biol* 9:5548-5562
14. Kumar V, Green S, Stack G, Berry M, Jin JR, Chambon P 1987 Functional domains of the human estrogen receptor. *Cell* 51:941-951
15. Fujimoto N, Katzenellenbogen BS 1994 Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: antiestrogen-selectivity and promoter-dependence. *Mol Endocrinol* 8:296-304
16. Oshima H, Simons Jr SS 1992 Modulation of transcription factor activity by a distant steroid modulatory element. *Mol Endocrinol* 6:416-428
17. Simons Jr SS, Oshima H, Szapary D 1992 Higher levels of control: modulation of steroid hormone-regulated gene transcription. *Mol Endocrinol* 6:995-1002
18. Depasquale JA, Samsonoff WA, Gierthy JF 1994 17- β -estradiol induced alterations of cell-matrix and intercellular adhesions in a human mammary carcinoma cell line. *Science* 107:1241-1245
19. Ali S, Metzger D, Bornert JM, Chambon P 1993 Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J* 12:1153-1160
20. Arnold SF, Obourn JD, Jaffe H, Notides AC 1994 Serine 167 is the major estradiol-induced phosphorylation site on the human estrogen receptor. *Mol Endocrinol* 8:1208-1214
21. Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS 1994 Phosphorylation of the human estrogen receptor: identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem* 269:4458-4466
22. Freedman LP, Luisi BF 1993 On the mechanism of DNA binding by nuclear hormone receptors: a structural and functional perspective. *J Cell Biochem* 51:140-150
23. Martinez E, Wahli W 1991 Characterization of hormone response elements. In: Parker M (ed) *Nuclear Hormone Receptors*. Academic Press, New York, pp 125-151
24. Baniahmad C, Nawaz Z, Baniahmad A, Gleeson MAG, Tsai M-J, O'Malley BW 1995 Enhancement of human estrogen receptor activity by SPT6: a potential coactivator. *Mol Endocrinol* 9:34-43
25. Cavaillès V, Dauvois S, L'Horsset F, Lopez G, Hoare S, Kushner PJ, Parker MG 1995 Nuclear factor RIP140 modulates transcriptional activation by the estrogen receptor. *EMBO J* 14:3741-3751
26. Halachmi S, Marden E, Martin G, MacKay H, Abbondanza C, Brown M 1994 Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription. *Science* 264:1455-1458
27. Le Douarin B, Zechel C, Garnier J, Luyt Y, Tora L, Pierrat B, Heery D, HG, Chambon P, Losson R 1995 The N-terminal part of TIF1, a putative mediator of the ligand-dependent activation function (AF-2) of nuclear receptors, is fused to B-raf in the oncogenic protein T18. *EMBO J* 14:2020-2033
28. Onate SA, Tsai SY, Tsai MJ, O'Malley BW 1995 Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270:1354-1357
29. Horwitz KB, Jackson TA, Bain DL, Richer JK, Takimoto GS, Tung T 1996 Nuclear receptor coactivators and corepressors. *Mol Endocrinol* 10:1167-1177
30. Burbach JP, da Silva SL, Cox JJ, Adan RA, Cooney AJ, Tsai M, Tsai S 1994 Repression of estrogen-dependent stimulation of the oxytocin gene by chicken ovalbumin promoter transcription factor I. *J Biol Chem* 269:15046-15053
31. Kraus WL, Weis KE, Katzenellenbogen BS 1995 Inhibitory cross-talk between steroid hormone receptors: differential targeting of estrogen receptor in the repression of its transcriptional activity by agonist- and antagonist-occupied progesterone receptors. *Mol Cell Biol* 15:1847-1857
32. Liu Y, Yang N, Teng CT 1993 COUP-TF acts as a competitive repressor for estrogen receptor-mediated activation of the mouse lactoferrin gene. *Mol Cell Biol* 13:1836-1846
33. McDonnell DP, Goldman ME 1994 RU486 exerts antiestrogenic activities through a novel progesterone receptor A form-mediated mechanism. *J Biol Chem* 269:11945-11949
34. Chiba H, Muramatsu M, Nomoto A, Kato H 1994 Two human homologues of *Saccharomyces cerevisiae* SW12/SNF2 and *Drosophila brahma* are transcriptional coactivators cooperating with the estrogen receptor and the retinoic acid receptor. *Nucleic Acids Res* 22:1815-1820
35. Gothard LQ, Hibbard JC, Seyfred MA 1996 Estrogen-mediated induction of rat prolactin gene transcription requires the formation of a chromatin loop between the distal enhancer and proximal promoter regions. *Mol Endocrinol* 10:185-195
36. Ausubel FM, Brent R, Kingston RE, Moore DD, Seidman JG, Smith JA, Struhl K 1992 *Short Protocols in Molecular Biology*, ed 2. John Wiley & Sons, New York
37. Sambrook J, Fritsch EF, Maniatis T 1989 *Molecular Cloning: A Laboratory Manual*, ed 2. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY
38. Aronica SM, Kraus WL, Katzenellenbogen BS 1994 Estrogen action via the cAMP signaling pathway: stimula-

- tion of adenylate cyclase and cAMP-regulated gene transcription. *Proc Natl Acad Sci USA* 91:8517-8521
39. Kunkel TA 1985 Rapid and efficient site-specific mutagenesis without phenotypic selection. *Proc Natl Acad Sci USA* 82:488-492
40. Mead D, Szczesna-Skorupa E, Kemper B 1986 Single-stranded DNA 'blue' T7 promoter plasmids: a versatile tandem promoter system for cloning and protein engineering. *Protein Eng* 1:67-74
41. Clarke R, Brünner N, Katzenellenbogen BS, Thompson EW, Norman MJ, Koppi C, Soonmyoung P, Lippman ME, Dickson RB 1989 Progression of human breast cancer cells from hormone-dependent to hormone-independent growth both *in vitro* and *in vivo*. *Proc Natl Acad Sci USA* 86:3649-3653
42. Chen C, Okayama H 1987 High efficiency transformation of mammalian cells by plasmid DNA. *Mol Cell Biol* 7:2745-2752
43. Reese JC, Katzenellenbogen BS 1991 Mutagenesis of cysteines in the hormone binding domain of the human estrogen receptor: alterations in binding and transcriptional activation by covalently and reversibly attaching ligands. *J Biol Chem* 266:10880-10887
44. Reese JR, Katzenellenbogen BS 1991 Differential DNA-binding abilities of estrogen receptor occupied with two classes of antiestrogens: studies using human estrogen receptor overexpressed in mammalian cells. *Nucleic Acids Res* 19:6595-6602

Erratum

In the article, "Identification of the sequences within the human complement 3 promoter required for estrogen responsiveness provides insight into the mechanism of tamoxifen mixed agonist activity" by J. Norris, D. Fan, and D. P. McDonnell (*Molecular Endocrinology* 10: 1605-1616, 1996), the Acknowledgments should have read as follows.

The authors would like to thank the friends and colleagues who offered helpful insights and suggestions during the course of this work and E. Allegretto for critical reading of the manuscript. We wish to thank Peter Kushner, Carol Anderson and Rosalie Uht (UCSF) for the plasmids SG5-HEO and E-GAL-E, and for sharing with us unpublished data relevant to our studies. We acknowledge also the gift of reagents from Dr. Geoff Greene, Dr. Eric Larson, and Dr. Alan Wakeling.

William L. McGuire Memorial Lecture

Antiestrogens: Mechanisms of action and resistance in breast cancer¹

Benita S. Katzenellenbogen, Monica M. Montano, Kirk Ekena, Mary E. Herman, and Eileen M. McNerney
Departments of Molecular and Integrative Physiology, and Cell and Structural Biology, University of Illinois and University of Illinois College of Medicine, Urbana, IL 61801-3704, USA

Key words: antiestrogens, tamoxifen, estrogen receptor, antiestrogen resistance, breast cancer, hormone sensitivity, endocrine therapy, resistance

Summary

Antiestrogens have proven to be highly effective in the treatment of hormone-responsive breast cancer. However, resistance to antiestrogen therapy often develops. In addition, although tamoxifen-like antiestrogens are largely inhibitory and function as estrogen antagonists in breast cancer cells, they also have some estrogen-like activity in other cells of the body. Thus, recent efforts are being directed toward the development of even more tissue-selective antiestrogens, i.e. compounds that are antiestrogenic on breast and uterus while maintaining the beneficial estrogen-like actions on bone and the cardiovascular system. Efforts are also being directed toward understanding ligand structure-estrogen receptor (ER) activity relationships and characterizing the molecular changes that underlie alterations in parallel signal transduction pathways that impact on the ER. Recent findings show that antiestrogens, which are known to exert most of their effects through the ER of breast cancer cells, contact a different set of amino acids in the hormone binding domain of the ER than those contacted by estrogen, and evoke a different receptor conformation that results in reduced or no transcriptional activity on most genes.

Resistance to antiestrogen therapy may develop due to changes at the level of the ER itself, and at pre- and post-receptor points in the estrogen receptor-response pathway. Resistance could arise in at least four ways: (1) ER loss or mutation; (2) Post-receptor alterations including changes in cAMP and phosphorylation pathways, or changes in coregulator and transcription factor interactions that affect the transcriptional activity of the ER; (3) Changes in growth factor production/sensitivity or paracrine cell-cell interactions; or (4) Pharmacological changes in the antiestrogen itself, including altered uptake and retention or metabolism of the antiestrogen. Model cell systems have been developed to study changes that accompany and define the antiestrogen resistant versus sensitive breast cancer phenotype. This information should lead to the development of antiestrogens with optimized tissue selectivity and agents to which resistance may develop more slowly. In addition, antiestrogens which work through somewhat different mechanisms of interaction with the ER should prove useful in treatment of some breast cancers that become resistant to a different category of antiestrogens.

¹ Presented at the 19th Annual San Antonio Breast Cancer Symposium, December 12, 1996.

Address for correspondence and offprints: Professor Benita S. Katzenellenbogen, Department of Molecular and Integrative Physiology, University of Illinois, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, Illinois 61801-3704, USA; *Tel:* 217-333-9769 or 217-333-7838; *Fax:* (217) 244-9906; *e-mail:* katzenel@uiuc.edu

Introduction

Tamoxifen, an antiestrogen in use for over 20 years, is the most commonly utilized agent in the treatment of hormone-responsive breast cancer. It is usually considered the treatment of choice because of its effectiveness and ease of use [1-4]. Recent clinical trials have confirmed the benefit of antiestrogens in preventing breast cancer recurrence and improving disease-free survival [5,6]. Tamoxifen may also be of benefit in preventing the development of breast cancer in women at high risk for the disease, a hypothesis being tested currently in major trials in the United Kingdom and the United States, although some concerns about its safety in long-term use have been raised [3].

Despite the clear effectiveness of antiestrogens, such that approximately 40% of breast cancer patients benefit substantially from such treatment, many of these women eventually suffer relapse because some of the breast cancer cells become resistant to tamoxifen. In addition, although tamoxifen-like antiestrogens are largely inhibitory and function as estrogen antagonists in breast cancer cells, they also have some estrogen-like activity in other cells of the body. Since women taking antiestrogen for breast cancer may be on prolonged therapy, the estrogen-like activities of tamoxifen become significant. Its stimulatory effects on uterus and liver may underlie the increased incidence of endometrial hyperplasia that may lead to cancer, as well as alterations in liver function [7-9]. On the other hand, the estrogen-like activities of tamoxifen are beneficial in bone cells and in the cardiovascular system, where this agent enhances bone maintenance, preserves a favorable blood lipid profile, and reduces risk of coronary problems [5,6,8-12]. Recently developed pure antiestrogens, such as ICI 164,384, ICI 182,780, and RU54,876, may perhaps prove to be more effective than tamoxifen in treating hormone-responsive breast cancer, but are not effective in preventing bone loss and may have detrimental effects on the cardiovascular system [13-17]. By altering the chemical struc-

ture of antiestrogens, it should be possible to potentiate their estrogen-like actions on bone and the cardiovascular system, but not their stimulatory activities in breast and uterus, while maintaining an appropriate balance of activities in the liver. Optimism in this regard is buoyed by the fact that there has already been the development of antiestrogens, termed selective estrogen receptor modulators (SERMs), that appear to show improved tissue selectivity in their actions [18,19].

Therefore, research with antiestrogens is aimed toward the development of agents that will circumvent or delay the onset of resistance, and ones that may show even greater tissue selectivity in their actions.

Mechanisms of action of antiestrogens

Since antiestrogens are believed to exert their beneficial effects in breast cancer cells by working largely through the estrogen receptor (ER) in these cells, we have focused much of our attention on understanding the interactions of antiestrogens and estrogens with this receptor protein. We will first summarize some of the current thinking about estrogen receptor action and the mechanisms by which antiestrogens suppress the activity and transcriptional effectiveness of the ER.

Antiestrogens are hormonal agents that act through the ER to regulate gene transcription [1,2,9]. Their pharmacology, however, is complex, and subtle differences in their structure, as well as alterations in the cellular milieu in which they are acting, can have marked effects on the level of their agonist or antagonist activities in different target tissues and on specific responses within these tissues [9,20-23]. These differences prove to be crucial in their uses for the prevention and treatment of breast cancer, as is known from experience with the few antiestrogens that have been extensively studied in women.

The presence of the estrogen receptor has proven to be important in predicting improved

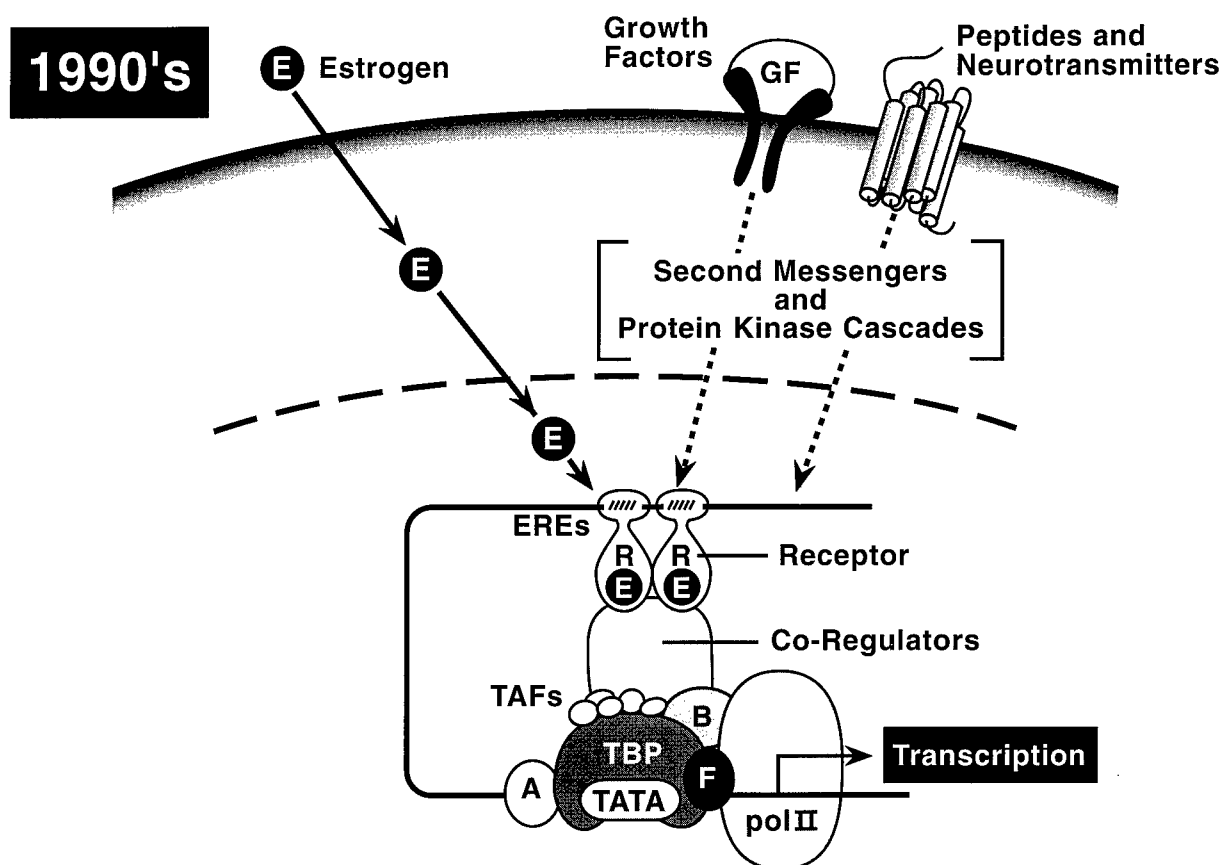


Figure 1. A model for estrogen receptor (ER) actions. The abbreviations used are E, estrogen; R, receptor; ERE, estrogen response element; GF, growth factor; TBP, TATA binding protein; TAFs, TBP-associated factors; pol II, RNA polymerase II. See text for details.

disease-free survival and in predicting response to tamoxifen therapy. In the absence of the estrogen receptor and progesterone receptor, response to tamoxifen is observed in only approximately 5% of breast cancer patients, while the presence of substantial levels of both the estrogen receptor and progesterone receptor predict that response to tamoxifen will occur in up to *ca.* 75% of such patients.

Studies on the estrogen receptor and its mechanisms and actions, begun about 25 years ago, indicated that the receptor interacted with chromatin after hormone binding, resulting in increases in specific mRNAs and hormone-induced proteins. In the 1990's model for estrogen action, shown in Figure 1, it is clear that our

understanding of estrogen action has expanded considerably beyond that of the 1970's. The sub-cellular distribution of the receptor is thought to be largely nuclear even in the absence of hormone. Very significantly, there are — besides the hormone and the estrogen receptor — other factors termed coregulators, as well as gene- and promoter-specific factors, that are crucial in regulating the activity of the receptor in target cells [9,24]. Other cell signaling pathways also impact on the bioactivity of the ER, and some of these aspects are discussed later in this article. These include modulation of ER activity by growth factors (including EGF, IGF-1, HER2/neu), neurotransmitters such as dopamine, and second messengers such as cAMP and others that

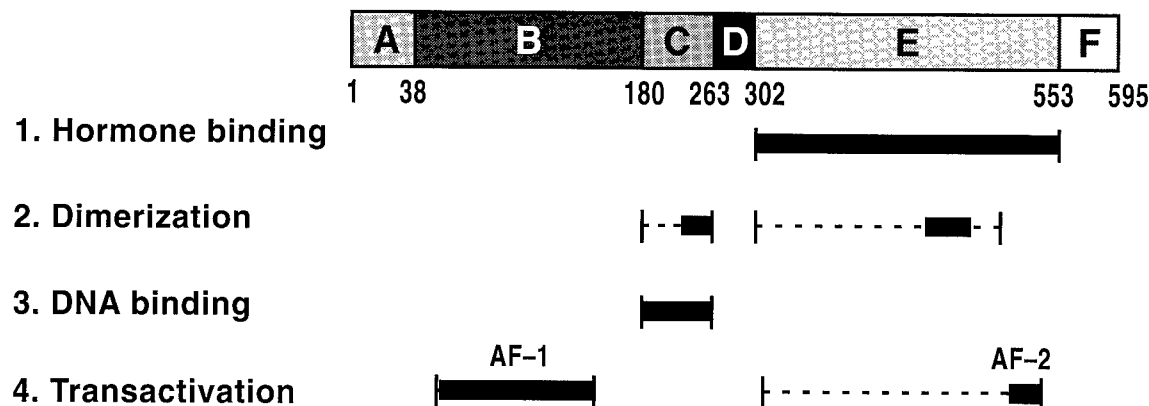


Figure 2. Schematic of the human estrogen receptor. The structural domains of ER (A/B, C, D, E, and F), as well as the hormone binding, dimerization, DNA binding, and transactivation (AF-1, AF-2) functional domains are shown. AF, activation function.

affect protein kinase cascades including the MAP kinase signaling pathway [2,8,25-29, and refs. therein].

We also now know a great deal more about this receptor protein, and how it interacts with estrogen and antiestrogen ligands and with other protein factors that regulate its transcriptional activity. The estrogen receptor (ER) is a 66 kilodalton, ligand-dependent transcription factor which regulates the transcription of estrogen-responsive genes (for reviews see [1,2,24,30]). Like other steroid hormone receptors, the ER is a modular protein (Figure 2) which can be divided into separable domains with specific functions, such as ligand binding, dimerization, DNA binding, and transactivation. In addition to a centrally located C domain, corresponding to the DNA binding domain, the ER contains two distinct activation functions [20,22,23,30]. The activation function located in the N-terminal A/B domain is termed activation function-1 (AF-1), and a second, hormone-dependent activation function (AF-2) is located in the E domain along with the hormone binding function of ER. AF-1 and AF-2 function in a synergistic manner and are required for full ER activity in most cell contexts [20,22,23,30]. Like other activation domains, the acti-

vation functions of ER are thought to be important targets for basal transcription factors or specific cellular proteins which function as co-activators. Of note, the activity of each of the two activation functions of the ER varies in different cellular contexts. Region F of the receptor, the most carboxyl-terminal domain, is not essential for hormone binding or transactivation, but we have shown that region F affects the agonist and antagonist activity of antiestrogens [31].

Binding of estrogen to the estrogen receptor stimulates the increased expression of some genes, including those for some growth factors and growth factor receptors resulting in the stimulation of DNA synthesis and cell proliferation, as well as the increased production of proteins such as plasminogen activator and collagenases that are believed to enhance the metastatic capability of breast cancer cells [32,33]. When antiestrogen binds to the estrogen receptor, the receptor is not available to bind estrogen, and the antiestrogen-estrogen receptor complex fails to effectively stimulate gene expression and DNA synthesis; instead, the receptor-antiestrogen complex enhances production of some growth inhibitory factors, including the TGF- β s, thereby preventing breast cancer growth

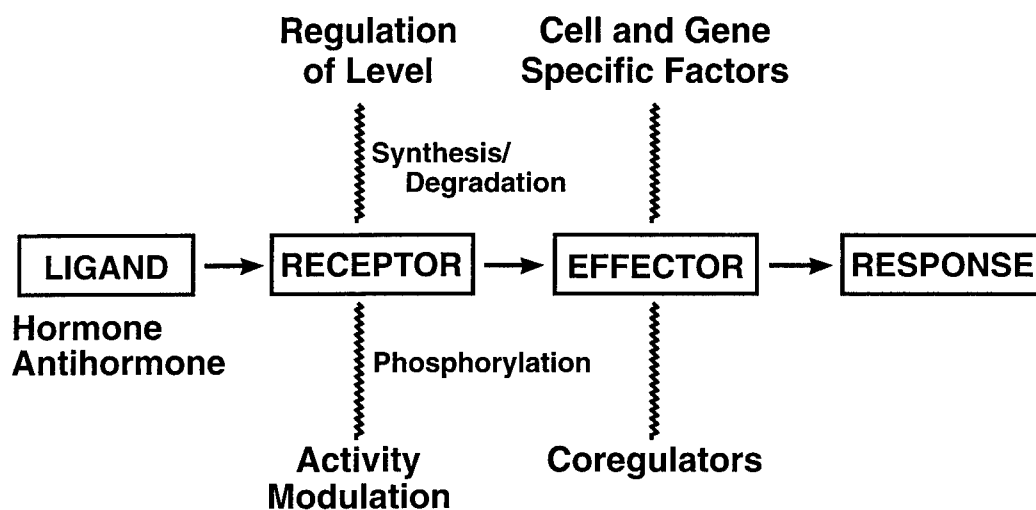


Figure 3. The flow of information from ligand-receptor-effector to response initiated by the binding of hormone (estrogen) or antihormone (antiestrogen) with the estrogen receptor. The response to a hormone is mediated by a tripartite interaction involving the ligand, the receptor, and effector sites through which the ligand-receptor complex regulates the response. The modulation of receptor activity by its state of phosphorylation is also indicated. See text for details.

and metastasis [34,35]. Recent findings indicate that antiestrogens also have anti growth-factor effects [28,36,37]; by changing the conformation of the estrogen receptor (and in the case of ICI164,384, also the concentration of estrogen receptor in cells [38,39]), antiestrogens can result in the inhibition of some growth factor-regulated genes. Antiestrogens effectively suppress angiogenesis and induce apoptosis, both beneficial in blocking tumor growth and development, and in evoking cell cycle arrest and killing of breast cancer cells [40,41]. Antiestrogens also increase the expression of wild type BRCA1, a tumor suppressive protein [42].

Studies have shown that the response of genes to estrogen and antiestrogen depends on several important factors: namely, the nature of the estrogen receptor (whether it is wild type or variant); the nature of the gene promoter; the cell context; and the ligand. The role of the recently characterized estrogen receptor subtype ER β [43,44] in mammary gland and breast cancer is currently unknown and needs to be investigated. In addition, gene responses elicited by the ER may be modulated by cAMP, growth factors, and agents that

affect protein kinases and cell phosphorylation. These may account for differences observed in the relative agonism/antagonism of compounds like tamoxifen with different genes and in different target cells. Thus, tamoxifen is a very effective antagonist of estrogen action in breast cancer, while having significant estrogen-like agonistic activity in uterus and bone. As shown in Figure 3, the biological response to an antiestrogen depends on differences in the interaction of anti-hormone versus hormone with the receptor, and differences in the coupling of these ligand-receptor complexes with the various effectors (cell-specific and gene-specific factors and coregulators) that determine the biological response, such as inhibition of cell proliferation by antiestrogens. As is discussed later, the state of phosphorylation of the estrogen receptor plays an important role in modulating receptor activity.

There are several modes of estrogen receptor activation of genes (Figure 4). Three of the different modes of gene activation by the liganded ER complex are shown. In system 1, there is interaction of receptor with the estrogen response element and direct interaction with general trans-

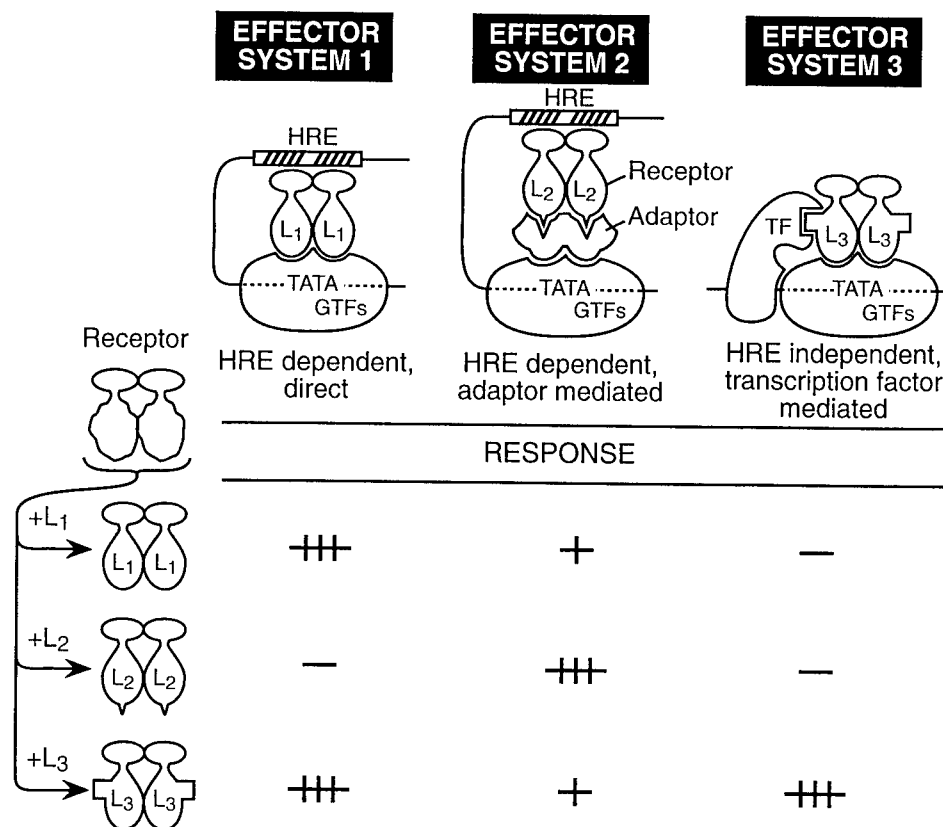


Figure 4. Different modes of nuclear receptor activation of genes. The top of this scheme illustrates three different modes for nuclear receptor activation of genes; for each mode, an optimal ligand-receptor-effector combination is shown. The bottom of the scheme illustrates the activity that each of the three ligand-receptor complexes might have at each of the three effector sites. Note that the receptor adopts a different conformation in its complex with the three ligands, and that these different "shapes" affect the nature of the receptor-effector coupling. In a tripartite scheme, the potency of a ligand is determined largely by its affinity of interaction with the receptor, but its biocharacter is determined by the interaction that the ligand-receptor complex has with various effector sites. Therefore, for each receptor, the biocharacter (and to some degree the potency) of a hormone cannot be uniquely assigned without reference to a specific response and effector interaction. Other modes of nuclear receptor gene activation than the three illustrated here are known, such as the remodeling of nucleosomal and chromatin architecture by hormone receptor complexes. However, for simplicity, only three are shown here as examples. The abbreviations are L, ligand; HRE, hormone response element; GTFs, general transcriptional factors; TF, transcription factor. (From ref [9].)

cription factors. In system 2, the DNA interaction is the same, but interaction with the general transcription factors is mediated by an adaptor or coregulator protein. In system 3, an estrogen response element (ERE) is not involved, and instead interaction with DNA is indirect, via a transcription factor such as Fos and Jun. These different modes of receptor activation of genes allow for considerable diversity in mechanism of gene turn-on, and can account for the fact that the

agonist/antagonist activity of a ligand such as tamoxifen may be response-specific. Since the shape that the receptor assumes around each ligand will be somewhat different, this can result in differential stimulation or failure of activation of different genes [9].

We and others [1,2,8, and refs. therein] have used affinity labeling with irreversibly attaching ligands, along with mutagenesis and deletion analyses, to study estrogen receptor ligand-receptor-

response relationships and to define regions and amino acid residues in the receptor that are critical for ligand binding and discrimination between estrogens and antiestrogens. These studies identified a region in the hormone binding domain near cysteine 530 that was important in discriminating between estrogen and antiestrogen [45], and a region from amino acid 510 to 530 that is very important for hormone binding [46-49]. Some point mutations in other parts of the E domain were also found to affect the affinity and temperature stability of hormone binding [50]. Through the use of alanine scanning mutagenesis across the amino acid 515-535 region of the receptor, we have identified four residues most important in estradiol binding (amino acids 521, 524, 525, and 528) and have observed that these form a compact unit on one face of a proposed α -helix in the hormone binding domain (Figure 5; [51]). Interestingly, the footprint over this region of the ER is somewhat different with antiestrogen, implying that receptor conformation is different with antiestrogen versus estrogen [52]. Proteolytic digestion studies on the antiestrogen liganded or estrogen liganded ER also support the view that receptor conformation is different with these different types of ligands [53,54].

The structures of some antiestrogens are shown in Figure 6. The antiestrogens can be non-steroidal or steroidal, based on either non-steroidal or steroidal estrogens, and usually have a bulky side chain that is charged or polar. The side chain is essential for antiestrogenic activity, as removal of the side chain results in a compound that shows only estrogenic activity. Recent studies have documented that changes both in the side chain and in the linker region can alter the relative agonistic and antagonistic activity of antiestrogens. In addition, by modifying the chemical nature of the antiestrogen, it is possible to generate antiestrogens that are purer estrogen antagonists [13-15], and also to develop compounds that may potentially have greater tissue selectivity [18,19], being strong antagonists in breast cancer cells, while showing little or no stimulation of uterus but maintaining estrogen-like

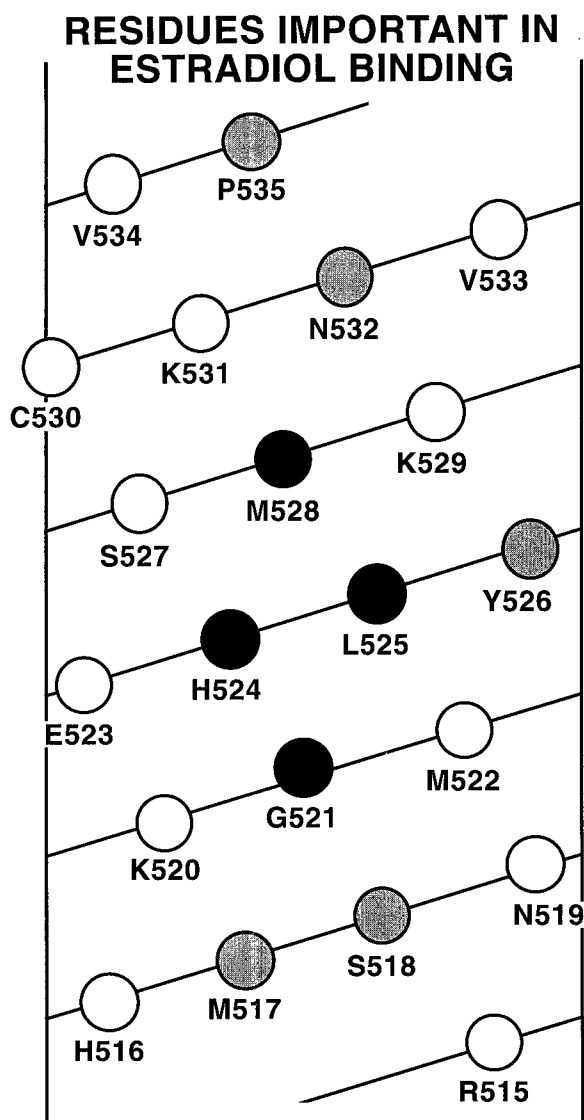


Figure 5. Residues in the region from amino acid 515 to 535 of the human estrogen receptor that are most important in estradiol binding. A helical face map of the 515-535 region of the human estrogen receptor is shown (α -helix split longitudinally and opened up). Darkly shaded circles represent positions where alanine substitution inhibits estradiol activity of the receptor 40-95%. Lightly shaded circles represent positions where alanine substitution inhibits estradiol activity of the receptor 20-40%. Note that the four residues most important in estradiol binding (residues 521, 524, 525, and 528) reside in a compact unit on one face of a proposed α -helix. (From ref [51].)

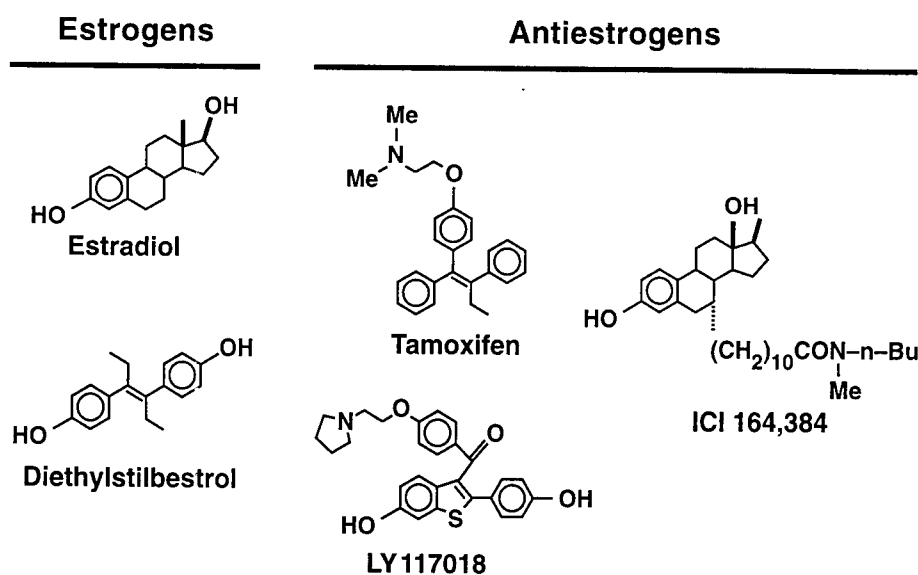


Figure 6. Ligands for the estrogen receptor. The structures of two estrogens and three antiestrogens are shown.

activity in bone along with cardiovascular and lipid profile benefits.

Different antiestrogens display a different spectrum or balance of agonist and antagonist activity. While antiestrogens have in the past been referred to as type I (partial antagonist) compounds such as tamoxifen, and type II (pure antagonist) compounds such as ICI 164,384 or ICI 182,780, it is now clear that there is a spectrum of activity that is often target cell- and gene-dependent [54,55]. In all cases, antiestrogen binding to the estrogen receptor, which occurs in a manner competitive with that of estradiol, results in a different receptor conformation. These conformational differences are manifest in different patterns of proteolytic cleavage [53,54] and coregulator interaction [56-59]. With compounds such as tamoxifen, binding to receptor fails to activate the hormone-dependent transcription activation function in domain E of the receptor (AF-2), while having no effect on the hormone-independent activation function (AF-1) in the A/B region of the receptor. Thus, tamoxifen is a partial agonist/partial antagonist on different genes, dependent on the promoter and the cell content of cell-specific factors and coregulators. With purer

antagonists, such as ICI 164,384, the receptor assumes a different conformation. This often results in acceleration of the rate of receptor degradation such that ER levels in breast cancer cells decline more rapidly over time. The reduced ER levels and the different conformation of any remaining ICI 164,384-receptor complexes result in a situation in which activation of transcription by the hormone-dependent AF-2 region in domain E, as well as the constitutive-transcriptional activation through AF-1 in the N-terminal A/B region, is thus not possible [20,31,38,39,54].

Estrogens have been shown to promote a ligand-dependent transcriptionally productive interaction of the amino- and carboxyl- terminal activation function regions of the estrogen receptor, allowing for optimal transcriptional activity of the receptor [23]. By contrast, when antiestrogen binds to the receptor, the antiestrogen-occupied receptor exhibits conformational changes that are distinct and different from those induced by estrogen. These lead to association of the amino- and carboxyl- terminal regions, but this interaction is transcriptionally unproductive. Thus, antiestrogens generally fail to activate gene transcription, or they do so only poorly [23,60,61].

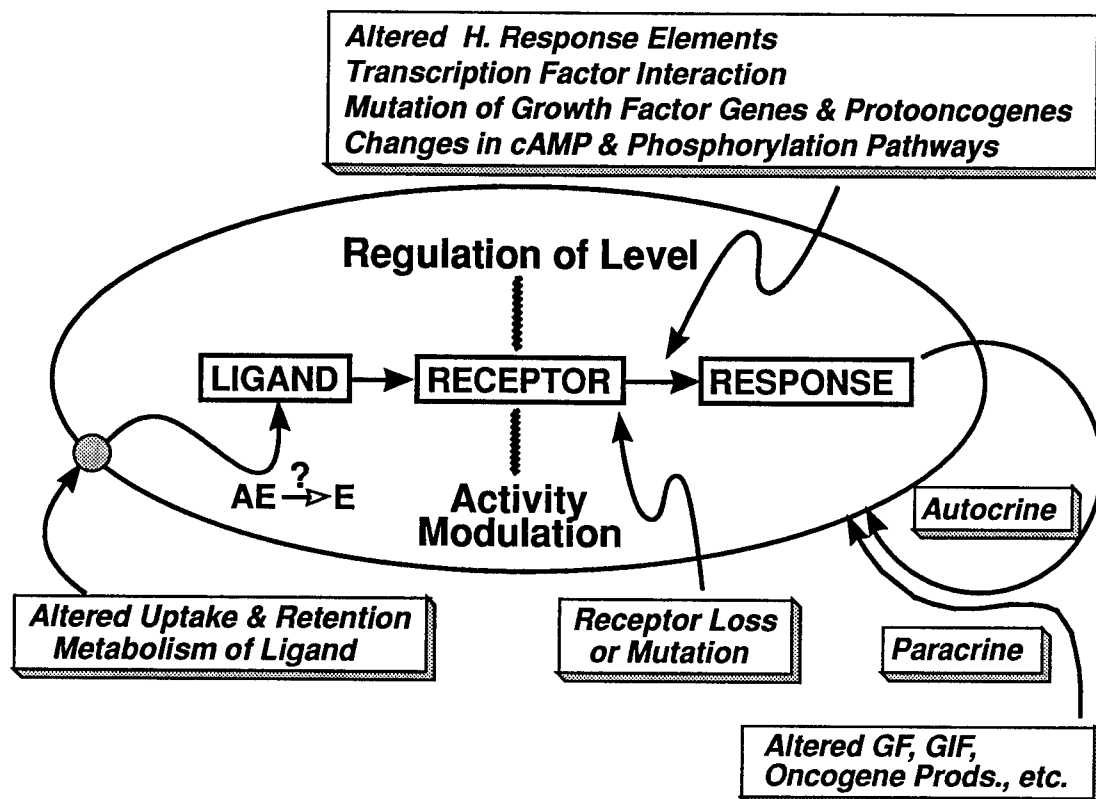


Figure 7. A model to explain cellular changes that may underlie hormonal resistance. Four major ways in which hormonal resistance may arise are shown in the boxed areas of the Figure. See text for details.

Analysis of antiestrogen resistance in breast cancer

One of the major problems in long-term, effective endocrine therapy for breast cancer is the development of hormonal resistance, in particular, resistance to antiestrogen therapy [3,62,63]. There are at least four major ways in which hormonal resistance could arise (Figure 7, boxed areas). First, there could be estrogen receptor loss or mutation, and there is evidence for this in human breast tumors [64-71], but this probably accounts for only a portion, perhaps 20%, of antiestrogen-resistant tumors [72]. Second, there may also be post-receptor alterations. These include changes in cAMP and phosphorylation pathways, which are known to affect the transcriptional activity of the receptor and to enhance the agonistic

activity of tamoxifen-like antiestrogens [55]. There may also be possible alterations in hormone response elements, coregulator and transcription factor interactions, or mutations of growth factor genes and protooncogenes [3]. Third, there may also be changes in growth factor production/sensitivity, i.e. altered production of autocrine factors or paracrine interactions from adjacent estrogen receptor negative breast cancer cells or stromal cells [73]. Fourth, there may be pharmacologic alterations in the antiestrogen itself, including altered uptake and retention, or metabolism of the antiestrogen [74]. There is evidence from work in several laboratories that changes in each of these four aspects can result in hormonal resistance. Changes may thus be at the level of the estrogen receptor itself, and at pre- or post-receptor points in the receptor-response pathway.

Although resistance would clearly result from the loss of the estrogen receptor protein or might be due to the presence of variant estrogen receptors in breast cancers, for which there is already considerable evidence [64-72], it is likely that such receptor variants account for only a portion of hormone-resistant breast cancers. As shown through the studies described above, point mutations in discrete regions in domain E of the estrogen receptor would eliminate estrogen or antiestrogen binding, and therefore eliminate response to either of these ligands. Likewise, changes in the levels of splice variant forms of ER may affect antiestrogen sensitivity [64,71]. Other changes in the DNA-binding ability of ER in breast tumors have also been reported [75]. In the carboxyl-terminal region of domain E, mutations can result in receptors which bind hormone but are altered in activation function-2 activity. While some of these mutant receptors fail to respond to either estrogen or antiestrogen [46], certain changes in this region, corresponding to the proposed helix 12 of the estrogen receptor, result in receptors which show no response to estrogen but surprisingly, can now be activated by antiestrogen. These ligand activity inversion mutants show inverted response to ligand — antiestrogen is now seen as a stimulator and estrogen as an antagonist [39,76]. With such a mutation, a tumor would be stimulated by antiestrogen rather than being suppressed by it. Furthermore, other specific point mutational changes in the activation function-2 region of the estrogen receptor result in estrogen receptors that show differential response to partial and pure antiestrogens [39]. These and related findings [45, 77,78] emphasize that even single amino acid changes can dramatically affect the conformation of the receptor and its functional interaction with the transcriptional machinery, resulting in receptors that may be either fully inactive or partially active, or in receptors that now misinterpret the nature of the ligand (i.e. see some antiestrogens as estrogens and estrogens as antagonists). They also highlight that the ER can distinguish exquisitely among different estrogen and antiestrogen

ligands.

It is clear, however, that in many cases, alterations in hormonal sensitivity/resistance occur despite the presence of significant levels of apparently normal estrogen receptors [73,79-81]. Perhaps most importantly, clinical experience has shown that hormonal resistance is often reversible, suggesting a cellular adaptation mechanism rather than a genetic alteration in many breast cancer patients. For example, patients who become resistant to tamoxifen often respond immediately to treatments with high dose estrogen or return to a state of tamoxifen responsiveness after a period of alternative therapy [82-84]. Therefore, any mechanism that would explain this form of tamoxifen resistance in these patients, would have to involve mechanisms that would be reversible or adaptational, in contrast to other mechanisms for tamoxifen resistance that might involve mutations in the estrogen receptor or other critical transcription factor or growth factor genes.

It has now been well documented that estrogen receptor activity is regulated by phosphorylation, and this may be, at least in part, how growth factors and cAMP influence estrogen receptor activity. Our studies have shown that cAMP and some growth factors enhance ER transcriptional activity, increase ER phosphorylation, and change the agonist/antagonist balance of some antiestrogens [2,8,55,85]. Agents which increased intracellular cAMP levels in MCF-7 breast cancer cells resulted in tamoxifen becoming more agonistic and a weaker antagonist of estrogen-stimulated transcriptional activity. In contrast, the purer antiestrogen ICI 164,384 did not have its transcriptional activity affected by increasing intracellular cAMP. Even in the presence of elevated intracellular cAMP, ICI 164,384 remained a complete estrogen antagonist without any intrinsic stimulatory activity [55].

In this regard, it is noteworthy that cAMP levels are significantly higher in breast tumors than in normal breast tissue or fibrocystic breast tissue [86,87] and that elevated concentrations of cAMP binding proteins are associated with early

disease recurrence and poor survival rates. It has been shown that the tumor content of cAMP binding proteins serves as a highly significant prognostic factor, equal in utility to that of the estrogen receptor, in predicting disease-free and overall survival in breast cancer [88,89], and that the content of cAMP binding proteins, in combination with estrogen receptor measurements, is very useful in identifying endocrine responsive tumors [90,91].

Thus, mutational changes in the ER itself, and changes in cAMP and phosphorylation pathways, could contribute to hormonal resistance. Changes in growth factor pathways that are normally under estrogen and antiestrogen regulation appear also to contribute to hormonal resistance as described below.

To understand better the antiestrogen-resistant phenotype that frequently develops in breast cancer patients receiving tamoxifen, we cultured MCF-7 breast cancer cells long-term (> 1 year) in the presence of the antiestrogen *trans*-hydroxy-tamoxifen (TOT) to generate a subline refractory to the growth-suppressive effects of TOT [73]. This subline (designated MCF/TOT) showed growth stimulation, rather than inhibition, with TOT and diminished growth stimulation with estradiol (E_2), yet remained as sensitive as the parental cells to growth suppression by another antiestrogen, ICI 164,384. Estrogen receptor (ER) was maintained at 40% of the level in the parent MCF-7 cells, but MCF/TOT cells failed to show an increase in progesterone receptor content in response to E_2 or TOT treatment. In contrast, the MCF/TOT subline behaved like parental cells in terms of E_2 and TOT regulation of ER and pS2 expression and transactivation of a transiently transfected estrogen-responsive gene construct. DNA sequencing of the hormone binding domain of the ER from both MCF-7 and MCF/TOT cells confirmed the presence of wild-type ER and exon 5 and exon 7 deletion splice variants, but showed no point mutations. Compared to the parental cells, the MCF/TOT subline showed reduced sensitivity to the growth-suppressive effects of retinoic acid and complete resistance to exogenous

TGF- β 1. The altered growth responsiveness of MCF/TOT cells to TOT and TGF- β 1 was partially to fully reversible following TOT withdrawal for 16 weeks. These findings underscore the facts that antiestrogen resistance is response-specific; that loss of growth suppression by TOT appears to be due to the acquisition of weak growth stimulation; and that resistance to TOT does not mean global resistance to other purer antiestrogens such as ICI 164,384, implying that these antiestrogens must act by somewhat different mechanisms. The association of reduced retinoic acid responsiveness and insensitivity to exogenous TGF- β with antiestrogen growth-resistance in these cells supports the increasing evidence for interrelationships among cell regulatory pathways utilized by these three growth-suppressive agents in breast cancer cells. In addition, our findings indicate that one mechanism of antiestrogen resistance, as seen in MCF/TOT cells, may involve alterations in growth factor and other hormonal pathways that affect the ER response pathway.

What we have learned from this model system is that the breast cancer cells which were originally suppressed by tamoxifen have become no longer growth inhibited by this antiestrogen and are in fact weakly stimulated by it, as shown by others as well [74,92,93]. The resistance to tamoxifen is partially reversible following removal of tamoxifen, suggesting cell adaptation rather than mutational changes in this model cell system. In addition, the MCF/TOT cells produce high levels of TGF- β , yet grow very rapidly and are not inhibited either by antiestrogen or by the TGF- β s that they are producing or that we add to their culture media. This resistance to TGF- β is not attributable to loss of TGF- β receptors, implying a possible change in the TGF- β signaling pathway [73]. On-going studies, employing differential display methods with mRNA from tamoxifen-resistant and tamoxifen-sensitive breast cancer cells, should allow a better understanding of the tamoxifen-resistant phenotype.

Hormonal resistance, therefore, can involve estrogen receptor and post-receptor changes. We

know that resistance can result from mutational changes in the estrogen receptor; changes in pathways (i.e. growth factors and cAMP) that impact on the estrogen receptor and the phosphorylation state of the cell; changes in co-regulators which interact with the estrogen receptor; and changes in growth factor pathways which are normally under estrogen and antiestrogen regulation (TGF- α , TGF- β s) and may now become constitutive.

What advances does the future hold? It is clear that a better understanding of the estrogen receptor-response pathway and further development of modified antiestrogen ligands should result in antiestrogens with improved tissue selectivity and agents that may engender resistance more slowly. At a minimum, new generation antiestrogens should provide an armament of reagents that will prove to be highly beneficial should resistance to one antiestrogenic agent develop. For example, it is already known that purer antiestrogens, which act through a somewhat different receptor mechanism than do the tamoxifen-like antiestrogens, are of benefit in some breast cancer patients when resistance to tamoxifen develops [17,94-96]. New information on ligand structure-receptor activity relationships and the characterization of molecular changes that underlie alterations in parallel signal transduction pathways that impact on the ER should lead to the development of new antiestrogens even more effective and tissue selective than those currently available for the treatment and ultimate prevention of breast cancer.

Acknowledgements

Research from our laboratory described in this article was supported by grants to B.S.K. from the NIH (2R37 CA18119 and CA60514) and US Army (DAMD-17-94-J-4205), and by postdoctoral fellowships from the Susan G. Komen Foundation (M.M.M.) and NIH (1F32 CA68653, K.E.)

References

1. Katzenellenbogen BS, Fang H, Ince BA, Pakdel F, Reese JC, Wooge CH, Wrenn CK: Estrogen receptors: Ligand discrimination and antiestrogen action. *Breast Cancer Res Treat* 27:17-26, 1993
2. Katzenellenbogen BS, Montano M, Le Goff P, Schodin DJ, Kraus WL, Bhardwaj B, Fujimoto N: Antiestrogens: Mechanisms and actions in target cells. *J Steroid Biochem Molec Biol* 53:387-393, 1995
3. Osborne CK, Elledge RM, Fuqua SAW: Estrogen receptors in breast cancer therapy. *Sci Am Sci Med* 3:32-41, 1996
4. Jordan VC, Murphy CS: Endocrine pharmacology of antiestrogens as antitumor agents. *Endocr Rev* 11: 578-610, 1990
5. Fisher B, Dignam J, Bryant J, DeCillis A, Wickerham DL, et al: Five versus more than five years of tamoxifen therapy for breast cancer patients with negative lymph nodes and estrogen receptor-positive tumors. *J Natl Cancer Inst* 88:1529-1542, 1996
6. Swedish Breast Cancer Cooperative Group: Randomized trial of two versus five years of adjuvant tamoxifen for postmenopausal early stage breast cancer. *J Natl Cancer Inst* 88:1543-1549, 1996
7. Jordan VC, Morrow M: Should clinicians be concerned about the carcinogenic potential of tamoxifen? *Eur J Cancer* 30A:1714-1721, 1994
8. Katzenellenbogen BS: Estrogen receptors: Bioactivities and interactions with cell signaling pathways. *Biol Reprod* 54:287-293, 1996
9. Katzenellenbogen JA, O'Malley BW, Katzenellenbogen BS: Tripartite steroid hormone receptor pharmacology: Interaction with multiple effector sites as a basis for the cell- and promoter-specific action of these hormones. *Mol Endocrinol* 10:119-131, 1996
10. Sarrel PM, Lufkin EG, Oursler MJ, Keefe D: Estrogen actions in arteries, bone and brain. *Sci Am Sci Med* 1:44-53, 1994
11. Kneifel MA, Katzenellenbogen BS: Comparative effects of estrogen and antiestrogen on plasma renin substrate levels and hepatic estrogen receptors in the rat. *Endocrinology* 108:545-552, 1981
12. Toney TW, Katzenellenbogen BS: Antiestrogen action in the medial basal hypothalamus and pituitary of immature female rats: insights concerning relationships among estrogen, dopamine and prolactin. *Endocrinology* 119:2661-2669, 1986
13. Wakeling AE, Bowler J: Biology and mode of action of pure antiestrogens. *J Steroid Biochem* 30:141-147, 1988
14. Wakeling AE, Dukes M, Bowler J: A potent specific pure antiestrogen with clinical potential. *Cancer Res* 51:3867-3873, 1991

15. Nique F, Van de Velde P, Bremaud J, Hardy M, Philibert D, Teutsch G: 11 β -amidoalkoxyphenyl estradiols, a new series of pure antiestrogens. *J Steroid Biochem Molec Biol* 50:21-29, 1994
16. Nicholson RI, Gee JMW, Francis AB, Manning DL, Wakeling AE, Katzenellenbogen BS: Observations arising from the use of pure antioestrogens on oestrogen-responsive (MCF-7) and oestrogen growth-independent (K3) human breast cancer cells. *Endocrine Related Cancer* 2:115-121, 1995
17. Wakeling AE: The future of new pure antiestrogens in clinical breast cancer. *Breast Cancer Res Treat* 25:1-9, 1993
18. Sato M, Rippey MK, Bryant HU: Raloxifene, tamoxifen, nafoxidine, or estrogen effects on reproductive and nonreproductive tissues in ovariectomized rats. *FASEB J* 10:905-912, 1996
19. Yang NN, Venugopalan M, Hardikar S, Glasebrook A: Identification of an estrogen response element activated by metabolites of 17 β -estradiol and raloxifene. *Science* 273:1222-1225, 1996
20. Berry M, Metzger D, Chambon P: Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-2818, 1990
21. Montano MM, Muller V, Trobaugh A, Katzenellenbogen BS: The carboxy terminal F-domain of the human estrogen receptor: role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 9:814-825, 1995
22. Tzukerman MT, Esty A, Santiso-Mere D, Danielian P, Parker MG, Stein RB, Pike JW, McDonnell DP: Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol Endocrinol* 8:21-30, 1994
23. Kraus WL, McInerney EM, Katzenellenbogen BS: Ligand-dependent, transcriptionally productive association of the amino- and carboxyl-terminal regions of a steroid hormone nuclear receptor. *Proc Natl Acad Sci USA* 92:12314-12318, 1995
24. Tsai MJ, O'Malley BW: Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 63:451-486, 1994
25. Kato S, Endoh H, Masuhiro Y, Kitamoto T, Uchiyama S, Sasaki H, Masushige S, Gotoh Y, Nishida E, Kawashima H, Metzger D, Chambon P: Activation of the estrogen receptor through phosphorylation by mitogen-activated protein kinase. *Science* 270:1491-1494, 1995
26. Pietras RJ, Arboleda J, Reese DM, Wongvipat N, Pegram MD, Ramos L, Gorman CM, Parker MG, Sliwkowski MX, Slamon DJ: HER-2 tyrosine kinase pathway targets estrogen receptor and promotes hormone-independent growth in human breast cancer cells. *Oncogene* 10:2435-2446, 1995
27. Cho H, Katzenellenbogen BS: Synergistic activation of estrogen receptor-mediated transcription by estradiol and protein kinase activators. *Molec Endocrinol* 7:441-452, 1993.
28. Cho H, Aronica SM, Katzenellenbogen BS: Regulation of progesterone receptor gene expression in MCF-7 breast cancer cells: a comparison of the effects of cyclic AMP, estradiol, IGF-1 and serum factors. *Endocrinology* 134:658-664, 1994
29. Ignar-Trowbridge DM, Nelson KG, Bidwell MC, Curtis SW, Washburn TF, McLachlan JA, Korach KS: Coupling of dual signaling pathways: Epidermal growth factor action involves the estrogen receptor. *Proc Natl Acad Sci USA* 89:4658-4662, 1992
30. Gronemeyer H: Transcriptional regulation by estrogen and progesterone receptors. *Annu Rev Genet* 25:89-123, 1991
31. Montano MM, Müller V, Trobaugh A, Katzenellenbogen BS: The carboxy-terminal F domain of the human estrogen receptor: Role in the transcriptional activity of the receptor and the effectiveness of antiestrogens as estrogen antagonists. *Mol Endocrinol* 9:814-825, 1995
32. Read LD, Katzenellenbogen BS: Characterization and regulation of estrogen and progesterone receptors in breast cancer. In: Dickson RB, Lippman ME (eds) *Genes, Oncogenes and Hormones: Advances in Cellular and Molecular Biology of Breast Cancer*. Kluwer Academic Publishers, Boston, 1991, pp 277-299
33. Dickson RB, Lippman ME: Estrogenic regulation of growth and polypeptide growth factor secretion in human breast carcinoma. *Endocr Rev* 8:29-43, 1987
34. Knabbe C, Lippman ME, Wakefield LM, Flanders KC, Kasid A, Derynck R, Dickson RB: Evidence that transforming growth factor- β is a hormonally regulated negative growth factor in human breast cancer cells. *Cell* 48:417-428, 1987
35. Knabbe C, Zugmaier G, Schmahl M, Dietel M, Lippman ME, Dickson RB: Induction of transforming growth factor β by the antiestrogens droloxifene, tamoxifen, and toremifene in MCF-7 cells. *Am J Clin Oncol (CCT)* 14(Suppl 2):S15-S20, 1991
36. Vignon F, Bouton MM, Rochefort H: Antiestrogens inhibit the mitogenic effect of growth factors on breast cancer cells in the total absence of estrogens. *Biochem Biophys Res Commun* 146:1502-1508, 1987
37. Katzenellenbogen BS, Norman MJ: Multihormonal regulation of the progesterone receptor in MCF-7 human breast cancer cells: Interrelationships among insulin/IGF-I, serum and estrogen. *Endocrinology* 126:891-898, 1990

38. Dauvois S, Danielian PS, White R, Parker MG: Anti-estrogen ICI 164,384 reduces cellular estrogen receptor content by increasing turnover. *Proc Natl Acad Sci USA* 89:4037-4041, 1992
39. Montano MM, Ekena KE, Krueger KD, Keller AL, Katzenellenbogen BS: Human estrogen receptor ligand activity inversion mutants: Receptors that interpret antiestrogens as estrogens and estrogens as antiestrogens and discriminate among different antiestrogens. *Mol Endocrinol* 10:230-242, 1996
40. Haran EF, Maratzek AF, Goldberg I, Horwitz A, Degani H: Tamoxifen enhances cell death in implanted MCF7 breast cancer by inhibiting endothelium growth. *Cancer Res* 54:5511-5514, 1994
41. McCloskey DE, Armstrong DK, Jackisch C, Davidson NE: Programmed cell death in human breast cancer cells. *Recent Prog Horm Res* 51:493-508, 1996
42. Jensen RA, Thompson ME, Jetton TL, Szabo CI, van der Meer R, Helou B, Tronick SR, Page DL, King MC, Holt JT: BRCA1 is secreted and exhibits properties of a granin. *Nature Genetics* 12:303-308, 1996
43. Kuiper GGJM, Enmark E, Peltö-Huikko M, Nilsson S: Cloning of a novel estrogen receptor expressed in rat prostate and ovary. *Proc Natl Acad Sci USA* 93: 5925-5930, 1996
44. Mosselman S, Polman J, Dijkema R: ER β : identification and characterization of a novel human estrogen receptor. *FEBS Letters* 392:49-53, 1996
45. Pakdel F, Katzenellenbogen BS: Human estrogen receptor mutants with altered estrogen and antiestrogen ligand discrimination. *J Biol Chem* 267:3429-3437, 1992
46. Wrenn CK, Katzenellenbogen BS: Structure-function analysis of the hormone binding domain of the human estrogen receptor by region-specific mutagenesis and phenotypic screening in yeast. *J Biol Chem* 268: 24089-24098, 1993
47. Katzenellenbogen BS, Bhardwaj B, Fang H, Ince BA, Pakdel F, Reese JC, Schodin DJ, Wrenn CK: Hormone binding and transcription activation by estrogen receptors: Analyses using mammalian and yeast systems. *J Steroid Biochem Molec Biol* 47:39-48, 1993
48. Fawell SE, Lees JA, White R, Parker MG: Characterization and colocalization of steroid binding and dimerization activities in the mouse estrogen receptor. *Cell* 60:953-962, 1990
49. Danielian PS, White R, Hoare SA, Fawell SF, Parker MG: Identification of residues in the estrogen receptor that confer differential sensitivity to estrogen and hydroxytamoxifen. *Mol Endocrinol* 7:232-240, 1993
50. Reese JC, Katzenellenbogen BS: Characterization of a temperature-sensitive mutation in the hormone binding domain of the human estrogen receptor: Studies in cell extracts and intact cells and their implications for hormone-dependent transcriptional activation. *J Biol Chem* 267:9868-9873, 1992
51. Ekena KE, Weis KE, Katzenellenbogen JA, Katzenellenbogen BS: Identification of amino acids in the hormone binding domain of the human estrogen receptor important in estrogen binding. *J Biol Chem* 271: 20053-20059, 1996
52. Ekena K, Weis KE, Katzenellenbogen JA, Katzenellenbogen BS: Different residues of the human estrogen receptor are involved in the recognition of structurally diverse estrogens and antiestrogens. *J Biol Chem* 272: 5069-5075, 1997
53. Allan GF, Leng XH, Tsai SY, Weigel NL, Edwards DP, Tsai M-J, O'Malley BW: Hormone and antihormone induce distinct conformational changes which are central to steroid receptor activation. *J Biol Chem* 267:19513-19520, 1992
54. McDonnell DP, Clemm DL, Hermann T, Goldman ME, Pike JW: Analysis of estrogen receptor function in vitro reveals three distinct classes of antiestrogens. *Mol Endocrinol* 9:659-669, 1995
55. Fujimoto N, Katzenellenbogen BS: Alteration in the agonist/antagonist balance of antiestrogens by activation of protein kinase A signaling pathways in breast cancer cells: Antiestrogen-selectivity and promoter-dependence. *Mol Endocrinol* 8:296-304, 1994
56. Oñate SA, Tsai SY, Tsai M-J, O'Malley BW: Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270: 1354-1357, 1995
57. Halachmi S, Marden E, Martin G, MacKay H, Abbonanza C, Brown M: Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription. *Science* 264:1455-1458, 1994
58. Cavallès V, Dauvois S, Danielian PS, Parker MG: Interaction of proteins with transcriptionally active estrogen receptors. *Proc Natl Acad Sci USA* 91: 10009-10013, 1994
59. Weis KE, Ekena K, Thomas JA, Lazennec G, Katzenellenbogen BS: Constitutively active human estrogen receptors containing amino acid substitutions for tyrosine 537 in the receptor protein. *Mol Endocrinol* 10:1388-1398, 1996
60. McInerney EM, Katzenellenbogen BS: Different regions in activation function-1 of the human estrogen receptor required for antiestrogen-dependent and estradiol-dependent transcription activation. *J Biol Chem* 271:24172-24178, 1996
61. McInerney EM, Tsai MJ, O'Malley BW, Katzenellenbogen BS: Analysis of estrogen receptor transcriptional enhancement by a nuclear hormone receptor coactivator. *Proc Natl Acad Sci USA* 93:10069-10073, 1996
62. Katzenellenbogen BS: Antiestrogen resistance: Mechanisms by which breast cancer cells undermine the

- effectiveness of endocrine therapy. *J Natl Cancer Inst* 83:1434-1435, 1991
63. Touchette N: Tamoxifen resistance in breast cancer. *J NIH Research* 4:67-72, 1992
64. McGuire WL, Chamness GC, Fuqua SAW: Estrogen receptor variants in clinical breast cancer. *Mol Endocrinol* 5:1571-1577, 1991
65. Miksicek RJ, Lei Y, Wang Y: Exon skipping gives rise to alternatively spliced forms of the estrogen receptor in breast tumor cells. *Breast Cancer Res Treat* 26:163-174, 1993
66. Fuqua SAW, Fitzgerald SD, Allred DC, Elledge RM, Nawaz Z, McDonnell DP, O'Malley BW, Greene GL, McGuire WL: Inhibition of estrogen receptor action by a naturally occurring variant in human breast tumors. *Cancer Res* 52:483-486, 1992
67. Garcia T, Sanchez M, Cox JL, Shaw PA, Ross JBA, Lehrer S, Schachter B: Identification of a variant form of the human estrogen receptor with an amino acid replacement. *Nucleic Acids Res* 17:8364-8368, 1989
68. Murphy LC, Dotzlaw H: Variant estrogen receptor mRNA species detected in human breast cancer biopsy samples. *Mol Endocrinol* 3:687-693, 1989
69. McGuire WL, Chamness GC, Fuqua SAW: The importance of normal and abnormal oestrogen receptor in breast cancer. *Cancer Surveys* 14:31-40, 1992
70. Fuqua SAW, Chamness GC, McGuire WL: Estrogen receptor mutations in breast cancer. *J Cell Biochem* 51:135-139, 1993
71. Castles CG, Fuqua SAW: Alterations within the estrogen receptor in breast cancer. In: Pasqualini JR, Katzenellenbogen BS (eds) *Hormone Dependent Cancer*. Marcel Dekker, New York, 1996, pp 81-105
72. Karnik PS, Kulkarni S, Liu X, Budd GT, Bukowski RM: Estrogen receptor mutations in tamoxifen-resistant breast cancer. *Cancer Res* 54:349-353, 1994
73. Herman ME, Katzenellenbogen BS: Response-specific antiestrogen resistance in a newly characterized MCF-7 human breast cancer cell line resulting from long-term exposure to trans-hydroxytamoxifen. *J Steroid Biochem Molec Biol* 59:121-134, 1996
74. Osborne CK, Coronado E, Allred DC, Wiebe V, DeGregorio M: Acquired tamoxifen resistance: Correlation with reduced breast tumor levels of tamoxifen and isomerization of trans-4-hydroxytamoxifen. *J Natl Cancer Inst* 83:1477-1482, 1991
75. Scott GK, Kushner P, Vigne J-L, Benz CC: Truncated forms of DNA-binding estrogen receptors in human breast cancer. *J Clin Invest* 88:700-706, 1991
76. Mahfoudi A, Roulet E, Dauvois S, Parker MG, Wahli W: Specific mutations in the estrogen receptor change the properties of antiestrogens to full agonists. *Proc Natl Acad Sci USA* 92:4206-4210, 1995
77. Wolf DM, Jordan VC: The estrogen receptor from a tamoxifen stimulated MCF-7 tumor variant contains a point mutation in the ligand binding domain. *Breast Cancer Res Treat* 31:129-138, 1994
78. Pakdel F, Reese JC, Katzenellenbogen BS: Identification of charged residues in an N-terminal portion of the hormone binding domain of the human estrogen receptor important in transcriptional activity of the receptor. *Mol Endocrinol* 7:1408-1417, 1993
79. Katzenellenbogen BS, Kendra KL, Norman MJ, Berthois Y: Proliferation, hormonal responsiveness, and estrogen receptor content of MCF-7 human breast cancer cells grown in the short-term and long-term absence of estrogens. *Cancer Res* 47:4355-4360, 1987
80. Clarke R, Brünner N, Katzenellenbogen BS, Thompson EW, Norman MJ, Koppi C, Soonmyoung P, Lippman ME, Dickson RB: Progression of human breast cancer cells from hormone-dependent to hormone-independent growth both in vitro and in vivo. *Proc Natl Acad Sci USA* 86:3649-3653, 1989
81. Jiang S, Wolf DM, Yingling JM, Chang C, Jordan VC: An estrogen receptor positive MCF-7 clone that is resistant to antiestrogens and estradiol. *Mol Cell Endocrinol* 90:77-86, 1992
82. Vassilomanolakis ME, Tsoussis S, Kandylis K, Hajichristou E, Efredmidis AP: Rechallenge by tamoxifen in metastatic breast cancer: Prospective study of different dose levels. *Breast Dis* 4:129-134, 1991
83. Stoll BA: Intermittent antiestrogen therapy in advanced breast cancer. *Cancer Treat Rep* 67:98, 1983
84. Stoll BA: Rechallenging breast cancer with tamoxifen therapy. *Clin Oncol* 9:347-351, 1983
85. Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS: Phosphorylation of the human estrogen receptor: Identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem* 269:4458-4466, 1994
86. Minton JP, Wisenbaugh T, Matthews RH: Elevated cyclic AMP levels in human breast-cancer tissue. *J Natl Cancer Inst* 53:283, 1974
87. Cohen LA, Chan P: Intracellular cAMP levels in normal rat mammary gland and adenocarcinoma: In vivo vs. in vitro. *Life Sci* 16:107-115, 1974
88. Miller WR, Elton RA, Dixon JM, Chetty U, Watson DMA: Cyclic AMP binding proteins and prognosis in breast cancer. *Br J Cancer* 61:263-266, 1990
89. Miller WR, Watson DMA, Jack W, Chetty U, Elton RA: Tumour cyclic AMP binding proteins: An independent prognostic factor for disease recurrence and survival in breast cancer. *Breast Cancer Res Treat* 26:89-94, 1993
90. Kvinnsland S, Ekanger R, Døskeland SO, Thorsen T: Relationship of cyclic AMP binding capacity and estrogen receptor to hormone sensitivity in human breast cancer. *Breast Cancer Res Treat* 3:67-72, 1983

91. Watson DMA, Hawkins RA, Bundred NJ, Stewart HJ, Miller WR: Tumour cyclic AMP binding proteins and endocrine responsiveness in patients with inoperable breast cancer. *Br J Cancer* 56:141-142, 1987
92. Gottardis MM, Jordan VC: Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res* 48:5183-5187, 1988
93. Osborne CK: Tamoxifen metabolism as a mechanism for resistance. *Endocrine-Related Cancer* 2:53-58, 1995
94. Howell A, DeFriend D, Robertson J, Blamey R, Walton P: Response to a specific antiestrogen (ICI 182780) in tamoxifen-resistant breast cancer. *Lancet* 345:29-30, 1995
95. Hu XF, Veroni M, De Luise M, Wakeling A, Sutherland R, Watts CKW, Zalcberg JR: Circumvention of tamoxifen resistance by the pure anti-estrogen ICI 182,780. *Int J Cancer* 55:873-876, 1993
96. Wakeling A: Physiological effects of pure antiestrogens. *In: Pasqualini JR, Katzenellenbogen BS (eds) Hormone Dependent Cancer*. Marcel Dekker, New York, 1996, pp 107-115

Mechanistic Aspects of Estrogen Receptor Activation Probed with Constitutively Active Estrogen Receptors: Correlations with DNA and Coregulator Interactions and Receptor Conformational Changes

Gwendal Lazennec, Tracy R. Ediger, Larry N. Petz,
Ann M. Nardulli, and Benita S. Katzenellenbogen

Department of Molecular and Integrative Physiology (G.L., L.N.P.,
A.M.N., B.S.K.) and
Department of Cell and Structural Biology (T.R.E., B.S.K.)
University of Illinois
Urbana, Illinois 61801

The estrogen receptor (ER) belongs to a large family of nuclear receptors, many of whose members function as ligand-dependent transcriptional activators. The mechanism by which the receptor is converted from an inactive into an activated state is not yet completely understood. To investigate the kind of changes in receptor conformation and interactions that are involved in this activation, we have used the wild type ER and a set of constitutively active ER point mutants that show from 20% to nearly 100% activity in the absence of estrogen. These mutants are of particular interest as they could mimic, in the absence of ligand, the activated state of the wild type receptor. We have analyzed several transcriptional steps that could be involved in the activation: the ability of these receptors 1) to interact with several coactivators (steroid receptor coactivator-1, SRC-1; transcription intermediary factor-1, TIF-1; and estrogen receptor-associated protein 140, ERAP 140) and with members of the preinitiation complex [TATA box-binding protein (TBP), transcription factor IIB (TFIIB)]; 2) to exhibit conformational changes revealed by proteolytic digest patterns similar to those observed for the wild type hormone-occupied ER; and 3) to bend estrogen response element-containing DNA, which is thought to be one of the important phenomena triggering transcriptional activation. Our results demonstrate that the interaction of these mutant receptors with coactivators is likely to be one of the features of the activated step, as the mutant receptors interacted with some coactivators in a ligand-independent manner in proportion to their extent of constitutive activity. However, the different

degrees of ligand-independent interaction of the mutant ERs with the three coactivators suggest that SRC-1, TIF-1, and ERAP 140 may play different roles in receptor activity. Limited proteolytic digest experiments reveal that the activated state of the receptor corresponds to a particular conformation of the receptor, which is fully observed with the mutant ER showing the highest activity in the absence of estrogen. Finally, it appears that in inactive or active states, the receptor exhibits distinctly different DNA-bending abilities. Addition of estradiol is able to modify the bending ability of only the wild type receptor, whereas estradiol has no influence on the constitutive receptors, which exhibited the same bending ability as that observed for the ligand-occupied wild type receptor. These data document that the ER undergoes major changes in its conformation and also in its functional properties when it is turned from an inactive into an active state and that mutational changes in the ER protein that result in constitutive, hormone-independent activation mimic many of the changes in ER properties that are normally under hormone regulation. (*Molecular Endocrinology* 11: 1375-1386, 1997)

INTRODUCTION

The estrogen receptor (ER) is a ligand-dependent transactivator that belongs to a large superfamily of nuclear receptors. Some members are active only in the presence of ligand, as is the case for the ER, but a large number have no identified ligand at the present time and could be, in some cases, ligand-independent factors (for review, see Ref. 1). All these receptors share a common structure of five domains named A/B,

C, D, E and F (2), and some key functions have been assigned to each domain. The N-terminal A/B domain contains the ligand-independent transcription activation function 1 (AF-1) (3, 4). The C domain has a characteristic helix-loop-helix structure stabilized by two zinc atoms and is responsible for the binding to estrogen response elements (EREs) (5, 6). The D domain appears to be a hinge region that can modulate the DNA-binding ability of the receptor (7). The E and F domains are involved in the ligand-binding function and exhibit also a strong ligand-dependent activation function (AF-2) (5, 8).

Numerous studies have focused on different aspects of the process by which the receptor is transformed from an inactive state in the absence of ligand to an activated state upon ligand exposure. However, this process is still not fully understood. In its inactive form, the receptor is associated with a number of other proteins (including at least hsp90, hsp70, and p23), forming a multiprotein complex with a sedimentation constant of 8–9 S (9–11). In the unliganded state, receptors for thyroid hormone (TR) or for retinoids [retinoic acid receptor (RAR) and retinoid X receptor (RXR)] can have an inhibitory effect on transcription and are associated with corepressors (12–17). Upon hormone exposure, most of the associated proteins are released, and the receptor appears as a 4–5 S sedimenting complex (18, 19). In this state, the receptor is able to dimerize and to associate with coactivators (20–27). The receptor is also known to undergo changes in its phosphorylation state (28, 29) and conformation (30, 31).

The aim of this study is to understand what kind of changes in function and conformation the receptor undergoes when it is converted to the active state. To address these questions, we have used the wild type ER and a set of three constitutive ER mutants that correspond to amino acid substitutions at residue 380 or residue 537 (32, 33). These mutants show constitutive activity in the absence of estradiol (E_2) ranging from 20 to nearly 100% of the activity of wild type receptor in the presence of E_2 . By comparing the properties of wild type and these mutant receptors in the absence and in the presence of estrogens or antiestrogens, we have analyzed their ability to interact with several known coactivators and members of the preinitiation complex (PIC). Moreover, the conformation of these receptors was studied by limited proteolytic digest experiments. Finally, we analyzed the ability of these receptors to bend ERE-containing DNA, which is thought to reflect in part the transcriptional ability of transcription factors (34–36). Altogether, these results demonstrate that, upon hormone exposure, the wild type receptor undergoes major changes in its conformation and in its properties, whereas no effect or only limited effects of hormone are observed with the constitutively active receptors, as these appear to have already undergone those changes that render them ligand-independent transcriptional activators.

RESULTS

Certain ER Mutants Show Constitutive Activity

To understand the mechanisms underlying the activated state of a normally hormone-activated nuclear receptor, we have analyzed the properties of the wild type ER as well as those of constitutively active mutant ERs. Several steps presumed to be involved in receptor activation upon ligand treatment were analyzed, *i.e.* the interaction of the wild type and mutant ERs with coactivators and members of the PIC, the conformational state of these receptors before and after treatment with estrogen, and their ability to bend ERE-containing DNA fragments.

Transcriptional activity of the wild type or mutant ERs was monitored in the presence and absence of hormone with estrogen-responsive CAT reporter genes containing a minimal (TATA) or a complex (pS2) promoter. As shown in Fig. 1, in the absence of ligand, the Y537S ER exhibited constitutive activity 65–90% that of the estrogen-occupied ER, the level of activity being dependent on the promoter used, while the Y537A and E380Q mutant ERs exhibited constitutive activity in the absence of ligand about 15% to 30% that of wild type activity in the presence of E_2 . Moreover, all three receptors were as potent as wild type ER in activating transcription in the presence of E_2 . The magnitude of constitutive activity of the ERs shown in Fig. 1 is consistent with our earlier findings (32, 33). Referring to the structure of the related nuclear receptors hRAR γ and rTR α 1, for which crystallographic information is available (37, 38), the Y537 residue would be in helix 12, containing the hormone-dependent activation function (AF-2) of nuclear receptors, and the E380 residue would be at the end of helix 4 of the ligand-binding domain. [Note: The helix numbering is from the RAR γ structure (37).]

Mutant and Wild Type ERs Interact in a Different Manner with Receptor-Associated Proteins

As numerous reports have emphasized the requirement of coactivator factors to promote full activity of nuclear receptors in the presence of their ligand, it was of interest to determine the extent to which wild type ER and the constitutively active mutants interacted with these factors. Pull-down experiments were performed utilizing glutathione-S-transferase (GST) fusion proteins with the hormone-binding domain of the different ERs. These fusions proteins were expressed in bacteria and adsorbed onto GSH-Sepharose columns. The interaction of coactivators with these ERs was then analyzed using *in vitro* translated coactivators, with equal inputs of coactivator in each experimental sample. Interactions were monitored in the absence of ligand and in the presence of estradiol or the antiestrogen *trans*-hydroxy-tamoxifen (TOT) (Fig. 2).

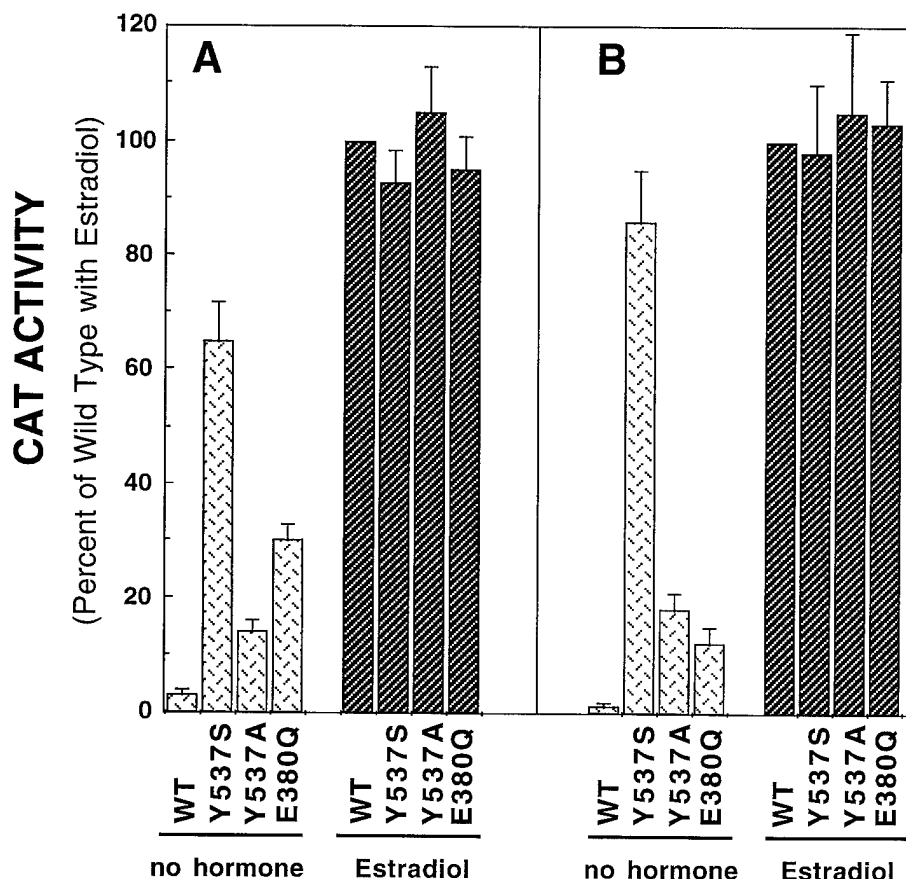


Fig. 1. Specific Mutations of the Tyrosine 537 and Glutamic Acid 380 Residues Create Constitutively Active ERs

Transcriptional activity of wild type or mutant (Y537S, Y537A, E380Q) ERs were monitored in the absence or presence of E_2 . A, CHO cells were cotransfected with wild type or mutant (Y537S, Y537A, E380Q) ER expression vectors, reporter gene construct 2ERE-TATA-CAT, and a pCH110 β -galactosidase internal reporter. Transfected cells were treated for 24 h with no hormone or E_2 (10^{-8} M). Values are the means and sds of three experiments after standardization with β -galactosidase activity and are expressed as the percentage of the CAT activity observed with wild type ER in the presence of E_2 . B, MDA-MB-231 cells were transfected in the same conditions as for CHO cells, except that the CAT reporter construct used was 2ERE-ps2-CAT.

For the wild type ER in the absence of ligand (control vehicle only, lane labeled C), no interaction was observed with SRC-1, TIF-1, or ERAP-140, as expected (23, 26, 39). The addition of E_2 promoted a good interaction with these factors, whereas TOT showed no ability to promote interaction between the wild type ER and these factors. When GST protein alone was used, no interaction was observed with any of the coactivators, either in the absence or in the presence of ligands (data not shown). Since equal inputs of radiolabeled coactivators were used in all samples, it is of note that in the presence of E_2 , ERAP-140 and TIF-1 showed a much lower extent of interaction with wild type ER than SRC-1, which could mean that these cofactors are somewhat less specific for ER than SRC-1.

The Y537S receptor showed a distinct interaction with each of these three coactivators. In the absence of E_2 , this mutant interacted with SRC-1 as strongly as did wild type receptor in the presence of E_2 ; this interaction in the absence of E_2 was not significantly

enhanced by the addition of E_2 , but was completely suppressed by TOT. The Y537S ER showed a moderate but readily detected interaction with TIF-1 in the absence of E_2 , and again this interaction was suppressed by TOT. However, as opposed to the interaction of Y537S ER with SRC-1, which is already maximal in the absence of E_2 , the binding of TIF-1 to the Y537S ER was markedly enhanced by addition of E_2 . Y537S ER shows yet a third pattern of interaction with the coactivator ERAP-140. There is a small interaction in the absence of E_2 , which was easily seen with longer times of autoradiography (data not shown), whereas for the same exposure times, no interaction was observed with either the unliganded wild type ER or any of the other constitutively active receptors (Fig. 2). Moreover, as with TIF-1, the binding of ERAP-140 to the Y537S ER was greatly increased by addition of E_2 .

Therefore, the fully constitutively active mutant ER, Y537S, showed a different extent of ligand-independent interaction with the three coactivators: full inter-

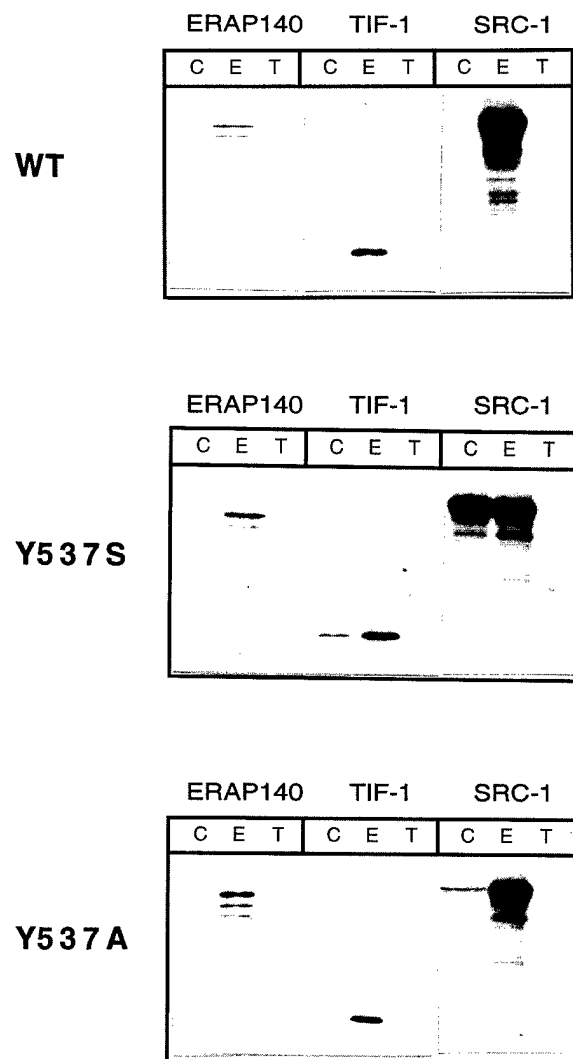


Fig. 2. Interaction of ERAP140, TIF-1, and SRC-1 Coactivators with Wild Type ER and with Constitutively Active ERs

GST fusion proteins with the wild type (WT) or Y537S, Y537A, E380Q ERs were incubated in the presence of the same amount of *in vitro* translated [³⁵S]methionine-labeled ERAP140, TIF-1, and SRC-1 coactivators. Incubations were performed in the absence of hormone (control 0.1% ethanol vehicle, C) or in the presence of E₂ (E: 1 μM) or TOT (T: 1 μM). After incubation and extensive washings of the glutathione Sepharose, the beads were boiled in Laemmli buffer, and samples were analyzed by SDS-PAGE followed by autoradiography.

action with SRC-1, some interaction with TIF-1, and little interaction with ERAP-140. Occupancy with E₂ was needed to achieve maximal interaction with TIF-1 and ERAP-140.

Concerning Y537A and E380Q mutant ERs, both receptors showed the same type of interaction with SRC-1, namely a weak interaction in the absence of E₂, which was greatly enhanced by addition of E₂. The interaction in the absence of ligand (control vehicle, C) was abolished by incubation in the presence of TOT. Both the Y537A and E380Q receptors required E₂ for

interaction with TIF-1 or ERAP-140, but even with E₂ treatment, the interaction of the E380Q mutant with ERAP-140 was much lower than that observed for E₂-occupied wild type receptor.

Mutant and Wild Type ERs Interact in a Ligand-Independent Manner with the PIC

The interaction of the ER with the PIC could be another step of the transcriptional process involved in the activation. Transcription factor II B (TFIIB) and TATA-box binding protein (TBP) are two members of the PIC that have been shown to be the target of numerous transcription factors. We therefore expressed these factors by *in vitro* translation and used them in pull-down experiments to test their interactions with our ER proteins. The wild type ER and the three mutants showed a much stronger interaction with TBP compared with TFIIB, but for both the mutant ERs and the wild type ER, interaction with TFIIB and with TBP occurred in the absence or presence of E₂ (Fig. 3). Among the three mutant receptors, the Y537S ER displayed the great-

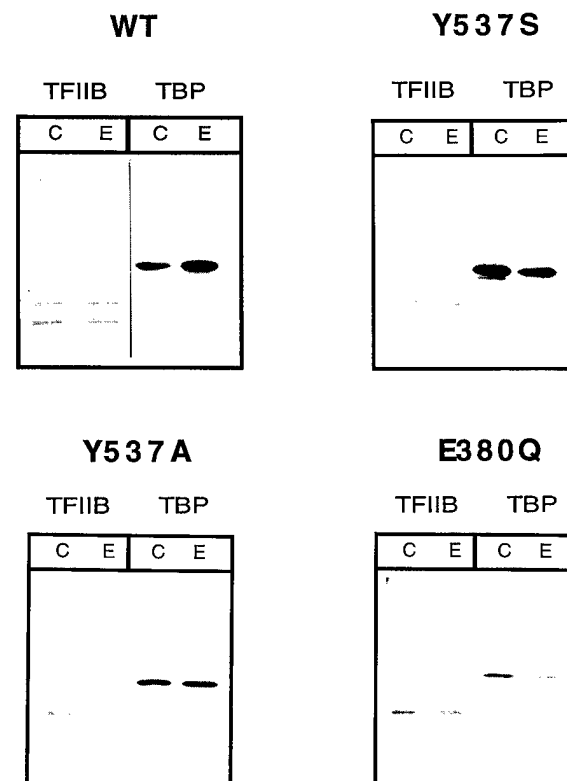


Fig. 3. Patterns of Interaction of Wild Type and Mutant ERs with Two Members of the PIC, TBP and TFIIB

Pull-down experiments were performed as described in Fig. 2 except that *in vitro* translated proteins were TBP and TFIIB in the absence or presence of E₂ (1 μM). *In vitro* translated TFIIB appeared as two major products, which both interacted with the wild type ER. The autoradiograms show the interactions of GST-WT, GST-Y537S, GST-Y537A, and GST-E380Q ERs with TBP and TFIIB.

est ability to interact with TBP, being comparable to that of wild type ER. There was less difference in the degree of interaction of all four receptors with TFIIB.

The Y537S Mutant Is in an Active Conformation in the Absence of Hormone: Proteolytic Digest Patterns

Previous studies have shown that differences in the conformation of unoccupied and hormone-occupied steroid receptors can be detected by differential sensitivity to protease digestion (30, 40). We therefore tested whether differences in proteolytic digestion patterns might provide a means to discriminate between constitutively active and inactive states among our mutant ERs. The proteolytic digestion patterns of [35 S]methionine-labeled wild type or

mutant ERs were analyzed by denaturing gel electrophoresis. The results of these analyses are shown in Fig. 4.

In the absence of ligand, ER is highly sensitive to trypsin and gives a proteolytic digestion pattern in which the fragment sizes decrease rapidly with increasing concentrations of trypsin, until two bands of approximately equal intensity appear and remain relatively stable (panel A, lanes 1–4). One of these is approximately 28 kDa (*arrow B*), while the other is approximately 25 kDa (*arrow A*). When the receptor is occupied with E_2 , only the upper band is strongly stabilized (lanes 5–8, *arrow B*), suggesting that receptor in an active conformation is protected from further cleavage by trypsin. A transiently stabilized fragment at about 35 kDa appears with E_2 treatment when receptor is digested with 5 μ g/ml trypsin (lane 6, *arrow*

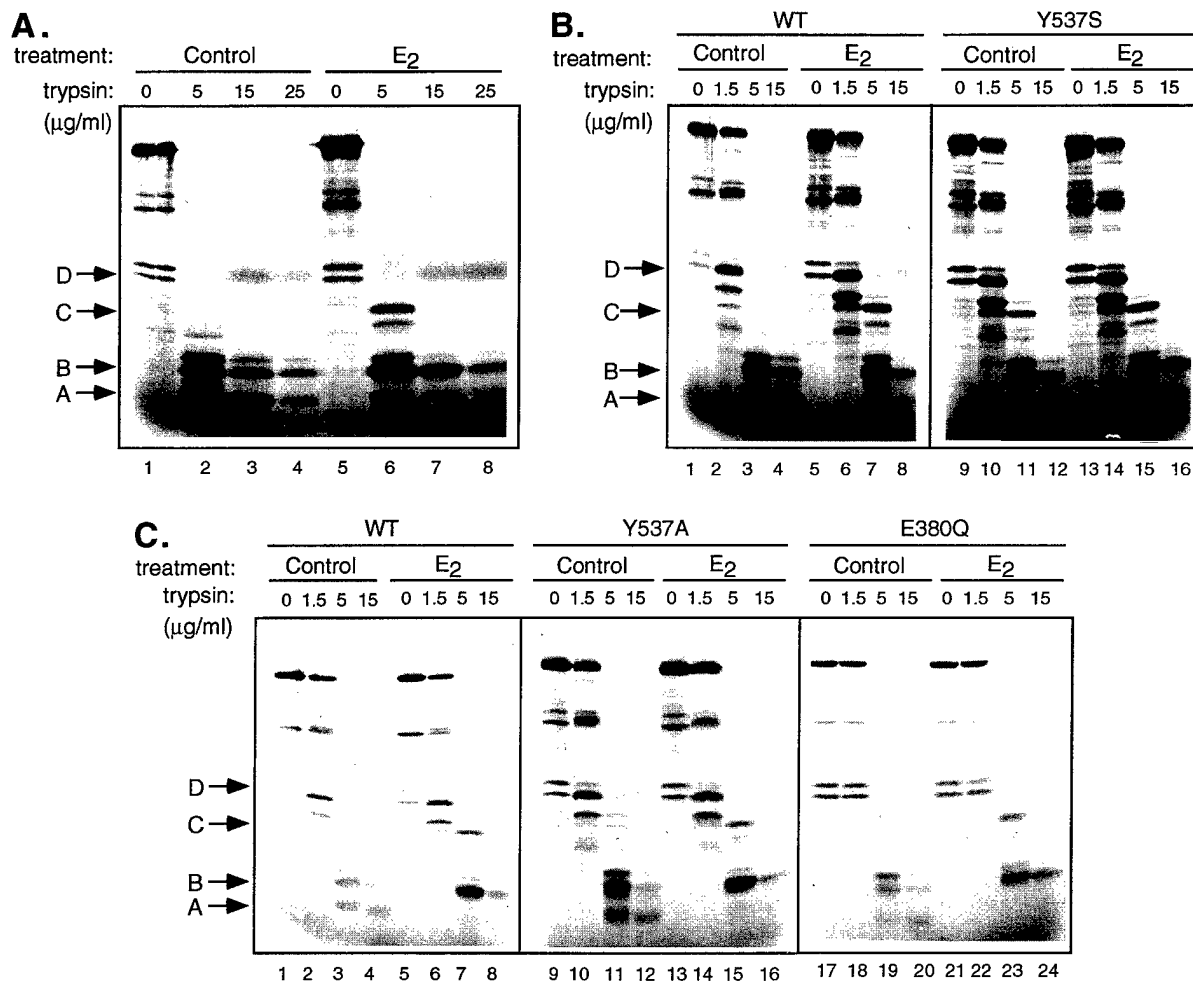


Fig. 4. Proteolytic Digest Patterns of Radiolabeled Wild Type or Constitutively Active ERs after Incubation with Increasing Concentrations of Trypsin

A, Unliganded (0.1% ethanol control vehicle, lanes 1–4) or E_2 -occupied (lanes 5–8) wild type ER incubated for 10 min at 22 C with 0, 5, 15, or 25 μ g/ml trypsin. B, Unliganded or E_2 -occupied wild type or Y537S ERs incubated for 10 min at 22 C with 0, 1.5, 5, or 15 μ g/ml trypsin. C, Unliganded or E_2 -occupied Y537A, wild type, or E380Q ERs incubated with trypsin as described for panel B. After trypsin exposure, samples were analyzed by SDS-PAGE. The radiolabeled products were visualized by autoradiography.

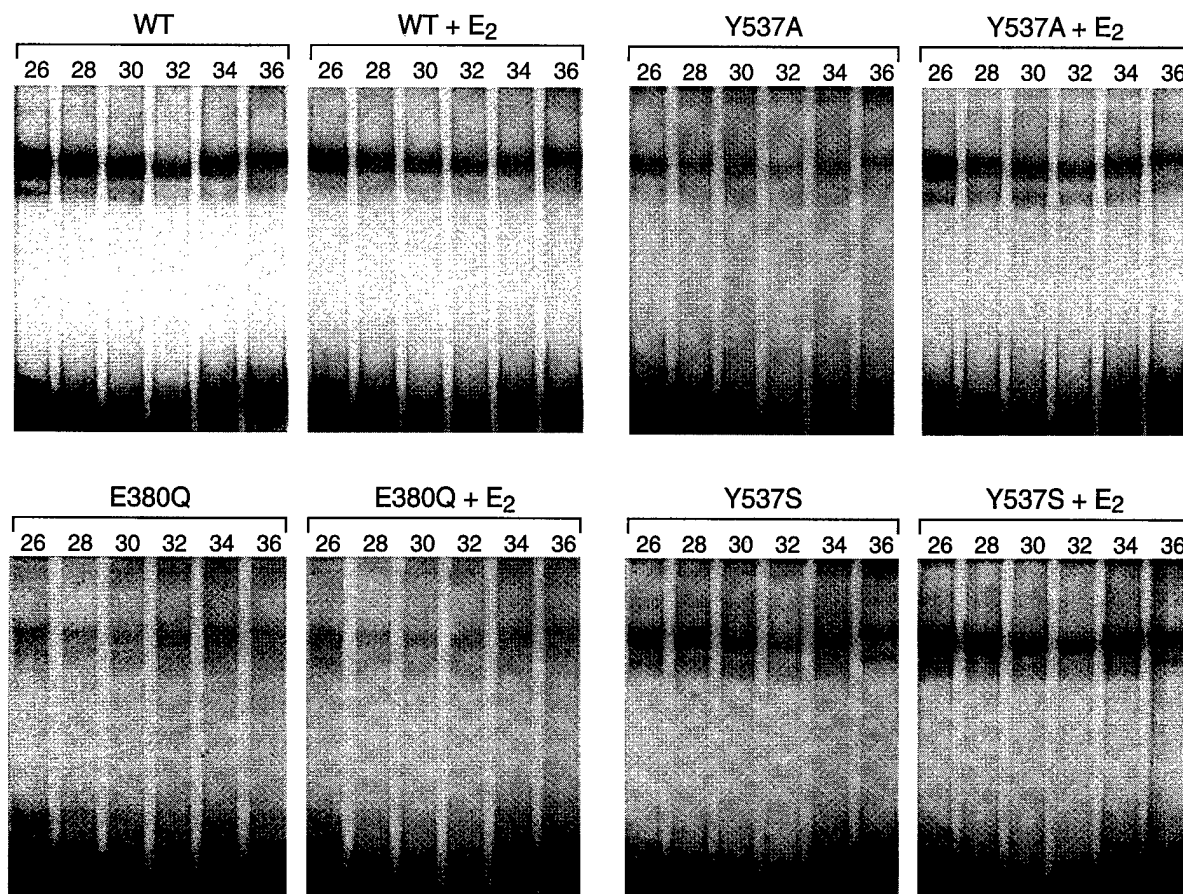


Fig. 5. Phasing Analysis of Wild Type and Mutant ERs with ERE-Containing DNA Fragments

Translation of receptors was performed in the absence or in the presence of $1 \mu\text{M}$ E_2 . Reticulocyte lysate-expressed receptors were incubated with ^{32}P -labeled DNA fragments containing an intrinsic bend separated from a consensus ERE by 26, 28, 30, 32, 34, or 36 nucleotides. The ER-DNA complexes were fractionated on a 8% polyacrylamide gel, dried, and visualized by autoradiography.

C), although this fragment is further digested when the trypsin concentration is increased.

The Y537S ER mutant, which shows nearly full constitutive activity in the absence of ligand, showed a proteolytic digestion pattern in both its unliganded or liganded state (compare lanes 9–12 with 13–16), which closely resembled the pattern seen for wild type ER treated with E_2 . Also, the transiently stabilized band at 35 kDa (arrow C), which in wild type ER is stabilized only in the presence of E_2 (lane 7), is strongly stabilized in Y537S ER without or with E_2 (lanes 11 and 15). There is, however, a small amount of protein present in the lower/inactive form (arrow A) in the absence of hormone (lanes 11 and 12), which shifts to the upper/active form (arrow B) when E_2 is added. Thus, the proteolytic digest pattern for the Y537S receptor reflects its constitutively active state.

The unoccupied E380Q and Y537A ERs, which possess partial constitutive activity, showed a slight enhancement of the transiently stabilized, high molecular mass bands, including the approximately 40-kDa band D and the 35-kDa band C, relative to the unoccupied wild type ER, especially noticeable at a trypsin

concentration of $5 \mu\text{g/ml}$ (compare lanes 11 and 19 vs. 3). Perhaps these subtle differences are indicative that the unliganded E380Q and Y537A receptors have taken on a partially active conformation. However, E_2 treatment resulted in a predominance of the B species after increasing trypsin treatment, as observed with the wild type E_2 -occupied receptor.

Transcriptionally Active Wild Type and Mutant ERs Induce Similar DNA-Bending Angles

Wild type and mutant ERs were expressed in a reticulocyte lysate system in the presence or absence of estrogen and used in DNA-phasing analysis experiments to determine the orientation and magnitude of ER-induced DNA bending (Fig. 5). The unoccupied and estrogen-occupied *in vitro* translated ERs were incubated with ^{32}P -labeled DNA fragments, each of which contained an intrinsic DNA bend separated from a consensus ERE by either 26, 28, 30, 32, 34, or 36 bp (41). Thus, when the ER bound to the ERE and induced DNA bending, the intrinsic and ER-induced DNA bends would either be in phase and form a larger DNA

bend or be out of phase and have the effect of straightening the DNA fragment. When wild type and mutant ERs were incubated with DNA fragments containing 26 or 36 bp between the intrinsic and ER-induced DNA bends, there was an increase in the mobilities of the receptor-DNA complexes through the acrylamide gel, indicating that the ER-induced and intrinsic DNA bends were in phase. When the wild type and mutant ERs were incubated with DNA fragments containing 32 bp between the intrinsic and ER-induced DNA bends, there was an increase in the mobilities of the receptor-DNA complexes indicating that the ER induced and intrinsic DNA bends were out of phase. Because the intrinsic and ER-induced DNA bends were on the same side of the DNA helix when they were out of phase (assuming 10.5 bp/helical turn), these findings do demonstrate that the ER-induced DNA bend opposes the intrinsic DNA bend, which is directed toward the minor groove of the DNA helix (42). Therefore, the unoccupied and estrogen-occupied wild type and mutant ERs induced DNA bends that were directed toward the major groove of the DNA helix. The magnitudes of the ER-induced DNA bends were determined from replicate phasing analysis experiments (see *Materials and Methods*) and are summarized in Table 1. The unoccupied wild type ER induced a DNA bend of 15.6°, the largest bending angle measured. In contrast, the estrogen-occupied wild type ER induced a much smaller bend of 7.3°. Interestingly, the three mutant receptors, Y537S, Y537A, and E380Q, which are active in the presence and in the absence of hormone, had statistically similar directed DNA-bending angles of approximately 7.5°–9°, in the absence or presence of hormone. Thus, the mutant receptors induced directed bending angles that were similar in magnitude to the bend induced by the estrogen-occupied ER. These findings are in good agreement with previous studies carried out with estrogen-occupied wild type and mutant ERs that had been expressed in COS cells (43) and support the idea that transcriptionally active wild type and mutant ERs induce directed bending angles of similar magnitude.

Table 1. Directed DNA Bending Angles Induced by Wild Type and Mutant ERs

Receptor	Directed Bending Angle (α_B) (for unoccupied receptor)	Directed Bending Angle (α_B) (for E ₂ -occupied receptor)
Wild type ER	15.6 \pm 2.7 (5)	7.3 \pm 1.6 (4)
Y537S ER	8.0 \pm 0.8 (3)	9.0 \pm 0.7 (3)
Y537A ER	8.3 \pm 2.1 (3)	9.2 \pm 1.0 (7)
E380Q ER	7.2 \pm 0.4 (3)	9.3 \pm 0.5 (4)

Directed bend angles (α_B) induced by ER binding to ERE-containing DNA phasing fragments observed for receptors translated in the absence (control) or presence of E₂. Values are reported as the mean \pm SEM. The number of independent determinations is indicated in parentheses.

DISCUSSION

Our findings show that the active state of the ER is characterized by several features including functional and conformational/structural properties that are distinct from the ones observed when the receptor is in an inactive state. Interestingly, some of these features are common for the liganded wild type receptor and for constitutively active receptors in the absence or presence of hormone, suggesting that the mechanisms underlying their biological activity are similar.

Figure 6 presents a model summarizing our findings regarding the interaction of several coregulators and basal transcription factors with the wild type and constitutively active Y537S ER and the effects of E₂ on these interactions and on DNA bending by these receptors. The figure emphasizes that occupancy of the wild type ER by E₂ elicits a reduction in the extent of DNA bending and an increase in the association with coactivators. The Y537S ER associates strongly with some, but not all, coactivators in the ligand-unoccupied state, and it exhibits the same DNA bend angle in its ligand-free or E₂-occupied state as does the E₂-occupied wild type receptor.

The most constitutively active ER (Y537S) had E₂-occupied ER-like character in all assays but two, and in those (association with TIF-1 and ERAP-140) it showed some wild type-liganded ER character (Table 2). The partially constitutively active receptors showed liganded ER character in some assays but not others. Of the three ER coregulators evaluated, SRC-1 interaction correlated best with the degree of transcriptional activity displayed by the mutant ERs.

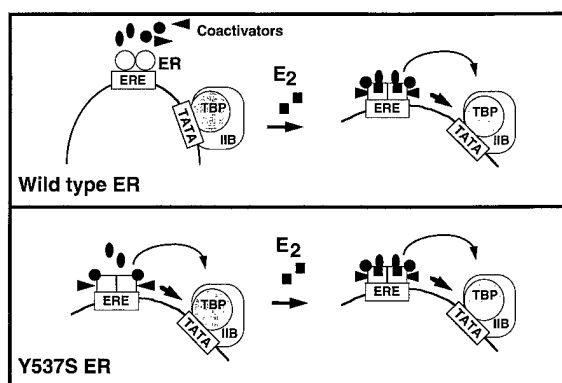


Fig. 6. Model Summarizing Our Findings Regarding the Interaction of Several Coregulators and Basal Transcription Factors with the Wild Type and Constitutively Active Y537S ER and the Effects of E₂ on These Interactions and on DNA Bending by These Receptors

The figure emphasizes that occupancy of the wild type ER by E₂ elicits a reduction in the extent of DNA bending and an increase in the association with coactivators. The Y537S ER associates strongly with some but not all coactivators in the ligand-unoccupied state, and it exhibits the same DNA bend angle in its ligand-free or E₂-occupied state as does the E₂-occupied wild type receptor.

Table 2. Parameters Assessing the Transcriptionally Active State of Wild Type and Constitutively Active Mutant ERs

Receptor	Transcription		Coactivators						Basal Factors				Protease Digestion Pattern		DNA Bend Angle	
			SRC1		TIF1		ERAP140		TBP		TFIIB					
	−E ₂	+E ₂	−E ₂	+E ₂	−E ₂	+E ₂	−E ₂	+E ₂	−E ₂	+E ₂	−E ₂	+E ₂	−E ₂	+E ₂	−E ₂	+E ₂
Wild type	—	● ^a	—	●	—	●	—	●	○	●	○	●	—	●	○	●
Y537S	○	●	○	●	○	●	○	●	○	●	○	●	○	●	○	●
Y537A	○	●	○	●	○	●	○	●	○	●	○	●	○	●	○	●
E380Q	○	●	○	●	○	●	—	●	○	●	○	●	○	●	○	●

^a Open circles indicate ligand-unoccupied receptor. Shaded circles indicate estrogen (E₂)-occupied receptor. Size of the circle, whether open or shaded, denotes magnitude of response, the larger the circle the greater the response. Shaded large circles represent the magnitude of the response obtained with the E₂-occupied wild type ER.

Of the various endpoints assessed, DNA bending was the most sensitive to the constitutively active character of the receptor (see Table 2). The receptor proteolytic digestion pattern and receptor coregulator association profiles only appear to clearly indicate the "active" character of the most constitutively active mutant ER Y537S; the other two receptors showed subtle differences in these assays consistent with some change toward the active state observed with the E₂-liganded wild type ER, but overall the E380Q and Y537A receptors resembled wild type ER. Although DNA bending did not distinguish between partially and fully constitutively active receptors, it appears to be very sensitive to the propensity of the receptor to be in the active conformation. By contrast, the protease digestion pattern differences may require that receptor be very strongly in the active conformation, as the hormone-occupied wild type-like ER pattern was only observed with the most constitutively active ER, Y537S. The fact that both partially active and fully constitutively active receptors gave the same directed DNA-bending angle suggests that the DNA may form a scaffold for the accumulation of other protein factors and coregulators important in determining receptor transcriptional activity, with DNA bending being the first of several important steps leading to full receptor transcriptional effectiveness. Interaction of the ER with basal factors did not require ligand, and it is therefore not surprising that interaction with TFIIB and TBP does not discriminate wild type from constitutively active ERs.

The wild type ER was able to interact with the three coactivators tested (ERAP140, TIF-1, and SRC-1) in the presence of E₂, but not in the presence of antiestrogens such as TOT. The strongest interaction was observed with SRC-1 and TIF-1 and to a lesser extent with ERAP140. In the presence of E₂, the three constitutive mutants displayed the same ability to interact with these coactivators as the wild type ER. However, in the absence of E₂, these mutants exhibited different abilities to interact with the coactivators. Indeed, the Y537S mutant was able to interact with SRC-1 in the absence of E₂ to the same extent as the wild type receptor in the presence of E₂. Moreover, this interaction was not enhanced by addition of E₂.

With the two other coactivators, the interaction with Y537S ER was still markedly enhanced by (TIF-1), or dependent on, ligand (ERAP 140). Concerning E380Q and Y537A mutants, they showed basically a common pattern of interaction with the coactivators: no interaction in the absence of E₂ with ERAP140 or TIF-1 and only a weak interaction with SRC-1 in the absence of ligand. Our observations suggest that SRC-1, TIF-1, and ERAP-140 play somewhat different roles in ER activity and may be involved to different degrees in the process of receptor-regulated transcription. That the activity of the mutants correlated well with binding to SRC-1 implies that SRC-1 is likely a functional mediator of ER transcriptional activity. The interaction of the three mutant receptors with coactivator in the absence of ligand was abolished by addition of *trans*-hydroxytamoxifen, suggesting that antiestrogen induces a different conformation of the receptor that is not compatible with the interaction with coactivator. Our data are in agreement with the model proposed by Chen and Evans (13), *i.e.* that the receptor bound to DNA could be in three states: in the absence of hormone, the receptor is inactive (or might be repressive as in the case of TR and RAR, which interact with corepressors). Once the ligand is added, the receptor is in an intermediary state, and basal transcription can take place. And finally, the liganded receptor is able to interact with coactivators, which would be a link with the PIC, and to activate strongly the transcription process.

The mechanism by which the nuclear receptors activate transcription remains unclear, but it is proposed that the receptors could stimulate PIC formation either by recruiting the different members of the PIC or by positioning a preformed PIC on the DNA. The PIC consists of at least seven basal transcription factors, namely TFIIA, TFIIB, TFIID [comprised of the TBP interacting with TBP-associated factors, TFIIE, TFIIIF, TFIIH, and TFIIJ (for review, see Ref. 44)]. A number of transcription factors, including nuclear receptors and, in particular, the ER, have been shown to interact *in vitro* with TFIID (45–48) or TFIIB (49–53). We have now investigated the interaction of the wild type and mutant ERs with members of the PIC. Basically, all the ERs tested, whether wild type or constitutively active,

interacted with TBP and TFIIB in the absence or presence of hormone. However, of note was the fact that TBP interaction, which was substantial in the absence of estrogen but was enhanced by E_2 in the case of wild type receptor, was not increased by E_2 for mutant receptors. These data suggest that the ability of a receptor to interact with the PIC is not a specific feature of activated receptors. However, it is possible that the interaction of inactive receptors (*i.e.* unoccupied wild type ER) with the PIC could be nonproductive. The transformation of the receptor into an active state would then not lead to an increase of the interactions but rather to a modification of the nature of these interactions.

To further investigate the conformational changes that could arise from treatment with estrogen, we performed limited proteolytic digestion experiments. Using the wild type receptor, we present evidence that the conformation of the receptor in the absence and in the presence of E_2 are different, which is in agreement with previous work (30, 54). Indeed, in the absence of ligand, the receptor was highly sensitive to trypsin and gives a proteolytic digestion pattern in which, for high concentrations of trypsin, two stable bands (~25 and 28 kDa) of approximately equal intensity appeared. Treatment with E_2 stabilized strongly the upper, 28-kDa band. The transformation of the receptor into a distinct, active conformation upon hormone exposure has been reported not only for ER but also for progesterone receptor and RAR/RXR (30, 31), suggesting that this is a phenomenon common to the ligand-activatable nuclear receptors. Of note, the fully constitutively active Y537S ER exhibited a proteolytic digestion pattern in the absence of E_2 , which was very similar to the pattern of the active wild type estrogen-occupied ER, suggesting that this mutant receptor was in the conformationally active state.

If the receptor can undergo conformational changes after activation, it is possible that these changes could modify the nature of its interactions with DNA. It has been reported by several groups that the binding of transcription factors to DNA can induce bending of DNA (34–36, 41, 43, 55–57). A change in the bending state of the DNA could reflect a difference in the ability of the receptor to activate transcription. Our results show that the unliganded wild type receptor induced a larger bend (~16°) and that the E_2 -occupied wild type receptor induced a dramatically smaller DNA bend (~7°). Of particular interest is the fact that the three constitutively active unoccupied and estrogen-occupied ER mutants induced DNA bends that were similar to the bending angle induced by the E_2 -occupied wild type receptor. This smaller DNA bend would be a characteristic of the active state. We propose that the distinct conformations induced by active and inactive receptors could be interpreted as activating and silencing signals. We should emphasize that the difference in DNA bending observed for the active and inactive wild type ER could be obtained using a totally

cell-free system, suggesting that cell context is not a factor in the effect of receptor on DNA bending.

Altogether, the data from studies with these constitutively active ERs provide evidence that the transformation of receptor into an active state involves a complex set of events that include conformational changes as well as distinct alterations in the ability of the receptor to interact with coregulator proteins and with DNA. Collectively, these changes in conformation and interactions "mark" the receptor as being transcriptionally productive.

MATERIALS AND METHODS

Chemicals

E_2 was from Sigma (St. Louis, MO), TOT and ICI 164,384 were kindly provided by Dr. Alan Wakeling (Zeneca Pharmaceuticals, Macclesfield, U.K.). [35 S]Methionine was from ICN (Costa Mesa, CA).

Plasmid Construction

The plasmid encoding SRC-1 (26) was kindly provided by Drs. Ming Tsai and Bert O'Malley. The plasmid encoding TIF-1 (amino acids 434–750) (39) was a gift from Dr. Pierre Chambon. The plasmid pGEX-2TK-ER, which contains the human ER spanning amino acids 282–595, and the plasmid encoding ERAP140 (23) were kindly provided by Dr. Myles Brown. The two expression vectors encoding TBP and TFIIB were kindly provided by Dr. Danny Reinberg. The GST-ER mutant plasmids for Y537A and Y537S were previously described (33). PCR was used to generate the E380Q ER fragment from amino acids 282 to 595 bearing *Bam*HI and *Eco*RI sites and cloned into PGEX-2TK.

Production of GST Fusion Proteins

Bacteria expressing GST fusion proteins were grown at 37°C in 500 ml of LB (Luria Bertani) broth until the absorbance (600 nm) reached 0.8. Then the induction was performed for 3 h at 30°C with 1 mM isopropyl β -D-thiogalactopyranoside (IPTG). Cells were collected by centrifugation at 4°C at 3000 $\times g$ for 15 min. The supernatant was discarded, and the pellet was rapidly frozen in liquid nitrogen and then kept for 1 h at –80°C. Frozen pellets were then thawed on ice and resuspended in 0.01 volume of NET buffer (20 mM Tris, pH 8.0/100 mM NaCl/1 mM EDTA) and sonicated twice for 30 sec at maximum level. The suspension was centrifuged for 10 min at 12,000 $\times g$, and the supernatant was then transferred to other tubes and centrifuged at 105,000 $\times g$ (30 min, 4°C). Protein concentration was estimated by the Bradford method. The levels of expressed fusion proteins were determined by *in vitro* binding assays followed by Western analysis with H222 monoclonal antibody.

In Vitro Translation of Receptor-Associated Proteins and Human ER Proteins

In vitro translation was performed using the TNT Promega kit (Promega, Madison, WI). Briefly, 1 μ g of expression vector was mixed with 25 μ l TNT rabbit reticulocyte lysate, 2 μ l TNT buffer, 1 μ l of mix containing all amino acids except methionine, 1 μ l RNasin (50 U/ μ l), 1 μ l T3 RNA polymerase (20 U/ μ l), and 4 μ l of [35 S]methionine (15 μ Ci/ μ l). The final reac-

tion volume was 50 μ l. The reaction was performed for 1.5 h at 30 C. The translation efficiency was checked by loading 1 μ l of lysate on an SDS-PAGE gel.

For gel mobility shift assays, the translation was performed in the presence of control vehicle (0.1% ethanol) or 1 μ M E_2 as above, except that labeled methionine was replaced with unlabeled methionine.

In Vitro Binding Assays with Glutathione Sepharose

Glutathione Sepharose (Pharmacia, Piscataway, NJ) was equilibrated with IP binding buffer (25 mM Tris-HCl (pH 7.9), 10% vol/vol glycerol, 0.1% NP-40, 0.5 mM dithiothreitol, 100 mM KCl). The *in vitro* translated products were first precleared for 2 h by incubation with 100 μ l of beads and 300 μ g of GST (which does not contain any insert). Crude bacterial extract (500 μ g) containing GST fusion proteins was incubated at 4 C with 25 μ l of beads for 2.5 h in the presence of vehicle (0.1% ethanol) or hormone (E_2 or TOT, at 1 μ M concentration). After three washes, the beads were incubated with 5 μ l of *in vitro* translated product for 2.5 h in the presence of vehicle or hormone at 4 C. The beads were washed three times with 1 ml of IP buffer and two times with 1 ml of IP buffer containing 300 mM KCl. After washing, beads were boiled in SDS sample buffer, and a quarter of the proteins were run on SDS-PAGE. The gel was fixed, dried, and submitted to autoradiography.

Protease Digestion Assays

Bluescript vector (Stratagene, La Jolla, Ca) was used for insertion of cDNA sequences of wild type ER, E380Q, Y537A, and Y537S ER mutants. Aliquots of *in vitro* translated, [³⁵S]-labeled proteins (25 μ l) were treated with control (0.1% EtOH) vehicle or ligand at a final concentration of 9×10^{-6} M for 20 min at room temperature. Aliquots (5 μ l) of the ligand-treated receptor were incubated without trypsin or with trypsin to a final concentration of 1.5, 5, 15, or 25 μ g/ml (Worthington Biochemicals, Freehold, NJ) After a 10-min incubation at room temperature, the digestion was stopped with 20 μ l of Laemmli buffer, and the samples were boiled for 5 min and then separated on a 12% SDS-PAGE gel. The radiolabeled products were visualized by autoradiography.

DNA-Bending Gel Mobility Shift Assays

The ERE-containing DNA-phasing vectors, ERE 26, ERE 28, ERE 30, ERE 32, ERE 34, and ERE 36 (41), were digested with *Eco*RI and *Hind*III, isolated on an acrylamide gel, and electrophoresed. The 281- to 291-bp DNA fragments containing the intrinsic DNA bend and the ERE were filled in with Klenow in the presence of [³²P]dATP and [³²P]dGTP and then purified using a G-25 Sephadex Quick Spin column (Boehringer Mannheim, Indianapolis, IN). Gel mobility shift assays were carried out as previously described (55) with minor modifications. Briefly, 10,000 cpm of the ³²P-labeled DNA phasing fragment was combined with 4 μ l (188 μ g total protein) reticulocyte lysate-expressed wild type or mutant ER and 1 μ g poly(deoxyinosinic-deoxycytidylic)acid (Sigma) in a buffer containing 10% glycerol, 50 mM KCl, 15 mM Tris, pH 7.9, 0.2 mM EDTA, and 0.4 mM dithiothreitol (20 μ l final volume) for 15 min at room temperature. Low ionic strength gels and buffers were prepared as described (58). Twenty-centimeter gels were prerun for 1 h at 300 V. Samples were fractionated for 3 h on an 8% (75:1 acrylamide to bis-acrylamide ratio) polyacrylamide gel. Water recirculation was used to maintain the gels at 4 C. Radioactive bands were visualized by autoradiography. The relative mobilities of the ER-DNA complexes and free probes were quantitated with a Molecular Dynamics PhosphorImager and Imagequant software (Molecular Dynamics, Sunnyvale, CA). The magnitudes of the receptor-induced directed DNA bending angles (α_B) were determined

for the wild type and mutant ERs using the empirical formula (59):

$$\tan(k\alpha_B/2) = \frac{A_{PH}/2}{\tan(k\alpha_C/2)}$$

where α_C is the intrinsic DNA bending angle, A_{PH} is the phasing amplitude, and k is a coefficient used to adjust for electrophoretic conditions. A value of $k = 0.991$ was determined by comparing the relative mobility of five sets of DNA-bending standards with their known bending angles (60).

Acknowledgments

We thank Myles Brown, Danny Reinberg, Pierre Chambon, Ming-Jer Tsai, and Bert O'Malley for providing plasmids.

Received April 2, 1997. Revision received and accepted May 21, 1997.

Address requests for reprints to: Dr. Benita S. Katzenellenbogen, Department of Molecular and Integrative Physiology, University of Illinois at Urbana-Champaign, 524 Burrill Hall, 407 South Goodwin Avenue, Urbana, Illinois 61801-3704.

This research was supported by NIH Grant R37 CA-18119 (to B.S.K.), a Susan G. Komen Foundation Postdoctoral Fellowship (to G.L.), and NIH Grant R29 HD-31299 (to A.M.N.).

REFERENCES

1. Evans RM 1988 The steroid and thyroid hormone receptor superfamily. *Science* 240:889-895
2. Krust A, Green S, Argos P, Kumar V, Walter P, Bornert JM, Chambon P 1986 The chicken oestrogen receptor sequence: homology with V-erbA and the human oestrogen receptor and glucocorticoid receptors. *EMBO J* 5:891-897
3. Metzger D, Ali S, Bornert JM, Chambon P 1995 Characterization of the amino-terminal transcriptional activation function of the human estrogen receptor in animal and yeast cell. *J Biol Chem* 270:9535-9542
4. Tora L, Gronemeyer H, Turcotte B, Gaub M-P, Chambon P 1988 The N-terminal region of the chicken progesterone receptor specifies target gene activation. *Nature* 333:185-188
5. Danielian PS, White R, Lees JA, Parker MG 1992 Identification of a conserved region required for hormone dependent transcriptional activation by steroid hormone receptors. *EMBO J* 11:1025-1033
6. Hård T, Kellenbach E, Boelens R, Maler BA, Dahlman K, Freedman LP, Carlstedt-Duke J, Yamamoto KR, Gustafsson JA, Kaptein R 1990 Solution structure of the glucocorticoid receptor DNA-binding domain. *Science* 215:157-160
7. Mader S, Chambon P, White JH 1993 Defining a minimal estrogen receptor DNA binding domain. *Nucleic Acids Res* 21:1125-1132
8. Kumar V, Green S, Staub A, Chambon P 1986 Localization of the oestradiol-binding and putative DNA binding domains of the human oestrogen receptor. *EMBO J* 5:2231-2236
9. Bresnick EH, Dalman FC, Sanchez ER, Pratt WB 1989 Evidence that the 90-kDa heat shock protein is necessary for the steroid binding conformation of the L cell glucocorticoid receptor. *J Biol Chem* 264:4992-4997
10. Miller MA, Mullick A, Greene GL, Katzenellenbogen BS 1985 Characterization of the subunit nature of nuclear estrogen receptors by chemical crosslinking and dense

- amino acid labeling. *Endocrinology* 117:515-522
11. Smith DF, Faber LE, Toft DO 1990 Purification of inactivated progesterone receptor and identification of novel receptor associated proteins. *J Biol Chem* 265:39996-40003
 12. Burris TP, Nawaz Z, Tsai M-J, O'Malley BW 1995 A nuclear hormone receptor-associated protein that inhibits transactivation by the thyroid hormone and retinoic acid receptors. *Proc Natl Acad Sci USA* 92:9525-9529
 13. Chen JD, Evans RM 1995 A transcriptional co-repressor that interacts with nuclear hormone receptors. *Nature* 377:454-457
 14. Chen JD, Umesono K, Evans RM 1996 SMRT isoforms mediate repression and anti-repression of nuclear receptor heterodimers. *Proc Natl Acad Sci USA* 93:7567-7571
 15. Hörlein AJ, Näär AM, Heinzel T, Torchia J, Glass B, Kurokawa R, Ryan A, Kamel Y, Söderström M, Glass CK, Rosenfeld MG 1995 Ligand-independent repression by the thyroid hormone receptor mediated by a nuclear receptor co-repressor. *Nature* 377:397-404
 16. Kurokawa R, Söderström M, Hörlein A, Halachmi S, Brown M, Rosenfeld MG, Glass CK 1995 Polarity-specific activities of retinoic acid receptors determined by a co-repressor. *Nature* 377:451-454
 17. Yen PM, Wilcox EC, Hayashi Y, Refetoff S, Chin WW 1995 Studies on the repression of basal transcription (silencing) by artificial and natural human thyroid hormone receptor- β -mutants. *Endocrinology* 136:2845-2851
 18. Nemoto T, Ohara-Nemoto Y, Denis M, Gustafsson JA 1990 The transformed glucocorticoid receptor has a lower steroid-binding affinity than the non-transformed receptor. *Biochemistry* 29:1880-1886
 19. Sanchez ER, Faver LE, Henzel WJ, Pratt WB 1990 The 56-59-kilodalton protein identified in untransformed receptor complexes is a unique protein that exists in cytosol in a complex with both the 70- and 90-kilodalton heat shock proteins. *Biochemistry* 29:5145-5152
 20. Baur EV, Zechel C, Heery D, Heine M, Garnier JM, Vivat V, Ledouarin B, Gronemeyer H, Chambon P, Losson R 1996 Differential ligand-dependent interactions between the AF-2 activating domain of nuclear receptors and the putative transcriptional intermediary factors msug1 and TIF1. *EMBO J* 15:110-124
 21. Cavaillès V, Dauvois S, Danielian PS, Parker MG 1994 Interaction of proteins with transcriptionally active estrogen receptors. *Proc Natl Acad Sci USA* 91:10,009-10,013
 22. Cavaillès V, Dauvois S, L'Horsset F, Lopez G, Hoare S, Kushner PJ, Parker MG 1995 Nuclear factor RIP140 modulates transcriptional activation by the estrogen Receptor. *EMBO J* 14:3741-3751
 23. Halachmi S, Marden E, Martin G, MacKay H, Abbonanza C, Brown M 1994 Estrogen receptor-associated proteins: possible mediators of hormone-induced transcription. *Science* 264:1455-1458
 24. Hong H, Kohli K, Trivedi A, Johnson DL, Stallcup MR 1996 GRIP1, a novel mouse protein that serves as a transcriptional coactivator in yeast for the hormone binding domains of steroids receptors. *Proc Natl Acad Sci USA* 93:4948-4952
 25. Kamei Y, Xu L, Heinzel T, Torchia J, Kurokawa R, Glass B, Lin SC, AHR, Rose DW, Glass CK, Rosenfeld MG 1996 A CBP integrator complex mediates transcriptional activation and AP-1 inhibition by nuclear receptors. *Cell* 85:403-414
 26. Onate SA, Tsai SY, Tsai MJ, O'Malley BW 1995 Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270:1354-1357
 27. Voegel JJ, Heine M, Zechel C, Chambon P, Gronemeyer H 1996 TIF2, a 160 kDa transcriptional mediator for the ligand-dependent activation function AF-2 of nuclear receptors. *EMBO J* 15:3667-3675
 28. Ali S, Metzger D, Bornert JM, Chambon P 1993 Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J* 12:1153-1160
 29. Orti E, Bodwell JE, Munck A 1992 Phosphorylation of steroid hormone receptors. *Endocr Rev* 13:105-128
 30. Allan GF, Leng X, Tsai SY, Weigel NL, Edwards DP, Tsai MJ, O'Malley BW 1992 Hormone and anti-hormone induce distinct conformational changes which are central to steroid receptor activation. *J Biol Chem* 267:19513-19520
 31. Valcarcel R, Holz H, Garcia Jeménez C, Baretino D, Stunnenberg HG 1994 Retinoid-dependent *in vitro* transcription mediated by the RXR/RAR heterodimer. *Genes Dev* 8:3068-3079
 32. Pakdel F, Reese JC, Katzenellenbogen BS 1993 Identification of charged residues in an N-terminal portion of the hormone binding domain of the human estrogen receptor important in transcriptional activity of the receptor. *Mol Endocrinol* 7:1408-1417
 33. Weis KE, Ekena K, Thomas JA, Lazennec G, Katzenellenbogen BS 1996 Constitutively active human estrogen receptors containing amino acid substitutions for tyrosine 537 in the receptor protein. *Mol Endocrinol* 10:1388-1398
 34. Frankel AD, Kim PS 1991 Modular structure of transcription factors: implication for gene regulation. *Cell* 65:717-719
 35. Horikoshi M, Bertuccioli C, Takada R, Wang J, Yamamoto T, Roeder RG 1992 Transcription factor TFIID induces DNA bending upon binding to the TATA element. *Proc Natl Acad Sci USA* 89:1060-1064
 36. Nardulli AM, Greene GL, Shapiro DJ 1993 Human estrogen receptor bound to an estrogen response element bends DNA. *Mol Endocrinol* 7:331-340
 37. Renaud JP, Rochel N, Ruff M, Vivat V, Chambon P, Gronemeyer H, Moras D 1995 Crystal structure of the RAR-g ligand-binding domain bound to all-trans retinoic acid. *Nature* 378:681-689
 38. Wagner RL, Apriletti JW, McGrath ME, West BL, Baxter JD, Fletterick RJ 1995 A structural role for hormone in the thyroid hormone receptor. *Nature* 378:690-697
 39. Le Douarin B, Zechel C, Garnier JM, Lutz Y, Tora L, Pierrat B, Heery D, Gronemeyer H, Chambon P, Losson R 1995 The N-terminal part of TIF-1, a putative mediator of the ligand-dependent activation function (AF-2) of nuclear receptors, is fused to B-RAF in the oncogenic protein T18. *EMBO J* 14:2020-2033
 40. McDonnell DP 1995 Unraveling the human progesterone receptor signal transduction pathway. *Trends Endocrinol Metab* 6:133-138
 41. Nardulli AM, Grobner C, Cotter D 1995 Estrogen receptor-induced DNA bending: orientation of the bend and replacement of an estrogen response element with an intrinsic DNA bending sequence. *Mol Endocrinol* 9:1064-1076
 42. Zinkel SS, Crothers DM 1987 DNA bend direction by phase sensitive detection. *Nature* 328:178-181
 43. Potthoff SJ, Romine LR, Nardulli AM 1996 Effects of wild type and mutant estrogen receptors on DNA flexibility, DNA bending and transcription activation. *Mol Endocrinol* 10:1095-1106
 44. Roeder RG 1991 The complexities of eukaryotic transcription initiation: regulation of preinitiation complex assembly. *Trends Biochem Sci* 16:402-408
 45. Sadovskiy Y, Webb P, Lopez G, Baxter JD, Fitzpatrick PM, Gizang-Ginsberg E, Cavaillès V, Parker MG, Kushner PJ 1995 Transcriptional activators differ in their responses to overexpression of TATA-box-binding protein. *Mol Cell Biol* 15:1554-1563
 46. Brou C, Wu J, Ali S, Scheer E, Lang C, Davidson I, Chambon P, Tora L 1993 Different TBP-associated factors are required for mediating the stimulation of tran-

- scription *in vitro* by the acidic transactivator GAL-VP16 and the two nonacidic activation functions of the estrogen receptor. *Nucleic Acids Res* 21:5-12
47. Ham J, Steger G, Yaniv M 1994 Cooperativity *in vivo* between the E₂ transactivator and the TATA box binding protein depends on core promoter structure. *EMBO J* 13:147-157
 48. Schulman IG, Chakravarti D, Juguilon H, Romo A, Evans RM 1995 Interactions between the retinoid X receptor and a conserved region of the TATA-binding protein mediate hormone-dependent transactivation. *Proc Natl Acad Sci USA* 92:8288-8292
 49. Blanco JCG, Wang IM, Tsai SY, Tsai MJ, O'Malley BW, Jurutka PW, Haussler MR, Ozato K 1995 Transcription factor TFIIB and the vitamin D receptor cooperatively activate ligand-dependent transcription. *Proc Natl Acad Sci USA* 92:1535-1539
 50. Ing NH, Beekman JM, Tsai SY, O'Malley BW 1992 Members of the steroid receptor superfamily interact with TFIIB(S300-II). *J Biol Chem* 267:17617-17623
 51. Lin YS, Green MR 1991 Mechanism of action of an acidic transcriptional activator *in vitro*. *Cell* 64:971-981
 52. Malik S, Karathanasis S 1995 Transcriptional activation by the orphan nuclear receptor ARP-1. *Nucleic Acids Res* 23:1536-1543
 53. McDonald PN, Sherman DR, Dowd DR, Jefcoat SC, Delisle RK 1995 The vitamin D receptor interacts with general transcription factor IIB. *J Biol Chem* 270:4748-4752
 54. Beekman JM, Allan GF, Tsai S, Tsai M, O'Malley BW 1993 Transcriptional activation by the estrogen receptor requires a conformational change in the ligand binding domain. *Mol Endocrinol* 7:1266-1274
 55. Nardulli AM, Lew D, Erijman L, Shapiro DJ 1991 Purified estrogen receptor DNA binding domain expressed in *Escherichia coli* activates transcription of an estrogen-responsive promoter in cultured cells. *J Biol Chem* 266:24070-24076
 56. Nardulli AM, Romine LE, Carpo C, Greene GL, Rainish B 1996 Estrogen receptor affinity and location of consensus and imperfect estrogen response elements influence transcription activation of simplified promoters. *Mol Endocrinol* 10:694-704
 57. Ptashne M 1986 Gene regulation by proteins acting nearby and at a distance. *Nature* 322:697-701
 58. Chodosh LA 1989 Mobility Shift DNA-Binding Assay Using Gel Electrophoresis. Greene Publishing Associates and Wiley Interscience, New York, pp 12.2.1-12.2.10
 59. Kerppola TK, Curran T 1991 DNA bending by Fos and Jun: the flexible hinge model. *Science* 254:1210-1214
 60. Thompson JF, Landy A 1988 Empirical estimation of protein-induced DNA bending angles: Applications to site-specific recombination complexes. *Nucleic Acids Res* 16:9687-9705



Estrogen Receptor Activation Function 1 Works by Binding p160 Coactivator Proteins

Paul Webb, Phuong Nguyen, Jeanette Shinsako,
Carol Anderson, Weijun Feng, Mimi P. Nguyen, Dagang Chen,
Shih-Ming Huang, Sujatha Subramanian, Eileen McKinerney,
Benita S. Katzenellenbogen, Michael R. Stallcup, and
Peter J. Kushner

Metabolic Research Unit (P.W., P.N., J.S., C.A., W.-J.F., M.P.N., P.J.K.)
University of California School of Medicine
San Francisco, California 94143-0540

Department of Pathology (D.C., S.-M.H., S.S., M.R.S.)
University of Southern California
Los Angeles, California 90033

Department of Physiology and Biophysics (E.M., B.S.K.)
University of Illinois
Urbana, Illinois 61801

Estrogen receptor- α contains two transactivation functions, a weak constitutive activation function (AF-1) and a hormone-dependent activation function (AF-2). AF-2 works by recruiting a large coactivator complex, composed of one or more p160s, CREB-binding protein (CBP)/p300, and P/CAF (p300 and CBP-associated factor), via direct contacts with the p160s. We report here that independent AF-1 activity also requires p160 contacts. Unlike AF-2, which binds signature NR boxes in the center of the p160 molecule, AF-1 binds to sequences near the p160 C terminus. We propose that the ability of AF-1 and AF-2 to interact with separate surfaces of the same coactivator is important for the ability of these transactivation functions to synergize. (Molecular Endocrinology 12: 1605-1618, 1998)

INTRODUCTION

Estrogen signal transduction is mediated by two receptors, α (hereafter ER) and β (ER β) (1-3). Both estrogen receptors (ERs) are members of the nuclear receptor family of conditional transcription factors (4, 5). In the absence of estrogen, the ERs are held in the nucleus in a complex with molecular chaperonins, such as HSP90. Upon estrogen binding, the receptors dissociate from the heat shock protein (HSP) complex and bind, as homodimers or heterodimers (6, 7), to a specific estrogen response element (ERE). From this

location on DNA, the receptor enhances transcription from the nearby promoter.

Transactivation is best understood within the context of the α -receptor. It is mediated by two activation functions, a weak constitutive activation function, AF-1, that lies within the ER N-terminal (AB) domain and a stronger estrogen-dependent activation function, AF-2, that lies within the ER ligand-binding domain (LBD) (8-14). Together, AF-1 and AF-2 synergize strongly to give the final overall level of estrogen activation. The ER transactivation functions are believed to work by binding coactivators and bringing them to the promoter (15-18). The AF-2 surface consists of a cluster of residues from helices 3, 5, and 12 (13, 19), that form a hydrophobic patch on the surface of the liganded LBD (19a). This hydrophobic patch binds a family of related proteins called the p160s, which include GRIP1/TIF2 (20-22), SRC-1 (23), and RAC3/pCIP/ACTR/AIB1 (24-27). In each case, AF-2 recognizes a specific signature motif (LXXLL), termed the NR box, that is found repeated throughout the p160s (22, 25, 28-30), and also within proteins that act as AF-2 repressors, such as RIP140 (28, 31) and TIF1 (32). GRIP1, for example, and its human homolog TIF2, contain three separate NR boxes (I, II, and III, respectively), of which NR boxes II and III are the most important for ER binding (22, 29). The p160s, in turn, interact with other coactivator proteins, including CREB-binding protein (CBP)/p300 (22, 25, 33, 34) and p300 and CBP-associated factor (P/CAF) (26, 35, 36). Together, this large coactivator complex, whose subunits possess both histone acetyl-transferase activity (26, 35, 37-39) and the capacity to bind the basal

transcription machinery (40, 41), is responsible for the ability of AF-2 to stimulate gene expression.

ER-AF-1, by contrast, is poorly understood. AF-1 usually displays little independent activity and serves only to synergize with AF-2 (10, 12, 14). For unknown reasons, however, AF-1 can also show strong independent activity in some cell types and on some promoters (12). Residues that are important for AF-1 activity are dispersed throughout a large region of the AB domain between amino acid 41 and amino acids 120–150, depending upon the cell type (13, 42, 43). Within this region, motifs that contribute to AF-1-independent activity (amino acids 41–64, hereafter Box 1) and synergism with the LBD (amino acids 87–108, hereafter Box 2) have been identified (44). AF-1 is also strong under conditions of mitogen-activated protein (MAP) kinase pathway stimulation (45–48). Several serine residues (S104, S106, S118) that are phosphorylated by MAP kinases, or cyclin-dependent kinases (49), have been identified, with the latter accounting for more than 90% of total ER phosphorylation (45, 46, 50). Each phosphorylated serine contributes to overall AF-1 activity (45, 46). Despite all this information, the AF-1 protein target or targets remain unknown. Furthermore, the reasons why AF-1 synergizes with AF-2 in some contexts, yet works independently in others, are also unknown.

While the exact nature of the prospective AF-1 coactivator complex is poorly defined, several lines of evidence suggest that it shares features with the AF-2 coactivator complex. First, AF-1 can mask weak mutations in AF-2, suggesting that AF-1 can compensate for reduced p160 recruitment (13). Second, AF-1 and AF-2 squelch each others activity, suggesting that they compete for common limiting target molecules (51). Third, the transcriptional activity of isolated AF-1 can be enhanced by introducing the free ER-LBD into the same cells (52). This activity requires that the LBD be liganded to estrogen and possesses an intact AF-2 surface. Furthermore, enhancement of AF-1 activity by the free LBD is potentiated by exogenous SRC-1 (53). Thus, the free LBD utilizes the AF-2 surface and p160 contacts to participate in the AF-1 coactivator complex.

In this paper, we ask whether AF-1 and AF-2 might indeed bind a similar complement of coactivator proteins. We find that overexpression of p160 proteins, especially GRIP1, increases the transcriptional activity of AF-1. Furthermore, AF-1 interacts with sequences near the GRIP1 C terminus, and this region is needed for the ability of GRIP1 to potentiate AF-1. We propose that p160s are a direct target for both AF-1 and AF-2.

RESULTS

GRIP1 Stimulates AF-1

To test whether AF-1 and AF-2 might utilize similar coactivators, we first examined whether GRIP1 would stimulate AF-1 activity. We transfected an ERE-dependent reporter [ERE-coll-CAT(choramphenicol

acetyltransferase)] into HeLa cells along with expression vectors for ER and GRIP1. We then determined the transcriptional activity of the promoter in cells treated with either ethanolic vehicle, estrogen, or the antiestrogen tamoxifen, which is a drug that inhibits AF-2 (54) but allows full AF-1 activity (12).

In the absence of exogenous GRIP1, wild-type ER elicited a strong estrogen response, but no tamoxifen response, from the ERE-coll-CAT promoter (Fig. 1A). This is consistent with previous observations that show that AF-1 has little independent activity at classical EREs in HeLa cells (10, 12, 55). In the presence of exogenous GRIP1, ER gave enhanced estrogen response but also gave significant activity both in the presence of tamoxifen and absence of ligand. The ability of GRIP1 to enhance the activity of tamoxifen-liganded ER was more clearly seen when we used an ER variant (ER-V400). This ER variant binds more tightly to the HSP complex than its wild-type counterpart and therefore lacks most of the spontaneous ligand-independent activity that ER usually displays in tissue culture (56, 57). Here, GRIP1 potentiated the estrogen response of ER-V400 and also elicited a large increase in tamoxifen response without the corresponding increase in basal activity. The tamoxifen response of ER-V400 was dependent upon the amounts of transfected GRIP1 and approached 40% of overall estrogen response at high amounts of GRIP1 (Fig. 1B). Several alternate ERE-responsive promoters also showed enhanced tamoxifen response in the presence of GRIP1 (Fig. 1C). We conclude that GRIP1 enhances the transcriptional activity of tamoxifen and estrogen-liganded ER.

We then confirmed that the effect of GRIP1 on tamoxifen-liganded ER was mediated by AF-1 (Fig. 1D). We asked whether GRIP1 would enhance the activity of mouse ERs (mORs) with either a truncation of the AB domain that eliminates AF-1 (mOR Δ AF-1), a double-point mutation that eliminates AF-2 (mORmAF-2), or both (mOR Δ AF-1/mAF-2) (13). In the absence of exogenous GRIP1, wild-type mOR elicited a strong estrogen response from the ERE-coll-CAT promoter. This response was reduced 50% by the AF-1 deletion and eliminated by the AF-2 mutation, consistent with the idea that estrogen response in the absence of GRIP1 predominantly reflects AF-2 activity. In the presence of GRIP1, wild-type mOR gave enhanced estrogen response and significant activity both in the presence of tamoxifen and absence of ligand, just as did the human ER. The AF-1 deletion (mOR Δ AF-1) retained strong activity in the presence of estrogen and absence of ligand, but showed no activity in the presence of tamoxifen. In contrast, the AF-2 mutant (mORmAF-2) gave strong tamoxifen and estrogen responses, but gave no activity in the absence of ligand. The double mutant (mOR Δ AF-1/mAF-2) showed no activity under any conditions. Thus, the spontaneous ligand-independent activity that is obtained in the presence of exogenous GRIP1 requires AF-2, and not AF-1. We speculate that this AF-2 dependent, ligand-independent, activity reflects stabilization of an ER-

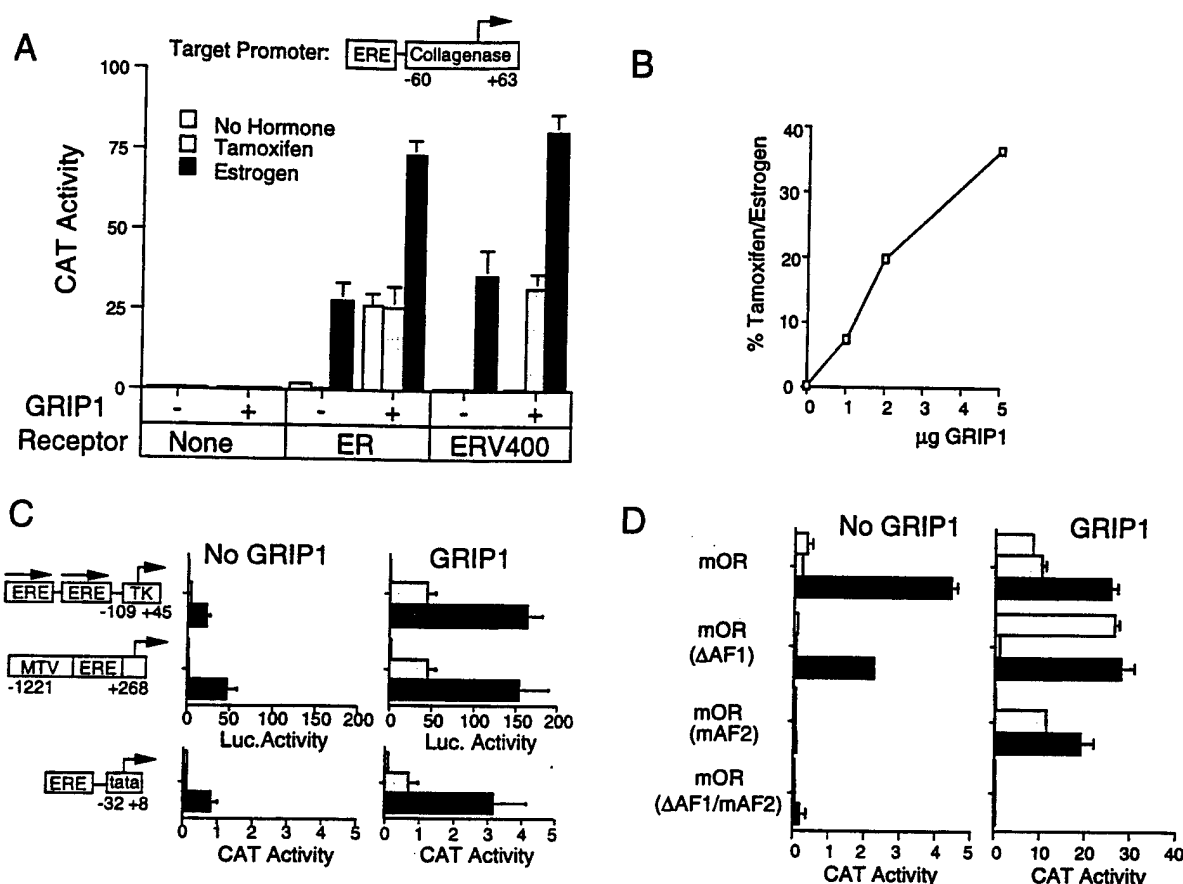


Fig. 1. GRIP1 Potentiates Tamoxifen Response

A, The effect of estrogen and tamoxifen upon expression of a transiently transfected ERE-coll-CAT reporter gene was examined in HeLa cells in the presence or absence of GRIP1. Where indicated, we also included empty expression vector SG5 (none) or ER expression vectors SG5-ER or SG5-ERV400. The transfected cells were treated with either ethanolic vehicle (\square), 5 μ M tamoxifen (gray bars) or 10 nM estradiol (black bars). CAT activities represent averages from triplicate wells from a representative transfection. B, Relative activity of tamoxifen vs. estrogen increases as a function of GRIP1 levels. Transfections utilizing SG5-ERV400 expression vector were performed as in Fig. 1A in the presence of increasing amounts of GRIP1 expression vector. The data are expressed as the percentage of tamoxifen response relative to estrogen response (set at 100%). C, GRIP1 enhances the activity of tamoxifen- and estrogen-liganded ER at a variety of promoters. The experiment was performed as described in Fig. 1A using a variety of ER-responsive reporters (from top to bottom): EREII-LUC (GL45), two vitellogenin EREs upstream of the herpes simplex virus TK proximal promoter ($-109/+45$); MTV-ERE-LUC, the MTV long terminal repeat region ($-1228/+268$) with a single palindromic ERE replacing the complex multiple hormone response element; ERE-TATA-CAT, a single ERE 19-mer upstream of the collagenase TATA box region ($-32/+8$). D, Analysis of tamoxifen action on ER mutants. Expression vectors for mouse ERs (mORs) were transfected into HeLa cells and their effect on ERE-coll-CAT expression was determined in the absence or presence of GRIP1. mORs were full length, contained a deletion of the first 120 amino acids of the AB domain (mOR Δ AF-1), an AF-2-disruptive double alanine for leucine substitution at positions 543 and 544 in LBD activation helix 12 (mORmAF-2), or both (mOR Δ AF-1/mAF-2).

LBD conformation that resembles that of the estrogen-liganded receptor by exogenous GRIP1. We also conclude that GRIP1 enhances the activity of the tamoxifen-liganded ER by enhancing the activity of AF-1 and not by enhancing the activity of AF-2.

p160s and p160-Associated Proteins Enhance AF-1 Activity

We next asked whether other coactivators would also enhance AF-1 activity. The p160 protein RAC3 enhanced both estrogen and tamoxifen response at

an ERE, and SRC-1a showed similar but weaker activity (Fig. 2A). This weak activity of SRC-1a is consistent with previous observations that show that this coactivator possesses little activity in HeLa cells (58), perhaps because SRC-1a contains an inhibitory function within its C-terminal domain (30). The AF-2 repressor RIP140 failed to enhance tamoxifen response. Of the p160-associated proteins, CBP and p300 enhanced estrogen and tamoxifen induction, especially in combination with GRIP1 (Fig. 2B). P/CAF only weakly enhanced tamoxifen activation (data not shown).

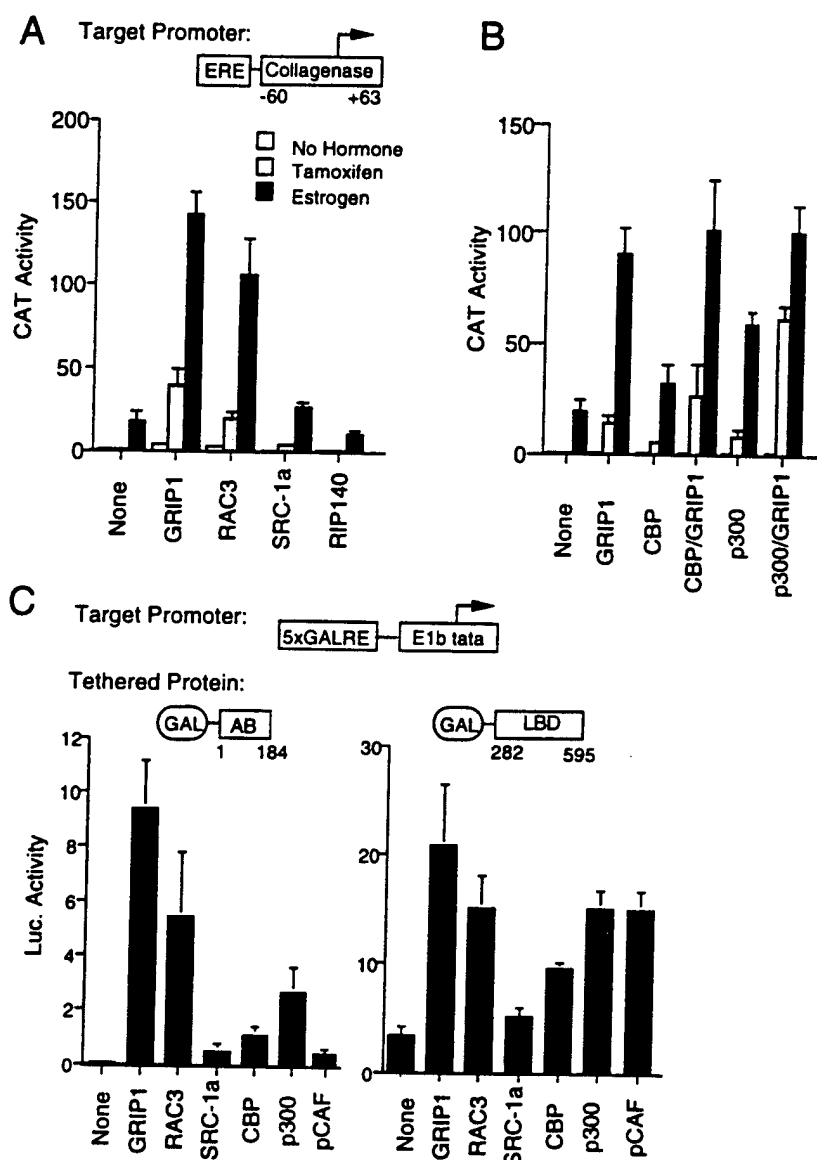


Fig. 2. A Survey of the Effect of Coactivators on Tamoxifen Response

A, Results of a transfection in which activity of ERE-coll-CAT reporter gene was monitored as above (Fig. 1A) in the presence of ER expression vector SG5-ERV400 and 5 μ g of each coactivator expression vectors. B, As above, using expression vectors for GRIP1, CBP, or p300. C, The effect of transient transfection of a range of coactivators upon transcription of a GAL4 responsive reporter gene (GK1) was determined in the presence of GAL4-AB or GAL4-LBD expression vectors. Individual bars represent luciferase activities determined from triplicate wells. In the case of the GAL-LBD, the cells were treated with 100 nM estradiol.

We then examined whether the same coactivators would enhance the activity of isolated AF-1. We transfected an expression vector for a GAL4 DNA-binding domain (DBD)/ER-AB domain fusion protein (GAL4-AB) into HeLa cells, along with expression vectors for various coactivators (Fig. 2C). In accordance with their effects on tamoxifen-liganded ER, GRIP1, RAC3, and SRC-1a each enhanced AF-1-dependent transcription from a GAL4-responsive reporter gene. CBP, p300, and, to a lesser extent, P/CAF enhanced the transcriptional activity of AF-1. As expected, each of the coactivators also enhanced the estrogen-dependent activity of a GAL4-LBD fusion protein. Thus, the p160s and

p300/CBP, and perhaps P/CAF, are involved in transactivation by both AF-1 and AF-2.

An Extended Region of AF-1 Is Required for GRIP1 Action

We next examined which region of AF-1 was needed for GRIP1 action. Three subregions of the AB domain have previously been implicated in AF-1 activity (see Fig. 3). Box 1 (amino acids 41-64) is needed for tamoxifen activation in MDA-231 breast cells (44). Box 2 (amino acids 87-108) is required for synergism with AF-2 in the same cell type. Sequences surrounding the

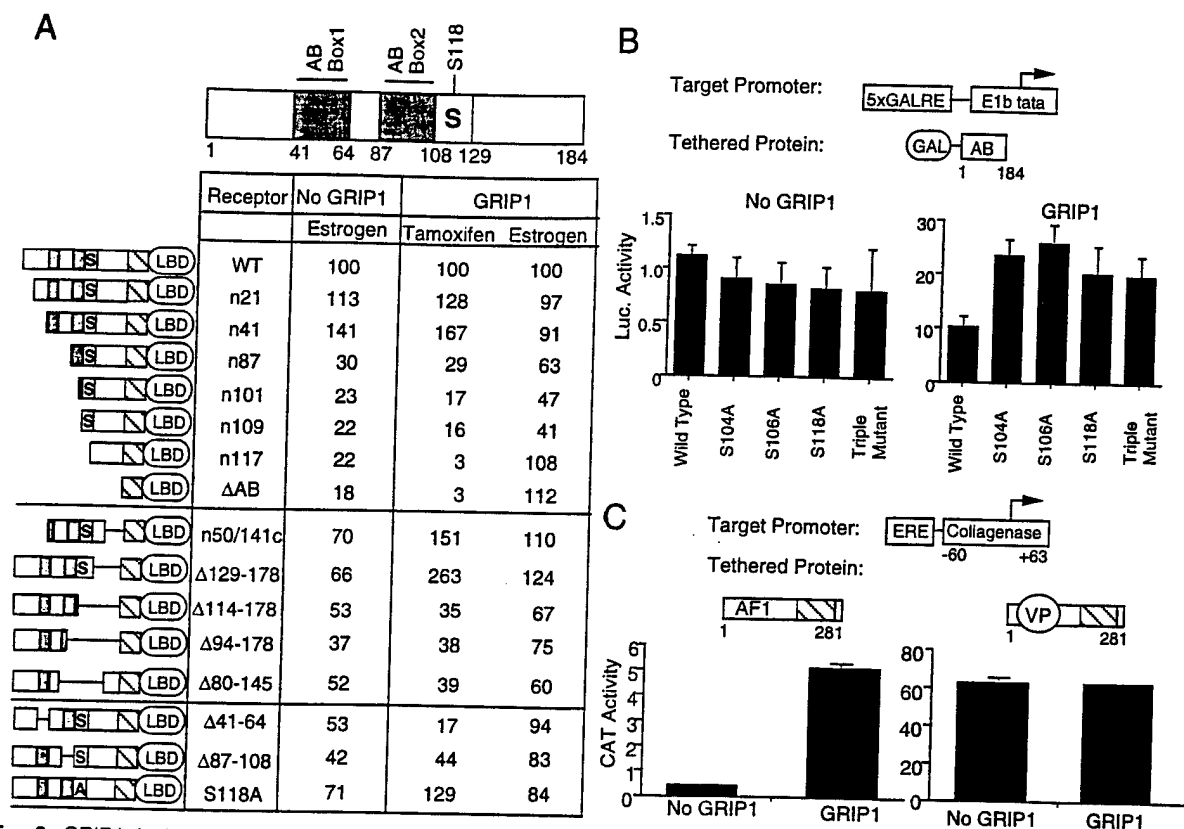


Fig. 3. GRIP1 Action on AF-1 Requires an Extended Region of the AB Domain

A, Effect of AB domain mutations upon ERE-dependent transcription in HeLa cells. A schematic of the ER AB domain showing each of the three AF-1 subregions, box 1 (41–64), box 2 87–108, and the KTR (108–129) is shown above. The locations of box 1 and box 2 are shown in gray. The KTR is boxed, and the major phosphorylation site Serine 118 is marked with S. A schematic of each mutation is shown on the left. In all cases, ERs contained mutations in the AB domain (amino acids 1–184) but retained an intact DBD and LBD. We tested the effect of mutations upon ER action on ERE-coll-CAT, as described in Fig. 1A, in the absence or presence of GRIP1. We set activity of wild-type ER (full length) at 100% in each column of data and compared the activity of mutant ER relative to this number under this condition. B, Effect of phosphorylation site mutations on GRIP1 action. HeLa cells were transfected, as in Fig. 2C, with a GRIP1 expression vector, where indicated, and GAL4-AB expression vectors bearing alanine substitutions at each, or all three, phosphorylation sites. C, Effect of a VP16 substitution on GRIP1 action. HeLa cells were transfected with expression vectors for either the ER AB-DBD region or a similar vector containing a VP16 transactivation domain inserted at amino acid 65, and either an empty expression vector or an expression vector for GRIP1.

main phosphorylated serine residue at position 118 [kinase target region (KTR), amino acids 108–129], also contribute to AF-1 activity (43). To test whether these subregions were needed for GRIP1 action, we transfected mutant human ERs into HeLa cells and assessed their ability to activate ERE-dependent transcription in the presence or absence of GRIP1 (Fig. 3A). In each case, the ERs contained mutations within the AB domain (amino acids 1–184), but retained an intact DBD and LBD. For simplicity, we compared the activity of mutant and wild-type ERs in the absence or presence of GRIP1 (see Fig. 1A) and set activity of wild-type ER at 100%.

We were able to confirm previous results (43) that showed that AF-1 activity in the absence of GRIP1 required an extended region of the AB domain (Fig. 3A, first column). Tamoxifen response in the presence of GRIP1 also required an extended region of the AB domain, but especially sequences in and around box 1 (Fig. 3A, second column). ERs lacking sequences up-

stream of amino acid 41 (n21, n41) showed normal, or enhanced, tamoxifen response, but an ER with an N-terminal deletion that eliminated box 1 (n87) showed markedly reduced tamoxifen response. ERs lacking box 1 and box 2 (n101, n109), or all three subregions (n117), gave further stepwise reductions in tamoxifen response. ERs lacking sequences downstream of amino acid 129 (n50/141c; Δ129–178) showed modestly enhanced tamoxifen response, but ERs with further C-terminal deletions that eliminate the KTR (Δ114–178), or both the KTR and box 2 (Δ93–178, Δ80–145), again showed reduced tamoxifen activation. Finally, an ER specifically lacking box 1 (Δ41–64) showed markedly reduced tamoxifen activation, and an ER specifically lacking box 2 (Δ87–108) showed a more modest reduction in tamoxifen activation. Thus, GRIP1 action requires each of the three AF-1 subregions and, especially, sequences within box 1.

We then specifically addressed the role of the AF-1 phosphorylation sites. An ER with an alanine substi-

tution at serine 118 (S118A) actually showed enhanced tamoxifen activation in the presence of GRIP1 (Fig. 3A). Similarly, introduction of alanine residues at each MAP kinase target serine (S104, S106, S118) also enhanced GRIP1 activation of a GAL4-AB fusion protein (Fig. 3B). Thus, GRIP1 action at AF-1 is independent of the phosphorylation sites.

Next, we compared the effect of GRIP1 on the ER AB-DBD region (AB-DBD), which only contains AF-1, with a similar ER truncation that contained an in-frame insertion of the VP16 activation function, just downstream of box 1 (V281C, Fig. 3C) (55, 59). As expected, GRIP1 enhanced the weak AF-1 activity that was obtained with the AB-DBD truncation. In contrast, GRIP1 had no effect on the stronger activity that was obtained with the VP16-ER fusion protein. Thus, insertion of a heterologous transactivation function into the AB domain abolishes the effect of GRIP1, supporting the notion that integrity of the proximal part of AF-1 (box 1) is essential for GRIP1 action. Together, our results show that GRIP1 enhancement of AF-1 activity requires an extended region of the AB domain and, especially, sequences in and around box 1.

ER AF-1 Binds p160s

We then asked whether the AB domain binds p160s (Fig. 4A). *In vitro* translated GRIP1 and SRC-1a bound to both the AB domain and the ER-LBD in a glutathione-S-transferase (GST) pull-down assay. Over the course of this study, the bacterially expressed AB domain retained about 5–20% of the amounts of GRIP1 and SRC-1a that were retained by equimolar amounts of bacterially expressed LBD. We were unable to demonstrate binding of the AB domain to CBP (data not shown). We conclude that the AB domain, like the LBD, binds p160s.

To examine which part of the AB domain was needed for GRIP1 binding, we produced a series of GST fusion proteins containing portions of the AB domain and asked whether they would bind *in vitro* translated GRIP1 (Fig. 4B). An AB domain fusion protein with an N-terminal deletion to amino acid 37 (38–161) bound GRIP1, but AB domains with larger N-terminal deletions (55–161; 67–184; 82–161) failed to bind GRIP1. Two AB fusion proteins with C-terminal deletions (1–145; 1–116) each bound GRIP1 efficiently. Indeed, the latter showed enhanced binding relative to the native AB domain. An AB fusion protein with a further deletion (5–90) retained some, albeit reduced, GRIP1 binding, but two portions of the AB domain that overlap sequences 5–90 (1–67 or 55–90) failed to bind GRIP1. Similar results were obtained with SRC-1a (data not shown). Thus, an extended region of the AB domain (amino acids 38–116), and especially sequences between amino acids 38–55 and upstream of amino acid 90, were needed to bind p160s. We conclude that both GRIP1 enhancement of AF-1 activity (Fig. 3A, second column), and GRIP1 binding (Fig. 4), require a similar region of the AB

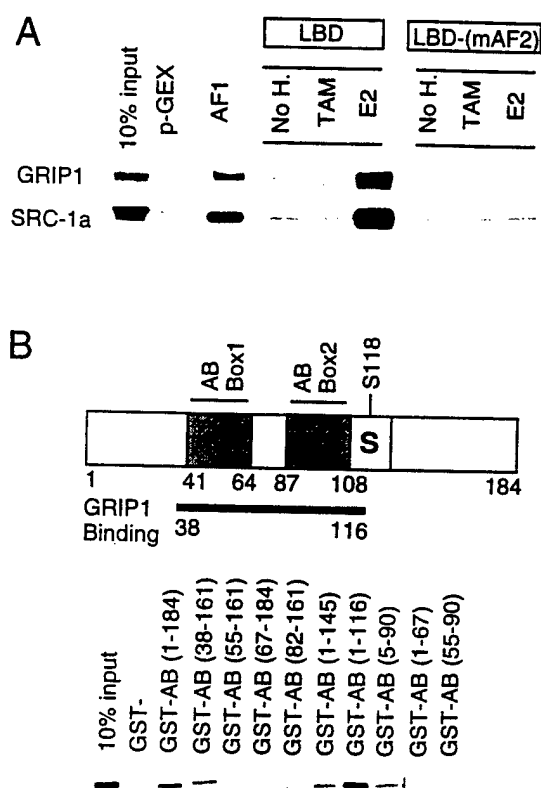


Fig. 4. The Estrogen Receptor AB Domain Binds p160s

A. The AB domain binds p160s. Autoradiographs of labeled proteins retained on beads in GST pull-down assays. The binding of either labeled GRIP1 or SRC-1a to beads coated with GST, GST-AB, or GST-LBD fusions. **B.** Binding of GRIP1 to the AB domain. A diagram of the AB domain is shown above with positions of previously described AB transactivation motifs, box 1, box 2, and the KTR-containing serine 118, marked. The extent of AB and the KTR implicated in GRIP1 binding is marked below with a thick black line. Below, autoradiography of labeled GRIP1 retained on GST beads is presented. Beads are coated with GST fused to portions of the AB domain between the amino acids that are described.

domain, and, especially, sequences in and around box 1. This suggests that AF-1, like AF-2, works by binding p160 proteins.

AF-1 Binds the GRIP C Terminus

We next asked how AF-1 recognized GRIP1 (Fig. 5A). Competition experiments revealed that a specific peptide that was homologous to GRIP1 NR box III blocked GRIP1 binding to the LBD but failed to block GRIP1 binding to the AB domain, or to the p160-binding region of CBP (35) (Fig. 5B). Similar results were also obtained with a peptide homologous to NR box II (data not shown), or when SRC-1a was substituted for GRIP1 and tested with either peptide (data not shown). Thus, the AB domain recognizes a region of GRIP1 and SRC-1a that is distinct from the NR boxes.

To localize the AB binding site within GRIP1, we analyzed binding of the AB domain and LBD to a series of GRIP1 truncations (Fig. 5C). The AB domain

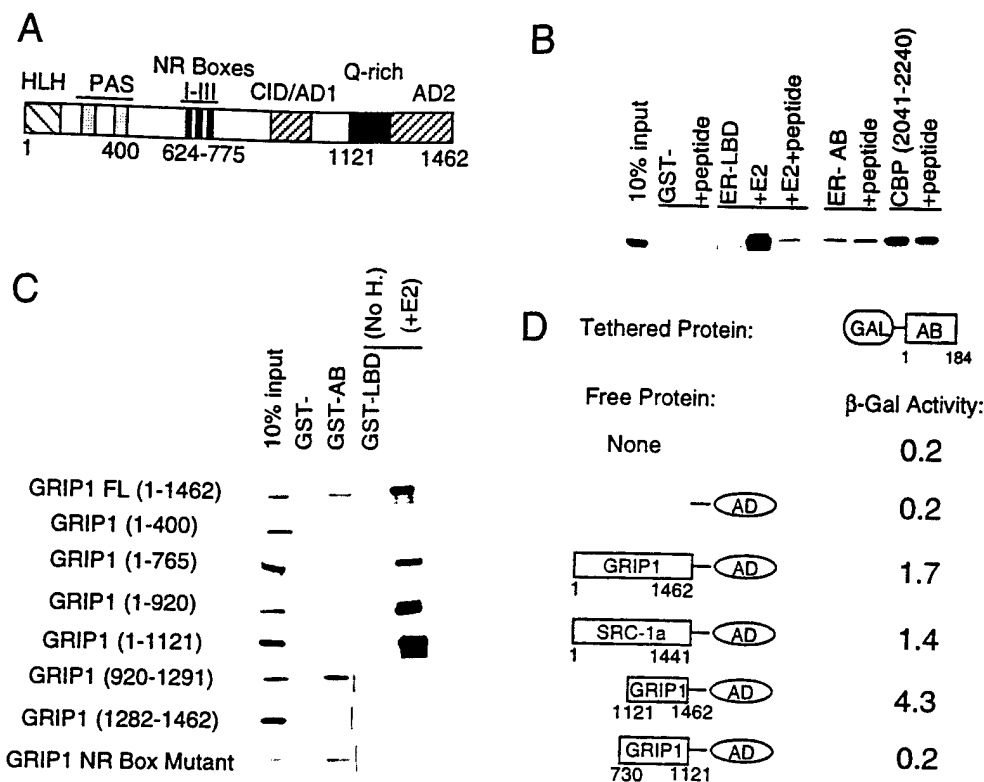


Fig. 5. The ER-AB Domain Binds to the GRIP1 C Terminus

A, Schematic of full-length GRIP1. Approximate positions of putative and defined functional domains are marked, including β HLH/PAS region, NR boxes I-III, CBP interaction domain (CID/AD1), Q-rich domain, and activation domain 2 (AD2). B, Autoradiography of GRIP1 protein retained on beads coated with either GST-LBD, GST-AB, or GST-CBP fusion proteins. The CBP fragment consists of amino acids 2041-2240 and contains the previously defined p160-binding motif. Where indicated (+peptide), binding to GRIP1 was performed in the presence of an excess of a synthetic peptide homologous to NR box III. We also performed controls with an NR box peptide that contained double alanine substitutions at key leucine residues (data not shown). C, Binding of GST-AB or GST-LBD to GRIP1 protein fragments. Autoradiograms show retention of labeled full-length GRIP1 or GRIP1 fragments on beads coated with GST-AB or GST-LBD fusion proteins. Amino acid numbers of all fragments were indicated. The GRIP1 NR box mutant has alanine substitutions in the key leucine doublets within NR boxes II and III (N-LXXLL-C to N-LXXAA-C). D, Interaction of ER-AF-1 and p160s in a yeast two-hybrid system. β -Galactosidase production from a GAL4-responsive reporter gene was determined in yeast strains containing GAL4DBD-AB and free GAL4AD-GRIP1 or GAL4AD-SRC-1a fusion proteins.

failed to bind GRIP1 truncations lacking amino acids beyond positions 400, 765, 920, or 1121, respectively, but did bind a GRIP1 fragment containing amino acids 920-1291 and, more weakly, to a fragment containing amino acids 1282-1462. In contrast, the LBD bound all C-terminal truncations that contained intact NR boxes (1-765; 1-920; 1-1121), but failed to bind either C-terminal GRIP1 fragment. As predicted, the AB domain bound to a GRIP1 NR box mutant (boxes II and III mutated) efficiently, and the LBD bound this molecule only weakly. Experiments using GST-GRIP1 fragments also indicated that GRIP1 sequences downstream of amino acid 1121, and especially sequences 1121-1282, were needed for ER binding (data not shown). These results confirm that AF-1 recognizes GRIP1 at a site that, most likely, lies between amino acids 1121 and 1462 and is distinct from the NR box cluster. Amino acids 1121-1282, which contribute most to AF-1 binding, overlap a glutamine rich (Q-rich)

domain that is conserved throughout the p160 family (26).

We then confirmed the location of the putative GRIP1 AB domain-binding site in a yeast two-hybrid system (Fig. 5D). A GAL4-AB fusion protein weakly enhanced transcription of a GAL4-responsive β -galactosidase gene in yeast, relative to the levels of transcription obtained with the GAL4-DBD alone (data not shown). This is consistent with previous studies indicating AF-1 is weakly active in yeast cells (12). ER-AF-1 activity was unaffected by coexpression of the GAL4 acid activation domain (GAL-AD), but was enhanced by coexpression of GAL4-AD/full-length GRIP1 or SRC-1a fusion proteins. A GAL-AD/GRIP1 fusion protein, containing C-terminal amino acids 1121-1462, also enhanced AF-1 activity. In contrast, a GAL-AD/GRIP1 fusion protein containing amino acids 730-1121 did not enhance AF-1 activity. Similarly, GAL-AD fusions to GRIP1 amino acids 563-1121 and

1-766 also failed to enhance AF-1 activity (data not shown). We stress that the GAL-AD/GRIP1 fusions containing GRIP1 fragments 730-1121 (20), 563-1121, and 1-766 (data not shown) all strongly enhanced the activity of a GAL-ER-LBD fusion protein in yeast, in agreement with the fact that each of these fragments contains NR boxes, and confirming that these fragments are efficiently expressed in yeast cells. Thus, GRIP1 (and SRC-1a) target the GAL4-AD to the AB domain in yeast, and this activity is specifically recapitulated by the GRIP1 C terminus. Furthermore, both the GST pull-down and yeast two-hybrid assays indicate that the C-terminal region of GRIP1 is the only region that interacts with the ER-AB domain in these systems.

AF-1 Function Requires the GRIP1 C Terminus

We next tested whether the GRIP1 C terminus was needed for AF-1 activity in mammalian cells. We co-expressed GAL4-AB or GAL4-LBD fusion proteins with either wild-type GRIP1, a GRIP1 truncation that cannot bind AF-1 (GRIP- Δ 1121C), or a GRIP1 molecule with NR box mutations (NR box mutant) (Fig. 6A). The GRIP1 truncation, GRIP- Δ 1121C, was completely deficient for AF-1 enhancement, but showed increased AF-2 enhancement relative to wild-type GRIP1. In contrast, the GRIP1 NR box mutant strongly enhanced AF-1 activity, but failed to enhance AF-2 activity. Thus, GRIP1 needs an intact C terminus to enhance AF-1, but this region is dispensable for GRIP1 enhancement of AF-2. Furthermore, NR boxes

II and III are not needed for GRIP1 enhancement of AF-1 activity.

We then asked how the GRIP1 truncation (Δ 1121C) would affect the activity of full-length ER (Fig. 6B). For these experiments we used a promoter that shows some tamoxifen activation in the absence of exogenous p160s (ERE-II-LUC; Fig. 6B, *inset*). As expected, GRIP- Δ 1121C enhanced the action of the estrogen-liganded ER at the ERE-II-LUC promoter, although this response was somewhat lower than that obtained with wild-type GRIP1. In contrast, GRIP- Δ 1121C eliminated the tamoxifen response that was seen in the absence of exogenous p160s (see *inset*). A combination of wild-type GRIP1 and GRIP- Δ 1121C gave increased estrogen response relative to the responses obtained with wild-type GRIP1 cotransfected with empty expression vector. [Note that this enhanced estrogen response occurs because wild-type GRIP1 and the GRIP- Δ 1121C truncation both activate isolated AF-2; therefore, this transfection contains more coactivator protein that is capable of activating AF-2 than the transfection containing wild-type GRIP1 alone]. Nonetheless, the tamoxifen response that was obtained with this combination of coactivators was significantly reduced relative to wild-type GRIP1 alone, showing that the GRIP- Δ 1121C truncation also inhibited the enhanced tamoxifen response that was obtained in the presence of exogenous GRIP1. We conclude that the GRIP1 C terminus is needed for enhancement of AF-1 activity and that GRIP- Δ 1121C acts as a dominant negative for tamoxifen response. Because GRIP- Δ 1121C is weakened in its ability to

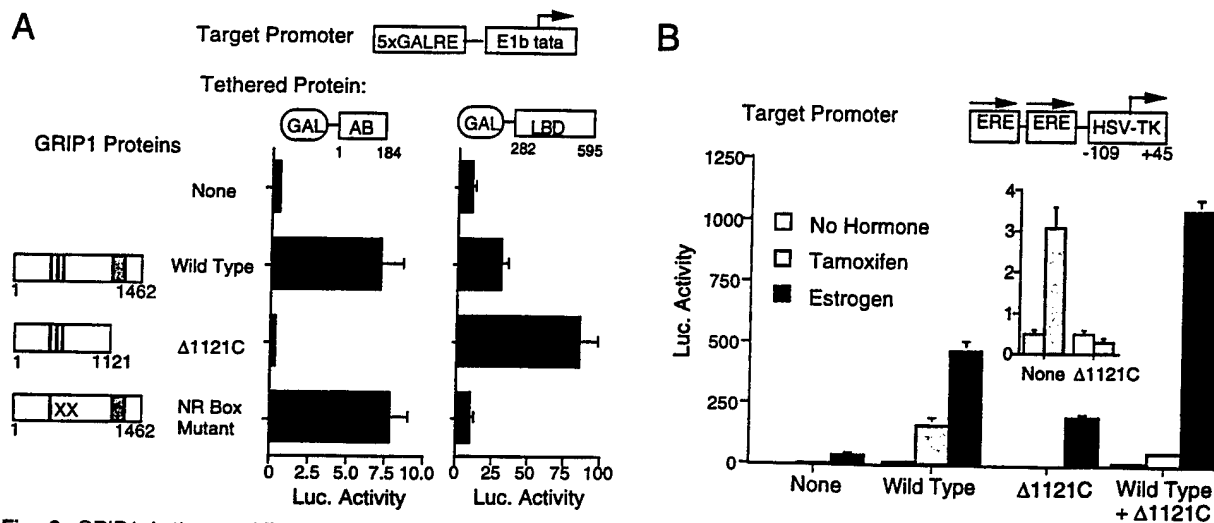


Fig. 6. GRIP1 Action on AF-1 Requires the GRIP1 C Terminus

A, Differential effects of GRIP1 mutations on AF-1 and AF-2. A schematic of the GRIP1 proteins is shown on the left of the figure. Intact NR boxes are marked with black lines, mutant NR boxes with an X, and the GRIP1 Q-rich domain with a dark gray box. The Δ 1121C truncation lacks sequences downstream of position 1121 and will not bind the AB domain. The GRIP1 NR box mutant contains double leucine to alanine substitutions within NR boxes II and III. The effect of the GRIP1 mutants upon GAL-AB and GAL-LBD fusion proteins was determined as in Fig. 3A. B, Requirement of the GRIP1 C terminus for ER action. The effect of tamoxifen and estrogen upon expression of an EREII-LUC reporter gene was examined in HeLa cells in the presence of ERV400 and GRIP1 or the GRIP- Δ 1121C truncation. The *inset* shows effect of wild-type and truncated GRIP1 upon EREII-LUC expression in the presence of tamoxifen, on an expanded scale.

enhance estrogen response in the presence of full-length ER, yet strengthened in its ability to enhance isolated AF-2, we also suggest that the GRIP1 C terminus is needed for AF-1/AF-2 synergism.

GRIP1 Is a Weak Activator of ER β AF-1

Previous observations have suggested that ER β , unlike ER(α), shows little activity in the presence of tamoxifen (60, 61). We therefore examined the role of p160s in ER β function. Wild-type GRIP1 strongly enhanced the activity of unliganded and estrogen-liganded ER β (Fig. 7A), consistent with similar results that were obtained with SRC-1 (60), but only weakly increased the activity of tamoxifen-liganded ER β . In parallel, GRIP1 markedly increased the activity of tamoxifen-liganded ER(α) and failed to affect the activity of an ER truncation lacking of the AB domain (ER Δ AB). Thus, ER β lacks an efficient AF-1 activity that responds to GRIP1.

We then compared binding of *in vitro* translated ERs to GST-GRIP1 fusion proteins (Fig. 7B). A GRIP1 fragment overlapping the NR boxes (563–1121) bound ER β , but only in the presence of estrogen and not

tamoxifen. In contrast, a GRIP1 C-terminal fragment (1121–1462) bound weakly to ER β but strongly to the ER(α) control. As expected, ER Δ AB also showed markedly reduced binding to this C-terminal fragment (data not shown). Thus, the strength of the interaction of the C terminus of GRIP1 with the two isoforms of ER parallels the ability of GRIP1 to potentiate the activity of either tamoxifen-liganded ER β or ER(α). This result further underlines the importance of the interactions between the ER(α) AB domain and the GRIP1 C terminus.

DISCUSSION

AF-1 Works by Binding p160s

This paper presents evidence that describes a direct target for the ER α AF-1 function. We find that AF-1 activity, in the context of tamoxifen-liganded ER or the isolated AB domain, is strongly enhanced by overexpression of p160s and CBP/p300 and, somewhat weakly, by overexpression of P/CAF. We find that the AB domain binds GRIP1 and SRC-1a. Furthermore, a similar extended region of the AB domain is required for GRIP1 potentiation of AF-1 activity *in vivo* and for GRIP1 binding *in vitro*. We were also able to identify a GRIP1 truncation that does not bind the AB domain and acts as a specific dominant negative for tamoxifen-liganded ER, either in the absence of exogenous p160s or the presence of overexpressed GRIP1. We therefore propose that AF-1 action requires a p160/p300 complex and that direct contacts with p160s are essential for recruitment of this complex. Our study complements a recently published study which shows that microinjection of anti-p160 or anti-CBP antibodies block tamoxifen response in cells (HepG2) where AF-1 activity is high (62).

GRIP1 strongly enhances independent AF-1 activity. In the absence of GRIP1, however, AF-1 largely serves to synergize with AF-2. We suggest that AF-1/p160 interactions may also play a role in AF-1/AF-2 synergism. ER N-terminal deletions have similar effects on estrogen response in the absence of GRIP1 and tamoxifen activation in the presence of GRIP1. Likewise, box 1 (amino acids 41–64), which is responsible for tamoxifen activation in breast cells (44), is needed both for AF-1/AF-2 synergism in the absence of GRIP1 and tamoxifen activation in the presence of GRIP1. Finally, the GRIP1 truncation (Δ 1121C), which does not bind the AB domain, does not potentiate the activity of estrogen-liganded ER as strongly as wild-type GRIP1, yet potentiates the activity of isolated AF-2 better than wild-type GRIP1 (Fig. 6). Thus, GRIP1- Δ 1121C is defective for potentiation of independent AF-1 activity and AF-1/AF-2 synergism. Together, these results suggest that the ability of AF-1 and AF-2 to bind separate surfaces of the same coactivator is important for the ability of these transactivation functions to synergize.

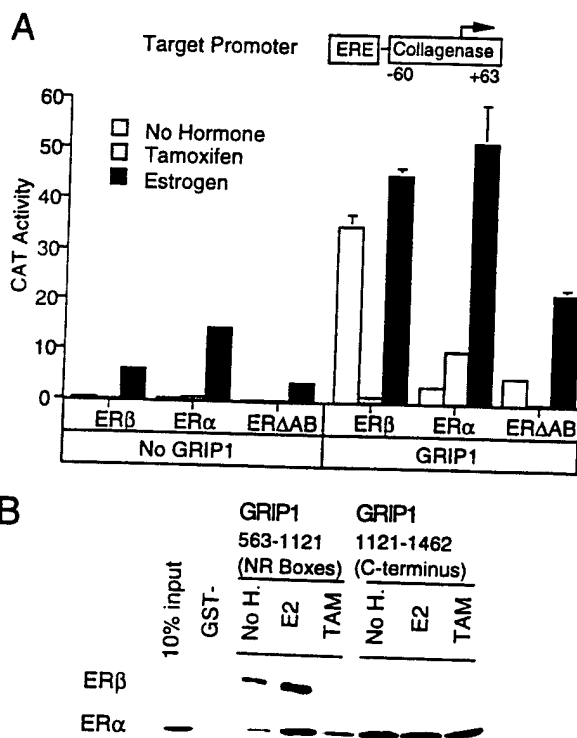


Fig. 7. GRIP1 Is a Weak Activator of ER β AF-1

A, GRIP1 action on ER β . We tested the effect of GRIP1 upon ERE-coll-CAT expression in the presence of CMV-driven expression vectors for human ER β , ER(α), or an ER lacking an AB domain (ER Δ AB). **B**, The GRIP1 C terminus binds ER constitutively. Autoradiograms of *in vitro* translated full-length ER β or ER(α) retained on glutathione beads coated with GST-GRIP1 fragments (563–1121 and 1121–1462) in the presence of ethanolic vehicle, tamoxifen, or estradiol.

While we, and others (62), have readily detected protein-protein interactions between ER AF-1 and p160s, two other groups failed to do so. One set of authors failed to see interaction between AF-1 and TIF-2 in a yeast two-hybrid system (22). In that study, rather than full-length TIF-2 fused to the GAL-AD, the authors used a TIF-2 fragment spanning amino acids 624-1288. Another set of authors failed to see interactions between mouse AF-1 and SRC-1 in GST pull-downs (30). While we have no explanation for this discrepancy, we note that the second set of authors did see that a C-terminal fragment of SRC-1, which overlaps the Q-rich domain, bound full-length mouse ER in an AF-2-independent manner. We speculate that this interaction may be similar to the one that we have described in this paper.

The idea that both AF-1 and AF-2 directly bind p160s may account for several unexplained observations. First, AF-1 can compensate for weak AF-2 mutations (13). This could be explained if AF-1/p160 contacts assist weakened AF-2 interactions with p160s. Indeed, the ability of AF-1 to mask AF-2 mutations is lost in AB domain truncations that, we now know, lack the ability to bind GRIP1. Second, AF-1 and AF-2 squelch each others activity (51). This could be explained if AF-1 and AF-2 were both able to bind and to sequester limiting target p160s molecules. Third, free ER-LBD can enhance the activity of isolated AF-1 *in trans* (53). This could be explained if the LBD binds the NR boxes of p160s that, themselves, are bound to AF-1. Fourth, it has been previously shown that SRC-1 enhances the ability of tamoxifen-liganded ER to activate the C3 complement promoter in CV-1 cells (58). This could be explained if SRC-1 were simply enhancing AF-1 activity. Finally, our results may explain why tamoxifen behaves as a complete antagonist of ER β action at EREs (60, 61). GRIP1 potentiates the activity of tamoxifen-liganded ER β weakly and the GRIP1 C-terminus binds ER β weakly. The lack of ER β -AF-1 activity may reflect poor interaction between ER β and the GRIP1 C terminus.

GRIP1 Action and GRIP1 Binding Are Mediated By A Similar Region of AF-1

In this study we asked which regions of the AB domain were needed for GRIP1 stimulation of AF-1 activity and for direct GRIP1 binding by the AB domain. We found marked similarities between AF-1 and the p160-binding unit. First, a similar extended region of the AB domain is required for GRIP1 activation (amino acids 41-129) and GRIP1 binding (amino acids 38-116). Second, box 1 (amino acids 41-64), which is also required for tamoxifen response in breast cells (44), is important for both GRIP1 activation and GRIP1 binding. Third, the kinase target residue serine 118 is dispensable for both GRIP1 action and GRIP1 binding. Lastly, sequences toward the C terminus of the AB domain inhibit both GRIP1 activation and GRIP1 binding. These results strongly suggest that GRIP1 activa-

tion of AF-1 is mediated by GRIP1 binding. We speculate that this extended region of the AB domain forms a folded AF-1 structure that, like AF-2 (19a), contains p160-binding residues along the length of its primary sequence. Our data particularly implicate box 1 in GRIP1 activation and GRIP1 binding. While we favor the possibility that box 1 contains residues that contact GRIP1, it remains possible that mutations in box 1 are particularly deleterious to overall AF-1 structure. It will be important to identify and mutate specific residues on the surface of the folded AF-1 structure to fully define the p160-binding site.

The fact that a large region of the AB domain is required for GRIP1 action raises the question of whether AF-1 might bind other target proteins. Our *in vitro* binding studies reveal that sequences within box 1 are absolutely required for GRIP1 binding (Fig. 4), yet ERs with deletions of box 1 (Δ 41-64; n87, Fig. 3) retain 17-29% of the GRIP1-induced tamoxifen response. One explanation for this discrepancy is that, because transcriptional activity was determined in the presence of AF-2, then residual interaction between GRIP1 and the LBD masks the phenotypes of the AF-1 deletions. Alternatively, the AF-1 mutants might fold differently *in vivo* and *in vitro*. More interestingly, if regions of AF-1 downstream of box 1 make contacts with other constituents of the coactivator complex, such as p300 or P/CAF, then transfected GRIP1 might stimulate AF-1 activity through these intermediate proteins, even in the absence of its own binding site.

Two other lines of evidence point to the existence of alternate AF-1 targets. While the AF-1 phosphorylation sites themselves are dispensable for GRIP1 action, they are needed for AF-1 activity under conditions of MAP kinase stimulation (Refs. 45 and 46 and W. J. Feng, P. Webb, J. Li, M. Karin, J. D. Baxter, and P. J. Kushner, manuscript submitted). This suggests that the AF-1 phosphorylation sites make contacts with other proteins. The existence of a subregion of the AB domain that inhibits GRIP1 action is also suggestive. Previous reports have indicated that tamoxifen-liganded ER binds the corepressors N-COR and SMRT (63) and that the overall level of activity of tamoxifen-liganded ER is regulated by the levels of corepressors (58, 62, 64, 65). Indeed, one group has shown that ER/corepressor interactions involve serine 118 (64). We are exploring the possibility that the inhibitory region of the AB domain binds corepressors.

The GRIP1 C Terminus Is Needed for AF-1 Activity

AF-1/GRIP1 interaction requires sequences at the GRIP1 C terminus and especially sequences that overlap the Q-rich domain. This region is conserved within the p160 family, suggesting that it may also be involved in the interaction of other p160s with the AB domain. The exact nature of the AF-1-binding site in the GRIP1 C terminus is likely to be complex. While the Q-rich domain is essential for strong AF-1 binding, a

GRIP1 fragment (1282–1462), which is downstream of the Q-rich region, also binds weakly to AF-1. Thus, the AF-1-binding site may be spread across a large region of GRIP1. We also find that the AB domain readily binds free GRIP1 in GST pull-down assays, but the reciprocal interaction between tethered GRIP1 and the free AB domain is weak (data not shown). We do see strong hormone-independent binding of full-length ER to the GRIP C terminus in GST pull-down assays (Fig. 8B), and this binding requires the AB domain (data not shown). Thus, contributions from the rest of the ER molecule assist AF-1/GRIP1 interactions. One explanation for this phenomenon is that interaction between the GRIP1 C terminus and ER-AF1 requires receptor dimerization functions, as does interaction between the p160 NR box region and ER-AF2 (30). Another explanation is that the ER DBD-LBD region contributes weak GRIP1 contacts that help stabilize the AF-1/GRIP1 interaction. We are actively investigating these possibilities.

Our work does not address whether AF-1 binding is the only role of the GRIP1 C terminus in AF-1 activity. Recent studies have suggested that transcription factors may bind the same coactivator, but require different functions within that coactivator (66, 67). It is therefore possible that AF-1 and AF-2 bind p160s but, at the same time, require different p160 functions. TIF2 (the human homolog of GRIP1) has two independent transactivation functions, AD1 and AD2 (22). AD1, which binds CBP/p300, lies downstream of the AF-2-binding site (NR boxes) in the middle of the p160 molecule. Intriguingly, AD2, whose target is unknown, lies downstream of the AF-1-binding site within the TIF2 C terminus. Likewise, p160 HAT activity (26, 39), which is dispensable for AF-2 (66, 67), maps to the same p160 Q-rich region that is involved in AF-1 binding. Finally, SRC-1a contains an inhibitory function within its C terminus (30). A GRIP1 (Δ 1121C) truncation activated isolated AF-2 better than wild-type GRIP1, suggesting that the GRIP1 C terminus may also contain inhibitory sequences. AF-1 action could specifically require any of these C-terminal GRIP1 functions.

The Choice Between AF-1/AF-2 Synergism and Independent AF1 Activity May Be Regulated by p160s

GRIP1 enhances the ability of AF-1 to work independently of AF-2. This suggests a simple hypothesis to explain why AF-1 synergizes with AF-2 in some cells, but works independently in others. We propose that the relatively weak AF-1/p160 contacts ordinarily support the stronger interaction of the ER-LBD with p160s, and this explains why AF-1 synergizes with AF-2. At increased p160 levels, however, AF-1/p160 contacts become sufficient to recruit p160s independently. Thus, the balance between AF-1/AF-2 synergism and AF-1 independent activity would be regulated by p160 levels. In a sense, the behavior of AF-1

in this model is analogous to the previously described behavior of weak AF-2 mutations that reduce GRIP1 binding, and whose phenotype is suppressed at high levels of GRIP1 (19a, 68).

It was previously suggested that the agonist/antagonist behavior of tamoxifen in breast cells is influenced by cell-specific differences in factors that interact with subregions of the AB domain, such as box 1 (44). The results presented here suggest that those factors are p160 proteins. It is known that p160 levels are increased in certain breast and ovarian cancers because one of the p160 genes, AIB1, lies in an area of the genome that is commonly amplified in tumors (27). We have previously examined the properties of ERE-coll-CAT in MCF-7 and MDA-MB-453 breast cells that, we now know, contain amplified AIB1 (55). In accordance with our latest hypothesis, we found strong AF-1 activity. The similarity between the behavior of AF-1 in MDA-MB-231 cells (53) and HeLa cells in the presence of GRIP1 (present paper) may suggest that MDA-MB-231 cells also contain amplified AIB1. It will be interesting to test how p160 overexpression might affect the tamoxifen or estrogen responsiveness of genes with different promoter architectures, such as spaced half-sites (69) or AP-1 sites (55), whose transcription may correlate better with estrogen and antiestrogen effects on growth.

MATERIALS AND METHODS

Mammalian Reporter Genes and Expression Vectors

ERE-coll60-CAT (55), ERE-II-LUC (61), MTV(ERE)-LUC (70), and ERE-tata-CAT (14) have been previously described. The GAL4-responsive reporter gene GK1 contains five GAL4 response elements upstream of a minimal adenovirus E1b promoter that has been previously described (68) but, for the purposes of these studies, was moved into a pUC vector backbone devoid of the pUC AP1 site (71).

ER expression vectors have been previously described. For ease of comparison, we have given ER expression vectors a consistent nomenclature. The previous names and sources of each construct are as follows: SG5-ER = HEG0; SG5-ERV400 = HE0 (57); n21; n41 = E41; n87 = A87; n109 = M109; ER Δ AB; Δ 41–64; Δ 87–108 (44), n101 = HE302; n117 = HE303; n50/141c = HE344/368; Δ 129–178 = HE316 (43). MOR Δ AF-1 = mOR121–599; MORmAF-2 = mORL543A,L544A; MOR Δ AF-1/mAF-2 = MOR121–599/543A,L544A (13).

Coactivator expression vectors (GRIP1, RAC3, SRC-1a, RIP-140, p300, CBP, and P/CAF) have each been previously described (24, 29, 33–35, 39, 72).

Cell Culture and Transfection

HeLa cells were maintained and transfected as previously described (55). Unless otherwise indicated, the transfections contained 5 μ g CAT or luciferase reporter, 1 μ g pJ3 β -galactosidase control, 1 μ g ER expression vector, and 5 μ g coactivator expression vector. Cell lysates were prepared 20–40 h after transfection and CAT, luciferase, and β -galactosidase assays were performed using standard methods (55). In each case CAT and luciferase activities were corrected for variations in transfection efficiency using the β -galactosidase activity.

GST-Fusion Proteins

GST-AB and GST-LBD fusions have been previously described (55). GST AB truncations were prepared by amplification of AB sequences with standard PCR methods and the resulting fragments were cloned in frame into pGEX-5x-1 using appropriate restriction sites. GST-AB fragment 1-67 was prepared by digesting GST-AB with *NotI* and self-ligating. Fragment 65-184 was prepared by cloning the excised *NotI* fragment from GST-AB into pGEX-5X-2.

The GST-GRIP1 fragments were amplified by standard PCR methods and cloned into pGEX-2TK (563-1121) as a *BamHI/EcoRI* fragment or pGEX-4T1 (1121-1462) as a *EcoRI/XhoI* fragment.

In Vitro Translation Vectors

GRIP1, GRIP1 NR mutant, and SRC-1a transcription/translation vectors have been previously described (29, 39). GRIP1 C-terminal truncations (400, 765, 920) were prepared by restriction digestion and run-off transcription/translation. The GRIP1 C-terminal truncation Δ 1121C was prepared by digesting full-length GRIP1 with *XhoI* and *XbaI* and inserting a comparable fragment from an original partial GRIP1 cDNA clone 730-1121 (20). GRIP1 fragments 920-1291 and 1282-1462 were prepared by standard PCR methods. In both cases, the 5'-oligonucleotide used for amplification contained an in-frame Kozak consensus sequence. These fragments were inserted into pSG5 between the *EcoRI* and *BamHI* sites.

Protein-Binding Assays

GST-pull downs were performed as previously described (55). Production and use of competitor peptides and yeast two-hybrid assays are described elsewhere (29). For these studies a cDNA fragment coding for the ER-AB domain (amino acids 1-184) was PCR amplified and cloned into the *EcoRI/BamHI* sites of pGBT9 (CLONTECH, Palo Alto, CA). GRIP1 and SRC-1a/GAL4-AD fusions were also previously described (29).

Acknowledgments

We thank Drs. P. Chambon, D. Metzger, C. Glass, M. Parker, Y. Nakatani, J. D. Chen, and B. W. O'Malley for generous gifts of plasmids.

Received April 17, 1998. Revision received June 2, 1998.
Accepted July 2, 1998.

Address requests for reprints to: Peter J. Kushner, Metabolic Research Unit, University of California, San Francisco, California 94143-0540. E-mail: kushner@itsa.ucsf.edu.

This work was supported by grants from the California Breast Cancer Research Program (to P.J.K., P.W., and M.S.) and from the NIH (to P.J.K., M.S., and B.K.)

REFERENCES

- Katzenellenbogen BS 1996 Estrogen receptors: bioactivities and interactions with cell signaling pathways. *Biol Reprod* 54:287-293
- Kuiper GGJM, Enmark E, Peltö-Huikko M, Nilsson S, Gustafsson J 1996 Cloning of a novel estrogen receptor expressed in rat prostate and ovary. *Proc Natl Acad Sci USA* 93:5925-5930
- Katzenellenbogen BS, Korach KS 1997 Editorial: a new actor in the estrogen receptor drama—enter ER beta. *Endocrinology* 138:861-862
- Parker MG 1993 Steroid and related receptors. *Curr Opin Cell Biol* 5:499-504
- Tsai MJ, O'Malley BW 1994 Molecular mechanisms of action of steroid/thyroid receptor superfamily members. *Annu Rev Biochem* 63:451-486
- Pettersson K, Grandien K, Kuiper GG, Gustafsson JA 1997 Mouse estrogen receptor beta forms estrogen response element-binding heterodimers with estrogen receptor alpha. *Mol Endocrinol* 11:1486-1496
- Cowley SM, Hoare S, Mosselman S, Parker MG 1997 Estrogen receptors alpha and beta form heterodimers on DNA. *J Biol Chem* 272:19858-19862
- Lees JA, Fawell SE, Parker MG 1989 Identification of two transactivation domains in the mouse oestrogen receptor. *Nucleic Acids Res* 17:5477-5488
- Kumar V, Green S, Stack G, Berry M, Jin JR, Chambon P 1987 Functional domains of the human estrogen receptor. *Cell* 51:941-51
- Tora L, White J, Brou C, Tasset D, Webster N, Scheer E, Chambon P 1989 The human estrogen receptor has two independent nonacidic transcriptional activation functions. *Cell* 59:477-87
- Bocquel MT, Kumar V, Stricker C, Chambon P, Gronemeyer H 1989 The contribution of the N- and C-terminal regions of steroid receptors to activation of transcription is both receptor and cell-specific. *Nucleic Acids Res* 17:2581-95
- Berry M, Metzger D, Chambon P 1990 Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestrogen 4-hydroxytamoxifen. *EMBO J* 9:2811-8
- Danielian PS, White R, Lees JA, Parker MG 1992 Identification of a conserved region required for hormone dependent transcriptional activation by steroid hormone receptors. *EMBO J* 11:1025-1033
- Sadovsky Y, Webb P, Lopez G, Baxter JD, Fitzpatrick PM, Gizang GE, Cavailles V, Parker MG, Kushner PJ 1995 Transcriptional activators differ in their responses to overexpression of TATA-box-binding protein. *Mol Cell Biol* 15:1554-1563
- Le Douarin B, vom Baur E, Zechel C, Heery D, Heine M, Vivat V, Gronemeyer H, Losson R, Chambon P 1996 Ligand-dependent interaction of nuclear receptors with potential transcriptional intermediary factors (mediators). *Philos Trans R Soc Lond B Biol Sci* 351:569-578
- Horwitz KB, Jackson TA, Bain DL, Richer JK, Takimoto GS, Tung L 1996 Nuclear receptor coactivators and corepressors. *Mol Endocrinol* 10:1167-1177
- Glass CK, Rose DW, Rosenfeld MG 1997 Nuclear receptor coactivators. *Curr Opin Cell Biol* 9:222-232
- Shibata H, Spencer TE, Onate SA, Jenster G, Tsai SY, Tsai MJ, O'Malley BW 1997 Role of co-activators and co-repressors in the mechanism of steroid/thyroid receptor action. *Recent Prog Horm Res* 52:141-164
- Henttu PM, Kalkhoven E, Parker MG 1997 AF-2 activity and recruitment of steroid receptor coactivator 1 to the estrogen receptor depend on a lysine residue conserved in nuclear receptors. *Mol Cell Biol* 17:1832-1839
- Feng WJ, Ribeiro RCJ, Wagner RL, Nguyen H, Apriletti JW, Fletterick RJ, Baxter JD, Kushner PJ, West BL 1998 Hormone-dependent coactivator binding to a hydrophobic cleft on nuclear receptors. *Science* 280:1747-1749
- Hong H, Kohli K, Trivedi A, Johnson DL, Stallcup MR 1996 GRIP1, a novel mouse protein that serves as a transcriptional coactivator in yeast for the hormone binding domains of steroid receptors. *Proc Natl Acad Sci USA* 93:4948-52
- Hong H, Kohli K, Garabedian MJ, Stallcup MR 1997 GRIP1, a transcriptional coactivator for the AF-2 trans-

- activation domain of steroid, thyroid, retinoid, and vitamin D receptors. *Mol Cell Biol* 17:2735-2744
22. Voegel JJ, Heine MJS, Tini M, Vivat V, Chambon P, Gronemeyer H 1998 The coactivator TIF2 contains three nuclear receptor-binding motifs and mediates transactivation through CBP binding dependent and independent pathways. *EMBO J* 17:507-519
 23. Onate SA, Tsai SY, Tsai MJ, O'Malley BW 1995 Sequence and characterization of a coactivator for the steroid hormone receptor superfamily. *Science* 270:1354-1357
 24. Li H, Gomes PJ, Chen JD 1997 RAC3, a steroid/nuclear receptor-associated coactivator that is related to SRC-1 and TIF2. *Proc Natl Acad Sci USA* 94:8479-8484
 25. Torchia J, Rose DW, Inostroza J, Kamei Y, Westin S, Glass CK, Rosenfeld MG 1997 The transcriptional coactivator p/CIP binds CBP and mediates nuclear-receptor function. *Nature* 387:677-684
 26. Chen H, Lin RJ, Schiltz RL, Chakravarti D, Nash A, Nagy L, Privalsky ML, Nakatani Y, RME 1997 Nuclear receptor coactivator ACTR is a novel histone acetyltransferase and forms a multimeric activation complex with P/CAF and CBP/p300. *Cell* 90:569-580
 27. Anzick SL, Kononen J, Walker RL, Azorsa DO, Tanner MM, Guan XY, Sauter G, Kallioniemi OP, Trent JM, Meltzer PS 1997 AIB1, a steroid receptor coactivator amplified in breast and ovarian cancer. *Science* 277:965-968
 28. Heery DM, Kalkhoven E, Hoare S, Parker MG 1997 A signature motif in transcriptional co-activators mediates binding to nuclear receptors. *Nature* 387:733-736
 29. Ding XF, Anderson CM, Ma H, Hong H, Uht RM, Kushner PJ, Stallcup MR 1998 Nuclear receptor-binding sites of coactivators glucocorticoid receptor interacting protein1 (GRIP1) and steroid receptor coactivator 1 (SRC-1): multiple motifs with different binding specificities. *Mol Endocrinol* 12:302-313
 30. Kalkhoven E, Valentine JE, Heery DM, Parker MG 1998 Isoforms of steroid receptor co-activator 1 differ in their ability to potentiate transcription by the oestrogen receptor. *EMBO J* 17:232-243
 31. L'Horsset F, Dauvois S, Heery DM, Cavailles V, Parker MG 1996 RIP-140 interacts with multiple nuclear receptors by means of two distinct sites. *Mol Cell Biol* 16:6029-6036
 32. vom Baur E, Zechel C, Heery D, Heine MJ, Garnier JM, Vivat V, Le Douarin B, Gronemeyer H, Chambon P, Losson R 1996 Differential ligand-dependent interactions between the AF-2 activating domain of nuclear receptors and the putative transcriptional intermediary factors mSUG1 and TIF1. *EMBO J* 15:110-124
 33. Chakravarti D, LaMorte V, Nelson M, Nakajima T, Schulman I, Juguilon H, Montminy M, Evans R 1996 Role of CBP/P300 in nuclear receptor signalling. *Nature* 383:99-103
 34. Kamei Y, Xu L, Heinzel T, Torchia J, Kurokawa R, Glass B, Lin SC, Heyman RA, Rose DW, Glass CK, Rosenfeld MG 1996 A CBP integrator complex mediates transcriptional activation and AP-1 inhibition by nuclear receptors. *Cell* 85:403-414
 35. Yang XJ, Ogryzko VV, Nishikawa J, Howard BH, Nakatani Y 1996 A p300/CBP-associated factor that competes with the adenoviral oncoprotein E1A. *Nature* 382:319-324
 36. Korzus E, Torchia J, Rose DW, Xu L, Kurokawa R, McInerney EM, Mullen TM, Glass CK, Rosenfeld MG 1998 Transcription factor-specific requirements for coactivators and their acetyltransferase functions. *Science* 279:703-707
 37. Ogryzko VV, Schiltz RL, Russanova V, Howard BH, Nakatani Y 1996 The transcriptional coactivators p300 and CBP are histone acetyltransferases. *Cell* 87:953-959
 38. Bannister AJ, Kouzarides T 1996 The CBP co-activator is a histone acetyltransferase. *Nature* 384:641-643
 39. Spencer TE, Jenner G, Burcin MM, Allis CD, Zhou J, Mizzen CA, McKenna NJ, Onate SA, Tsai SY, Tsai MJ, O'Malley BW 1997 Steroid receptor coactivator-1 is a histone acetyltransferase. *Nature* 389:194-8
 40. Takeshita A, Yen PM, Misiti S, Cardona GR, Liu Y, Chin WW 1996 Molecular cloning and properties of a full-length putative thyroid hormone receptor coactivator. *Endocrinology* 137:3594-7
 41. Swope DL, Mueller CL, Chrivia JC 1996 CREB-binding protein activates transcription through multiple domains. *J Biol Chem* 271:28138-28145
 42. Imakado S, Koike S, Kondo S, Sakai M, Muramatsu M 1991 The N-terminal transactivation domain of rat estrogen receptor is localized in a hydrophobic domain of eighty amino acids. *J Biochem (Tokyo)* 109:684-689
 43. Metzger D, Ali S, Bornert JM, Chambon P 1995 Characterization of the amino-terminal transcriptional activation function of the human estrogen receptor in animal and yeast cells. *J Biol Chem* 270:9535-9542
 44. McInerney EM, Katzenellenbogen BS 1996 Different regions in activation function-1 of the human estrogen receptor required for antiestrogen- and estradiol-dependent transcription activation. *J Biol Chem* 271:24172-24178
 45. Ali S, Metzger D, Bornert JM, Chambon P 1993 Modulation of transcriptional activation by ligand-dependent phosphorylation of the human oestrogen receptor A/B region. *EMBO J* 12:1153-1160
 46. Le Goff P, Montano MM, Schodin DJ, Katzenellenbogen BS 1994 Phosphorylation of the human estrogen receptor. Identification of hormone-regulated sites and examination of their influence on transcriptional activity. *J Biol Chem* 269:4458-4466
 47. Kato S, Endoh H, Masuhiro Y, Kitamoto T, Uchiyama S, Sasaki H, Masushige S, Gotoh Y, Nishida E, Kawashima H, Metzger D, Chambon P 1995 Activation of the estrogen receptor through phosphorylation by mitogen-activated protein kinase. *Science* 270:1491-1494
 48. Bunone G, Briand PA, Miksicek RJ, Picard D 1996 Activation of the unliganded estrogen receptor by EGF involves the MAP kinase pathway and direct phosphorylation. *EMBO J* 15:2174-2183
 49. Trowbridge JM, Rogatsky I, Garabedian MJ 1997 Regulation of estrogen receptor transcriptional enhancement by the cyclin A/Cdk2 complex. *Proc Natl Acad Sci USA* 94:10132-10137
 50. Lahooti H, White R, Danielian PS, Parker MG 1994 Characterization of ligand-dependent phosphorylation of the estrogen receptor. *Mol Endocrinol* 8:182-188
 51. Tasset D, Tora L, Fromental C, Scheer E, Chambon P 1990 Distinct classes of transcriptional activating domains function by different mechanisms. *Cell* 62:1177-1187
 52. Kraus WL, McInerney EM, Katzenellenbogen BS 1995 Ligand-dependent, transcriptionally productive association of the amino- and carboxyl-terminal regions of a steroid hormone nuclear receptor. *Proc Natl Acad Sci USA* 92:12314-12318
 53. McInerney EM, Tsai MJ, O'Malley BW, Katzenellenbogen BS 1996 Analysis of estrogen receptor transcriptional enhancement by a nuclear hormone receptor coactivator. *Proc Natl Acad Sci USA* 93:10069-10073
 54. Gronemeyer H, Benhamou B, Berry M, Bocquel MT, Gofflo D, Garcia T, Lerouge T, Metzger D, Meyer ME, Tora L, Vergezac A, Chambon P 1992 Mechanisms of antihormone action. *J Steroid Biochem Mol Biol* 41:217-221
 55. Webb P, Lopez GN, Uht RM, Kushner PJ 1995 Tamoxifen activation of the estrogen receptor/AP-1 pathway: potential origin for the cell-specific estrogen-like effects of antiestrogens. *Mol Endocrinol* 9:443-456
 56. Aumais JP, Lee HS, Lin R, White JH 1997 Selective interaction of hsp90 with an estrogen receptor ligand-

- binding domain containing a point mutation. *J Biol Chem* 272:12229-12235
57. Tora L, Mullick A, Metzger D, Ponglikitmongkol M, Park I, Chambon P 1989 The cloned human oestrogen receptor contains a mutation which alters its hormone binding properties. *EMBO J* 8:1981-1986
58. Smith CL, Nawaz Z, O'Malley BW 1997 Coactivator and corepressor regulation of the agonist/antagonist activity of the mixed antiestrogen, 4-hydroxytamoxifen. *Mol Endocrinol* 11:657-66
59. Elliston JF, Tsai SY, O'Malley BW, Tsai MJ 1990 Superactive estrogen receptors. Potent activators of gene expression. *J Biol Chem* 265:11517-11521
60. Tremblay GB, Tremblay A, Copeland NG, Gilbert DJ, Jenkins NA, Labrie F, VG 1997 Cloning, chromosomal localization, and functional analysis of the murine estrogen receptor beta. *Mol Endocrinol* 11:353-365
61. Paech K, Webb P, Kuiper GGJM, Nilsson S, Gustafsson J, Kushner PJ, Scanlan TS 1997 Differential ligand activation of estrogen receptors ER alpha and ER beta at AP1 sites. *Science* 277:1508-1510
62. Lavinsky RM, Jepsen K, Heinzel T, Torchia J, Mullen TM, Schiff R, Del-Rio AL, Ricote M, Ngo S, Gemsch J, Hilsenbeck SG, Osborne CK, Glass CK, Rosenfeld MG, Rose DW 1998 Diverse signaling pathways modulate nuclear receptor recruitment of N-CoR and SMRT complexes. *Proc Natl Acad Sci USA* 95:2920-2925
63. Jackson TA, Richer JK, Bain DL, Takimoto GS, Tung L, Horwitz KB 1997 The partial agonist activity of antagonist-occupied steroid receptors is controlled by a novel hinge domain-binding coactivator L7/SPA and the corepressors N-CoR or SMRT. *Mol Endocrinol* 11:693-705
64. Wagner BL, Norris JD, Knotts TA, Weigel NL, McDonnell DP 1998 The nuclear corepressors NCoR and SMRT are key regulators of both ligand- and γ -bromo-cyclic AMP-dependent transcriptional activity of the human progesterone receptor. *Mol Cell Biol* 18:1369-1378
65. Zhang X, Jeyakumar M, Petukhov S, Bagchi MK 1998 A nuclear receptor corepressor modulates transcriptional activity of antagonist-occupied steroid hormone receptor. *Mol Endocrinol* 12:513-524
66. Korzus E, Torchia J, Rose DW, Xu L, Kurokawa R, McInerney EM, Mullen TM, Glass CK, Rosenfeld MG 1998 Transcription factor specific requirements for coactivators and their acetyltransferase functions. *Science* 279:703-707
67. Kurokawa R, Kalafus D, Ogliastro MH, Kioussi C, Xu L, Torchia J, Rosenfeld MG, Glass CK 1998 Differential use of CREB binding protein-coactivator complexes. *Science* 279:700-703
68. Saatcioglu F, Lopez G, West BL, Zandi E, Feng W, Lu H, Esmaili A, Apriletti JW, Kushner PJ, Baxter JD, Karin M 1997 Mutations in the conserved C-terminal sequence in thyroid hormone receptor dissociate hormone-dependent activation from interference with AP-1 activity. *Mol Cell Biol* 17:4687-4695
69. Norris JD, Fan DJ, Wagner BL, McDonnell DP 1996 Identification of the sequences within the human complement 3 promoter required for estrogen responsiveness provides insight into the mechanism of tamoxifen mixed agonist activity. *Mol Endocrinol* 10:1605-1616
70. Umesono K, Evans RM 1989 Determinants of target gene specificity for steroid/thyroid hormone receptors. *Cell* 57:1139-1146
71. Lopez G, Schaufele F, Webb P, Holloway JM, Baxter JD, Kushner PJ 1993 Positive and negative modulation of Jun action by thyroid hormone receptor at a unique AP1 site. *Mol Cell Biol* 13:3042-3049
72. Cavailles V, Dauvois S, L'Horsset F, Lopez G, Hoare S, Kushner PJ, Parker MG 1995 Nuclear factor RIP140 modulates transcriptional activation by the estrogen receptor. *EMBO J* 14:3741-3751

**International Society for Clinical Densitometry (ISCD)
5th Annual Meeting, New Orleans, Louisiana, USA
January 14-17, 1999**

For more information, contact the ISCD Headquarters Office, 1200 19th Street, NW, Suite 300, Washington, DC 20036-2422, USA. Telephone (202) 828-6056; Fax (202) 857-1102; E-mail iscd@dc.sba.com.

ISCD Certification Course, New Orleans, Louisiana, USA, January 12-13, 1999. For more information, contact ISCD Professional Certification and Site Accreditation, 1104 Main Street, Suite 600, Vancouver, WA 98660, USA. Telephone (360) 694-5010; Fax (360) 694-5396; E-mail certify@iscd.org.